

Australian Government

**Department of Health and Aged Care** Office of the Gene Technology Regulator

# **Risk Assessment and Risk Management Plan** for

# **DIR 208**

Clinical trial of a GM vaccinia virus for the treatment of solid tumours

Applicant – Novotech (Australia) Pty Limited

11 March 2025

### PAGE INTENTIONALLY LEFT BLANK

### Summary of the Risk Assessment and Risk Management Plan for Licence Application No. DIR 208

### Decision

The Gene Technology Regulator (the Regulator) has decided to issue a licence for this application for the intentional release of a genetically modified organism (GMO) into the environment. A Risk Assessment and Risk Management Plan (RARMP) for this application has been prepared by the Regulator in accordance with the *Gene Technology Act 2000* (the Act) and corresponding state and territory legislation, and finalised following consultation with a wide range of experts, agencies and authorities, and the public. The RARMP concluded that the proposed trial poses negligible risk to human health and safety and the environment and that any risks posed by the dealings can be managed by imposing conditions on the release.

The applicant, Novotech (Australia) Pty Limited (Novotech), proposes to conduct a clinical trial to evaluate the safety and efficacy of a genetically modified (GM) vaccinia virus (VACV), for the treatment of solid tumours.

The proposed GM VACV has been designed to preferentially replicate in and kill cancer cells. The GM VACV would be manufactured overseas and imported into Australia. It would be administered by intravenous infusion in up to 40 patients with solid tumours at clinical facilities and hospitals in Australia.

Clinical trials in Australia are conducted in accordance with requirements of the *Therapeutic Goods Act 1989*, which is administered by the Therapeutic Goods Administration (TGA). Therefore, in addition to approval by the Regulator, Novotech would require authorisation from the TGA before the trial commences. Clinical trials conducted in Australia must also be conducted in accordance with the *National Statement on Ethical Conduct in Human Research* and with the *Guidelines for Good Clinical Practice* of the International Council for Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use. Novotech would also require approval from the Department of Agriculture, Fisheries and Forestry (DAFF) for import of the GMO into Australia. In addition, they may require approval from the Chief Inspector of Stock before bringing the GMO into South Australia; an authorisation from the Department of Jobs, Skills, Industry and Regions - Agriculture Victoria in Victoria and a Prohibited Matter Permit from New South Wales, Queensland and Western Australia if they wish to conduct dealings in those states.

Risk Assessment and Risk Management Plan (RARMP) for this application has been prepared by the Regulator in accordance with the Act and corresponding state and territory legislation, and finalised following consultation with a wide range of experts, agencies and authorities, and the public. The RARMP concludes that the proposed clinical trial poses negligible risks to human health and safety and the environment

Project Title	Clinical trial of a GM vaccinia virus for the treatment of solid tumours	
Parent organism	Vaccinia virus (VACV)	
Principal purpose	The proposed trial is a Phase 1 study designed to evaluate the safety and efficacy of a genetically modified (GM) vaccinia virus, for the treatment of patients with solid tumours.	
Genetic modifications	<ul> <li>Introduced genes<sup>1</sup>:</li> <li>Three separate genes related to immune function of human origin, which enhance anti-tumour immune responses.</li> <li>Deleted genes<sup>1</sup>:</li> <li>The deletion of three VACV genes, which improves the efficacy and safety of the GMO.</li> </ul>	
Previous clinical trials	This is a first in human clinical trial using this GMO	
Limits and controls		
Duration	5 years	
Number of participants	Up to 40 clinical trial participants in Australia	
Locations	The proposed trial would be conducted at a number of hospitals and clinics across Australia. The exact clinical trial sites are yet to be identified	
Controls	<ul> <li>Transport and storage of the GMO according to the Regulator's <i>Guidelines</i> for the Transport, Storage and Disposal of GMOs</li> <li>Require staff handling the GMO to be trained and to use personal protective equipment</li> <li>Staff with immunosuppressive disorders are excluded from handling the GMO</li> <li>Disposal of waste that may contain GMO according to clinical site procedures appropriate for risk group 2 organisms</li> <li>Provide patients with detailed instructions regarding the care of any skin-related reactions post-treatment and the use of good hygiene practices</li> </ul>	

### The application

### Risk assessment

The risk assessment process considers how the genetic modifications and proposed activities conducted with the GMO might lead to harm to people or the environment. Risks are characterised in relation to both the seriousness and likelihood of harm, taking into account information in the application (including proposed controls), relevant previous approvals and current scientific/technical knowledge. Both short- and long-term impacts are considered.

Credible pathways to potential harm that were considered included the; potential exposure of people or animals to the GMO; and the potential for the GMO to transfer or acquire genetic material from other viruses. The potential for the GMO to be released into the environment and its effects were also considered.

<sup>&</sup>lt;sup>1</sup> Confidential Commercial Information (CCI): Some details about the inserted and deleted genes have been declared as CCI under section 185 of the Act. This information will be made available to the prescribed experts and agencies. CCI is not available to the public.

The risk assessment concludes that the trial poses negligible risks to human health and safety and to the environment. No specific risk treatment measures are required to manage these negligible risks. Important factors in reaching the conclusions of the risk assessment included that the GM VACV treatment is enriched for replication in cancer cells, and unintended exposure to the GMOs would be minimised by the limits and controls.

### **Risk management**

The risk management plan describes measures to protect the health and safety of people and to protect the environment by controlling or mitigating risk. The risk management plan is given effect through licence conditions.

As the level of risk is considered negligible, specific risk treatment is not required. However, since this is a clinical trial, the licence includes limits on the number of trial participants, types of facilities used, limits on the duration of the trial, as well as a range of controls to minimise the potential for the GMO to spread in the environment. In addition, there are several general conditions relating to ongoing licence holder suitability, auditing and monitoring, and reporting requirements which include an obligation to report any unintended effects.

### **Table of contents**

Summary o	f the Risk Assessment and Risk Management Plan (Consultation Version)I
Abbreviatio	nsVI
Chapter 1	Risk assessment context1
Section 1	Background1
1.1	Interface with other regulatory schemes2
Section 2	The proposed dealings3
2.1	The proposed limits of the trial (duration, scale, location, people)
2.2 envire	The proposed controls to restrict the spread and persistence of the GMOs in the onment
2.3	Details of the proposed dealings5
Section 3	Parent organism – Vaccinia virus8
3.1	Classification and genome characteristics8
3.2	Origin, geographic distribution and host range8
3.3	Infection cycle9
3.4	VACV persistence in infected hosts10
3.5	Pathology of VACV10
3.6	Transmission and shedding10
3.7	Recombination
3.8	Environmental stability and methods of decontamination for VACV11
3.9	VACV as a treatment
3.10	Adverse reactions to VACV infections12
3.11	Treatment of adverse reactions13
3.12	Risk group of VACV13
Section 4	The GMO – nature and effect of the genetic modification
4.1	Genetic modifications to VACV14
4.2	Shedding and safety of the GMO14
4.3	Stability in the environment and decontamination14
Section 5	The receiving environment15
5.1	Clinical trial sites15
5.2	Relevant environmental factors15
5.3	Related viral species in the receiving environment15
5.4	Presence of the introduced genes and encoded proteins in the environment
Section 6	Relevant Australian and international approvals16
6.1	Australian approvals16
6.2	International approvals16

Chapter 2	Risk assessment	17
Section 1	Introduction	17
Section 2	Risk identification	18
2.1	Risk source	18
2.2	Causal pathway	19
2.3	Potential harm	19
2.4	Postulated risk scenarios	20
Section 3	Uncertainty	27
Section 4	Risk evaluation	28
Chapter 3	Risk management plan	30
Section 1	Background	30
Section 2	Risk treatment measures for substantive risks	30
Section 3	General risk management	30
3.1	Limits and controls on the clinical trial	30
3.2	Other risk management considerations	33
3.2 Section 4	Other risk management considerations Issues to be addressed for future releases	33 34
3.2 Section 4 Section 5	Other risk management considerations Issues to be addressed for future releases Conclusions of the consultation RARMP	33 34 35

°C	Degrees Celsius
CCI	Confidential Commercial Information
CDC	Centers for Disease Control and Prevention
DAFF	Department of Agriculture, Fisheries and Forestry
DIR	Dealings involving Intentional Release
DNA	Deoxyribonucleic acid
GM	Genetically modified
GMO	Genetically modified organism
kb	kilobase
HREC	Human Research Ethics Committee
ΙΑΤΑ	International Air Transport Association
IBC	Institutional Biosafety Committee
ICH-GCP	<i>Guidelines for Good Clinical Practice</i> of the International Council for Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use
i.v.	Intravenous
NLRD	Notifiable Low Risk Dealings
NYCBH	New York City Board of Health
OGTR	Office of the Gene Technology Regulator
OPV	Orthopoxvirus
pfu	pock-forming units; plaque forming units
PPE	Personal protective equipment
RARMP	Risk Assessment and Risk Management Plan
Regulations	Gene Technology Regulations 2001
Regulator	Gene Technology Regulator
TGA	Therapeutic Goods Administration
the Act	Gene Technology Act 2000
TSD	Regulator's Guidelines for the Transport, Storage and Disposal of GMOs
USA	United States of America
VACV	Vaccinia virus
VARV	Variola virus
VIG	Vaccinia immunoglobulin
WHO	World Health Organisation

### Abbreviations

### Chapter 1 Risk assessment context

### Section 1 Background

1. An application has been made under the *Gene Technology Act 2000* (the Act) for Dealings involving the Intentional Release (DIR) of genetically modified organisms (GMOs) into the Australian environment.

2. The Act and the Gene Technology Regulations 2001 (the Regulations), together with corresponding State and Territory legislation, comprise Australia's national regulatory system for gene technology. Its objective is to protect the health and safety of people, and to protect the environment, by identifying risks posed by or as a result of gene technology, and by managing those risks through regulating certain dealings with GMOs.

3. Section 50 of the Act requires that the Gene Technology Regulator (the Regulator) must prepare a Risk Assessment and Risk Management Plan (RARMP) in response to an application for release of GMOs into the Australian environment. Sections 50, 50A and 51 of the Act and sections 9 and 10 of the Regulations outline the matters which the Regulator must take into account and who must be consulted when preparing the RARMP.

4. The *Risk Analysis Framework* (OGTR, 2013) explains the Regulator's approach to the preparation of RARMPs in accordance with the Act and the Regulations. The Regulator has also developed operational policies and guidelines that are relevant to DIR licences. These documents are available from the Office of the Gene Technology Regulator (<u>OGTR</u>) website.

5. Figure 1 shows the information that is considered, within the regulatory framework, in establishing the risk assessment context. This information is specific for each application. Potential risks to the health and safety of people or the environment posed by the proposed release are assessed within this context. Chapter 1 provides the specific information for establishing the risk assessment context for this application.

RISK ASSESSMENT CONTEXT
-------------------------

The GMO Modified genes Novel traits

**Parent organism (comparator)** Origin and taxonomy Cultivation and use Biology Proposed GMO dealings Activities Limits Controls

**Previous releases** Australian approvals International approvals

Receiving environment Environmental conditions: abiotic and biotic factors Production practices Related organisms Similar genes and proteins

Figure 1. Summary of parameters used to establish the risk assessment context, within the legislative requirements, operational policies and guidelines of the OGTR and the Risk Analysis Framework.

6. In accordance with Section 50A of the Act, this application is considered to be a limited and controlled release application, as the Regulator was satisfied that it meets the criteria prescribed by the Act. Therefore, the Regulator was not required to consult with prescribed experts, agencies and authorities before preparation of the RARMP.

7. Section 52 of the Act requires the Regulator to seek comment on the RARMP from agencies the Gene Technology Technical Advisory Committee (GTTAC), State and Territory Governments, Australian Government authorities or agencies prescribed in the Regulations, Australian local councils and the Minister for the Environment - and from the public. The advice from the prescribed experts, agencies and authorities and how it was taken into account is summarised in Appendix A. One public submission was received and its consideration is summarised in Appendix B.

### 1.1 Interface with other regulatory schemes

8. Gene technology legislation operates in conjunction with other regulatory schemes in Australia. The GMOs and any proposed dealings conducted under a licence issued by the Regulator may also be subject to regulation by other Australian government agencies that regulate GMOs or GM products, including Food Standards Australia New Zealand, the Australian Pesticides and Veterinary Medicines Authority, the Therapeutic Goods Administration (TGA), the Australian Industrial Chemicals Introduction Scheme and the Department of Agriculture, Fisheries and Forestry (DAFF).

9. The DAFF regulates products imported into Australia to protect Australia from biosecurity risks. Under the *Biosecurity Act 2015*, the importation of biological material such as live GM treatments requires a permit from the DAFF. In addition, they may require approval from the Chief Inspector of Stock before bringing the GMO into South Australia; an authorisation from the Department of Jobs, Skills, Industry and Regions - Agriculture Victoria in Victoria and a Prohibited Matter Permit from New South Wales, Queensland and Western Australia if they wish to conduct dealings in those states.

10. Medicines and other therapeutic goods for use in Australia are required to be assessed for quality, safety and efficacy under the *Therapeutic Goods Act 1989* and must be included in the Australian Register of Therapeutic Goods. The TGA is responsible for administering the provisions of this legislation. Clinical trials of therapeutic products that are experimental and under development, prior to a full evaluation and assessment, are also regulated by the TGA through the Clinical Trial Approval (CTA) scheme or the Clinical Trial Notification (CTN) scheme.

11. Approval by a Human Research Ethics Committee (HREC) is also a fundamental requirement of a clinical trial. HREC review is a part of the research governance process carried out by an institution that is responsible for the quality, safety and ethical acceptability of research carried out under their auspices. HRECs review research proposals involving human participants to ensure that they are ethically acceptable and meet relevant standards and guidelines. Elements of research to be considered include research merit and integrity, justice, beneficence, and participant consent.

12. The National Health and Medical Research Council (NHMRC) has issued the *National Statement on Ethical Conduct in Human Research, 2023* (National Health and Medical Research Council et al., 2023) which is the principal ethics guideline setting out the requirements for the ethical design, review and conduct of human research in Australia. The *Therapeutic Goods Act 1989* requires an HREC to review and monitor all clinical trials of unregistered therapeutic goods. The HREC must be registered with the NHMRC and constituted and operating in accordance with the National Statement.

13. In terms of risk to individuals participating in a clinical trial, the TGA (as the primary regulatory agency of investigational products), the trial sponsor, the investigators and the HREC responsible for each trial site all have roles in ensuring participant's safety under the *Therapeutic Goods Act 1989* and the requirements of the National Statement. However, where the trial involves a GMO, authorisation is also required under gene technology legislation. To avoid duplication of regulatory oversight, and as risks to trial participants are addressed through the above mechanisms, the Regulator's focus is on assessing risks posed to people other than those participating in the clinical trial, and to the environment. This includes risks to people preparing and administering the GMO, and risks associated with import, transport and disposal of the GMO.

14. The International Council for Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use – Guideline for Good Clinical Practice is an international ethical and scientific quality standard for designing, conducting, recording and reporting trials that involve the

participation of human subjects (ICH, 2016). The guideline was developed with consideration of the current good clinical practices of the European Union, Japan, and the United States of America, as well as those of Australia, Canada, the Nordic countries and the World Health Organization. The TGA has adopted the Integrated addendum to ICH E6(R1): Guideline for good clinical practice E6(R2) (Therapeutic Goods Administration), which provides overarching guidance for conducting clinical trials in Australia which fall under TGA regulation.

15. Some dealings with the GMO will be conducted at clinical trial sites, which are medical facilities including out-patient settings, hospitals and associated pharmacies. Analysis of biological samples collected from trial participants administered with the GMO may occur at clinical trial sites or at pathology laboratories.

16. The State and Territory governments regulate hospitals and other medical facilities in Australia. All public and private hospitals and day procedure services need to be accredited to the National Safety and Quality Health Service (<u>NSQHS</u>) Standards developed by the Australian Commission on Safety and Quality in Healthcare (the Commission) and endorsed by the State and Territory Health Ministers. The Commission coordinates accreditation processes via the Australian Health Service Safety and Quality Accreditation scheme. The NSQHS Standards provide a quality assurance mechanism that tests whether relevant systems are in place to ensure that the minimum standards of safety and quality are met. The safety aspects addressed by the NSQHS Standards include the safe use of sharps, disinfection, sterilisation and appropriate handling of potentially infectious substances. Additionally, the Commission has developed the National Model Clinical Guidance Framework, which is based on, and builds on NSQHS Standards to ensure that clinical governance systems are implemented effectively and to support better care for patients and consumers.

17. The National Pathology Accreditation Advisory Council (<u>NPAAC</u>) advises Commonwealth, State and Territory Health Ministers on matters relating to the accreditation of pathology laboratories. NPAAC plays a key role in ensuring the quality of Australian pathology services and is responsible for the development and maintenance of standards and guidelines for pathology practices. The standards include safety precautions to protect the safety of workers from exposure to infectious microorganisms in pathology laboratories. While compliance with NPAAC standards and guidelines is not mandatory, there is a strong motivation for pathology services to comply, as Medicare benefits are only payable for pathology services if conducted in an appropriate Accredited Pathology Laboratory category, by an Approved Pathology Practitioner employed by an Approved Pathology Authority. Accreditation of pathology services is overseen by Services Australia (formerly Department of Human Services), and currently, the only endorsed assessing body for pathology accreditation is the National Association of Testing Authorities.

18. Hospitals and pathology laboratories, including their workers, managers and executives, all have a role in making the workplace safe and managing the risks associated with handling potentially infectious substances including the proposed GMO. There are minimum infection prevention practices that apply to all health care in any setting where health care is provided. These prevention practices were initially developed by the Centers for Disease Control and Prevention, and are known as the standard precautions for working with potentially infectious material. The standard precautions are described in the <u>Australian Guidelines for the Prevention and Control of Infection in Healthcare (2024)</u>.

### Section 2 The proposed dealings

19. Novotech has proposed a Phase 1 clinical trial of a live GM VACV that preferentially replicates in cancer cells. The purpose of the clinical trial is to assess the safety and efficacy of the GM treatment in patients with solid tumours.

- 20. The dealings involved in the proposed clinical trials are:
  - (a) import the GMO;
  - (b) conduct the following experiments with the GMO:

- i. prepare the GMO for administration to trial participants;
- ii. administer the GMO to clinical trial participants by intravenous (i.v.) infusion;
- iii. collect samples from trial participants;
- iv. analyse the samples;
- (c) transport the GMO; and
- (d) dispose of the GMO;

and the possession (including storage), supply and use the GMO for the purposes of, or in the course of, any of these dealings.

### 2.1 The proposed limits of the trial (duration, scale, location, people)

21. The clinical trial is proposed to take place over a five-year period from the date of issue of the licence. Up to 40 patients in Australia would receive a single dose of the GMO via i.v. infusion.

22. The trial would take place at hospitals and clinical sites in Australia, these sites have not yet been identified.

## **2.2** The proposed controls to restrict the spread and persistence of the GMOs in the environment

23. The applicant has proposed a number of controls to minimise exposure to the GMO, and to restrict the spread and persistence of the GMOs in the environment. These include:

- Only trained personnel would conduct dealings with the GMO. Staff preparing and administering the GMO would also be experienced in the use and disposal of sharps.
- Staff considered to be at high-risk (see paragraph 33) would be excluded from handling the GMO.
- Staff preparing/administering the GMO would be required to wear appropriate PPE (e.g. gown, gloves, and eye protection) during the procedures, and instructed on how to remove and/or dispose of PPE to avoid contamination and wash the hands after PPE removal.
- Transport, storage and disposal of the GMO and any contaminated waste generated at a clinical trial site must be in accordance with the current version of the Regulator's *Guidelines* for the Transport, Storage and Disposal of GMOs.
- Disinfecting surfaces and equipment that come into contact with the GMO using an effective disinfectant (including but not limited to 70% ethanol, 50% isopropyl alcohol, and/or 10% bleach).

24. The applicant has proposed several measures to limit the GMO exposure of close contacts and animals to the GMO. These include:

- The participants would be instructed to clean any household areas that may have been exposed to the GMO, including contaminated clothing, and bedding with a 10% bleach solution.
- Ensuring the administration site is kept clean and dry to prevent infection. Use of sterile, occlusive dressings to cover any pustules that develop. Changing dressings regularly and disposing of used dressings in biohazard waste bags.
- Trial participants will be instructed to avoid touching or scratching the infusion site to prevent autoinoculation and spread of the virus. Pustule kits and biohazard supplies will be provided to clinical trial participants in case of development of any pustules during the trial period.
- Instructing clinical trial participants to monitor themselves for any signs of infection or adverse reactions, such as fever, increased redness, or pus at the infusion site. Report any concerning symptoms or adverse events to a healthcare provider and the study investigator immediately.

• Participants will be instructed to avoid contact with animals and high-risk groups such as pregnant women, infants, immunocompromised individuals, and those with eczema or atopic dermatitis.

### 2.3 Details of the proposed dealings

### 2.3.1 Manufacture of the GMO

25. The GM VACV would be manufactured overseas in accordance with applicable Good Manufacturing Practice (GMP) regulations and imported into Australia (see Section 2.3.5).

### **Clinical trial sites**

26. The clinical trial would be carried out at clinical trial sites and hospitals, which are yet to be confirmed. Clinical trial sites would be assessed by the applicant for their ability to adhere to infection prevention practices outlined in the *Australian Guidelines for the Prevention and Control of Infection in Healthcare* (2019) (National Health and Medical Research Council, 2019). Sites would also be selected on an ability to comply with the International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use (ICH) Good Clinical Practice (GCP) guidelines (<u>ICH Guideline for Good Clinical Practice</u>).

### 2.3.2 The clinical trial

27. The international sponsor for the trial is ViroMissile Inc which is headquartered in California (CA), United States of America (USA). Novotech, as a clinical research organisation (CRO), is responsible for conducting the proposed clinical trial in Australia. As the licence holder, Novotech is responsible for ensuring that the licence conditions are met.

28. The GM VACV would be extracted from the vial using a needle attached to a syringe and diluted in a sterile buffer solution. Once at the desired dilution, the GMO would be transferred using needle and syringe to an infusion bag. The GMO would be administered via i.v. infusion over 60 minutes and the infusion line would then be flushed with sterile saline.

29. The infusion site would be covered with an occlusive dressing until complete wound healing. The dressing should be changed daily. Contaminated dressings would be placed in a biohazard waste container (provided to the patient) and returned to the clinical trial site at the next scheduled site visit.

### 2.3.3 Dose levels

30. This is a Phase 1, first-in-human (FIH), open-label, dose-escalation and single arm study to determine the Maximum-Tolerated Dose (MTD) or Maximum Feasible Dose (MFD) and assess the Dose Limiting Toxicities (DLT) of the GMO. The safety and tolerability of the investigational product will be assessed in adult participants with advanced or refractory solid tumours. The total number of participants in the study will depend on the number of dose-escalation cohorts with a maximum of 40 trial participants enrolled in Australia. The applicant has stated that they may amend the protocol to include combination cohorts and expansion cohorts which may also include additional treatments. The treatment arm will follow an escalation scheme. Participants will be treated at increasing doses of study drug until all dose levels have been investigated or any dose level is found to exceed the MTD. If the DLT or the MTD are not reached, the MFD will be defined as the highest dose level.

### 2.3.4 Selection of trial participants and behavioural requirements

31. Relevant inclusion criteria to this assessment proposed by the applicant include that trial participants <u>must</u>:

- be ≥18 years-of-age at the time of signature of the informed consent form (ICF).
- women of childbearing potential (WOCBP) must have a negative serum pregnancy test prior to study entry.

- refrain from gamete donation from the first dose of the GMO, throughout the study, and for 3 months from the last dose of the GMO.
- male or female participants: Male participants with female partners of childbearing potential and female participants of childbearing potential are required to use two forms of acceptable contraception, including one barrier method, during their participation in the study and for 3 months following the last dose of the GMO.
- be willing and able to comply with all study procedures, requirements, and follow-up examinations.
- 32. Relevant exclusion criteria include participants who:
  - are experiencing any active infections (bacterial, viral, or fungal) for which systemic antimicrobials are required.
  - have known HIV infection history.
  - have clinically significant immunodeficiency (e.g., due to underlying illness and/or medication) in themselves or their household contacts.
  - have open wounds with current or history of severe skin disease.
  - are taking certain medications (details of which have been declared Confidential Commercial Information (CCI). Under Section 185 of the Act. This information is available to the prescribed experts and agencies that will be consulted on this application upon request in the course of them performing duties or functions under the Act or under a corresponding State law. CCI is not available to the public.

33. For the purposes of this RARMP, persons who are pregnant or have immunosuppressive disorders are considered persons at a higher-risk of a serious adverse event when exposed to the GM VACV.

### 2.3.5 Transport and storage of the GMO

34. The GMO would be imported according to the packaging and labelling requirements of the International Air Transport Association (IATA) code UN 3373.

35. Transport of the GMO from the Australian border would be directly to the clinical sites or storage facility. Once at a clinical site or storage facility, the GMO would be stored in a freezer, with access restricted to appropriately trained personnel. The GMO will be contained within a sealed, unbreakable primary container and also be contained within a sealed unbreakable secondary container. The external surface of the primary and secondary container will be decontaminated before and after transport.

36. Procedures will be in place to ensure that all transported GMOs can be accounted for, and that a loss of GMOs during transport can be detected; and access to the GMOs will be restricted to authorised persons conducting dealings under the licence, who have been informed by the licence holder of any licence conditions that apply to them. This includes situations where containers are left for collection in a holding area.

37. The proposed method of supply and storage of the GMOs, as advised by the applicant, would be in accordance with the Regulator's <u>*Guidelines for the Transport, Storage and Disposal of GMOs*</u> (TSD).

### 2.3.6 Sample collection and processing

38. Biological samples such as blood, urine and anal swabs will be collected for efficacy and shedding analysis on days 1,2, 3, 4, 14 and 28.

39. Blood samples will be collected by clinical site staff wearing appropriate PPE. Staff must ensure that the sample collection area is clean and sterile.

40. Urine and anal swab samples will be self-collected by trial participants within clinical settings during follow-up visits. Appropriate instruction and training will be given to trial participants before the sample collection.

41. After sample collection, clinical site staff may need to process the samples according to the study protocol. This may involve centrifugation to separate blood components, aliquoting of samples into smaller volumes for storage or analysis, or immediate processing of tissue samples for analysis. All sample processing steps will be performed following appropriate safety precautions and in compliance with Good Clinical Laboratory Practices (GCLP) or other relevant guidelines.

42. Whilst some samples such as whole blood will be analysed at site of collection, most will be shipped to a central laboratory.

### 2.3.7 Personal protective equipment and exclusion criteria

43. The applicant advised that persons handling the GMO, including preparation and administration of the GMO to trial participants and clean-up of potential spills, would be instructed to wear appropriate PPE, including gown, gloves, mask and eye protection.

44. The applicant has proposed that those who are pregnant or immunosuppressed would be excluded from handling the GMO.

### 2.3.8 Decontamination and disposal of the GMOs (including waste contaminated with the GMOs)

45. The applicant states that all GMO-contaminated waste will be handled as per the clinical waste streams of the hospital or clinic being used. This includes specific and designated containers for the disposal of biohazardous materials, including used dressings. Biohazard bags or containers will be appropriately labelled, and securely sealed to prevent leakage or accidental exposure to the virus. The bag or container would then be disposed of according to institutional or regulatory guidelines for biohazardous waste disposal, which may involve incineration or other approved methods for destruction. GMO-contaminated waste generated outside of the clinical setting, such as bandages, dressings and other materials used to care for vaccinia related lesions, would also be disposed of in biohazard bins provided to patients. The biohazard bins would be returned to the clinical trial site for disposal. In all cases, the applicant commits to follow the Regulator's *Guidelines for the Transport, Storage and Disposal of GMOs*.

46. Unused GMO(s) at clinical trial sites, storage/distribution facilities and/or analytical facilities will be destroyed onsite, transported for disposal by external service providers or returned to ViroMissile. Decontamination at the trial site would be either by chemical treatment (e.g., using chemical disinfectants to treat work areas and reusable equipment) or by autoclaving. GMO spills would be decontaminated using a fresh dilution of 10% bleach, with at least 10 minutes of contact time.

### 2.3.9 Training of clinical trial personnel

47. Novotech has the responsibility to ensure the training of personnel and compliance with OGTR licence conditions.

48. The applicant has advised that appropriate training materials (e.g. training in all procedures specific to the GMO including preparing, handling, administration, spill procedures, containment and disposal, etc.) will be provided to all the personnel involved in the study.

49. The study drug infusion will be prepared by trained pharmacists or pharmacy technicians in a Class II Biosafety cabinet (BSC-2). Those staff would be trained on the preparation of the GMO and handling of sharps to minimise the likelihood of exposure. Additionally, at least two trained clinical trial staff need be present when the GMO administration is performed.

### 2.3.10 Contingency plans

50. In case of exposure of people to the GMO via sharps injury or contact with broken skin, the applicant proposes that persons who have had accidental direct contact with the GMO would be instructed to:

• bleed from the affected site; and

• scrub contaminated skin for several minutes with a 10% povidone solution (Betadine) and copious amounts of water.

51. In case of exposure of people to the GMO via aerosols, airborne droplets or direct contact with facial mucosa, the persons who have had accidental direct contact with the GMO will be instructed to rinse the affected area for a minimum of 15 minutes in eye wash or flushed with water.

52. The exposed person(s) will be monitored closely. The incident would be reported to the Principal Investigator, who would report to Novotech as soon as is practical. Novotech would then notify the Regulator.

53. In case of unintentional release of the GMO due to an accidental spill, the spill would be reported to Novotech by clinical trial staff trained in the OGTR reporting requirements. Novotech would on-report to the OGTR. The local Institutional Biosafety Committee (IBC) would also be notified of loss of containment or suspected loss of containment.

### 2.3.11 Accountability and monitoring

54. The applicant has proposed to instruct clinical trial participants to monitor themselves for any signs of infection or adverse reactions, such as fever, increased redness, or pus at the infusion site and report any concerning symptoms or adverse events to a healthcare provider and the study investigator immediately.

55. Any unintended exposure to the GMO through injury or direct contact would be reported to the OGTR.

### Section 3 Parent organism – Vaccinia virus

56. The parent organism is *Vaccinia* virus (VACV). The specific strain of VACV being used by the applicant has been declared as CCI. The characteristics of the non-GM parent organism provide a baseline for comparing the potential for harm from dealings with GMOs. As such, the relevant biological properties of VACV will be discussed here.

57. VACV was used globally as a vaccine against smallpox prior to the latter's declared eradication in 1980. It was a highly effective vaccine because it is a mild pathogen that stimulates an immune response to the closely related and often lethal smallpox agent *Variola virus* (Middaugh et al., 2016). The biology of VACV has been described in detail in the RARMPs for <u>DIR 116</u> and <u>DIR 140</u> (clinical trials with GM VACV) and more recently in RARMPs for <u>DIR 170</u> (trial with GM VACV in horses) and <u>DIR-179</u> (trial with GM VACV for treatment of tumours). A summary is presented in this section.

### 3.1 Classification and genome characteristics

58. VACV is an enveloped virus belonging to the *Orthopoxvirus* (OPV) genus, subfamily *Chlordopxvirinae*, family *Poxviridae*. This genus also includes the human pathogen *Variola virus* (causative agent of smallpox), cowpox virus, horsepox virus, mpox virus, mousepox virus and others (McLysaght et al.; Tulman et al., 2006). *Vaccinia* virus has a roughly 195 kilobases (kb) long double stranded DNA genome which is extremely AT rich (~66%) (Li et al., 2006) and replicates in membrane bound segments localised entirely in the cytoplasm. These segments encode for the production of close to 200 proteins which facilitate viral entry, transcription of viral genes, DNA synthesis, assembly of virus particles, and suppression of the host anti-viral response (Liu et al., 2014).

### 3.2 Origin, geographic distribution and host range

59. Originally thought to be cowpox virus, the primary agent of smallpox vaccines was discovered to be VACV in 1939 (Downie, 1939). VACV is more closely related to horsepox and likely arose from the recombination and selection of several related *Orthopox* viruses which had been randomly combined and sampled in the pursuit of potent vaccines against smallpox (Esparza et al., 2017; Qin et al., 2015).

60. Due to their evolution in different parts of the world over nearly two centuries of smallpox vaccination, many strains of VACV exist (e.g. Paris, Copenhagen, Bern, Ankara, Lister and New York City Board of Health (NYCBH) strains). These differ in viral characteristics, host range, pathogenicity and prevalence of adverse reactions following vaccination (see Paragraph 85). Specific information about the historic use of VACV based vaccines in Australia, as well as more modern examples of use in the U.S military are discussed in the RARMP for <u>DIR-179</u>.

61. The host specificity of pox viruses varies widely, some such as VACV, can infect multiple hosts, whilst others such as Variola virus (VARV) are reliant on one (Oliveira et al., 2017a; Qin et al., 2015). The natural host of VACV is not known, but in the environment and in laboratories, VACV is able to infect and cause disease in humans, several monkey species, a variety of rodents and marsupials, buffalo, dairy cattle, sheep, horses, domestic cats and dogs (Abrahão et al., 2010; Adams et al., 2007; Artois et al., 1990; Bennett et al., 1989; Brochier et al., 1989; Dumbell and Richardson, 1993; Felipetto Cargnelutti et al., 2012; Miranda et al., 2017; Oliveira et al., 2015; Riyesh et al., 2014; Robinson and Mercer, 1988). Birds are not known to be a host for VACV, but a study of a GM VACV-based rabies vaccine demonstrated sufficient viral replication in several Canadian bird species to permit seroconversion (Artois et al., 1990).

62. Naturally occurring infections with VACV or close relatives have been documented in South America, India, Indonesia, Egypt and other countries. In Brazil, outbreaks of zoonotic disease (transferable between animals and people) caused by VACV-like viruses and affecting dairy cattle and rural workers as well as significant levels of VACV infection found in remote Amazonian wildlife have also been reported (Abrahão et al., 2010; José da Silva Domingos et al., 2021; Miranda et al., 2017; Oliveira et al., 2014).

63. There are few poxviruses in circulation in Australia, of which, mpox and *Molluscum contagiosum* are likely the only candidates with community transmission in humans since it has been several decades since the discontinuation of the smallpox vaccination campaign (Hammerschlag et al., 2022; Konya and Thompson, 1999). Mpox was first identified in Australia in 2022 and has been circulating within the community since then (Healthdirect Australia, 2024). *Molluscum contagiosum* is more prevalent but is a mild condition which predominantly affects young children or those who are immuno-compromised (Healthdirect Australia 2024).

### 3.3 Infection cycle

64. VACV infection is usually transmitted through physical contact with an infected lesion or pustule (Lane and Fulginiti, 2003; Wertheimer et al., 2012). Once inside the host, infectious VACV particles fuse to host cells through a process which involves at least 16 different proteins (Laliberte et al., 2011). This level of complexity is an oddity among enveloped viruses, with most requiring only one or two proteins for entry. In addition to cell fusion, VACV can also enter host cells through macropinocytosis, a form of endocytosis, or via uptake from an intracellular vesicle (Mercer et al., 2010).

65. Once inside the cell, VACV disassembles, and its core navigates the microtubule network to deposit itself in a perinuclear position within the cytoplasm (Schmidt et al., 2013). Its entire replication cycle occurs in the cytoplasm, rendering the virus incapable of integrating into the host genome (Liu et al., 2014). As an additional consequence of being spatially restricted from the nucleus, VACVs are unable to use host replication enzymes and their genomes encode enzymes for DNA replication and gene transcription (Greseth and Traktman, 2022; Schramm and Locker, 2005).

66. VACV genes are expressed in three phases, early, intermediate and late (Yang et al., 2011). The mRNAs for the early phase genes are largely required for DNA replication and are expressed shortly after the virus enters the cell (Baldick and Moss, 1993). Expression of intermediate and late genes occurs post-DNA replication and needs *de novo* RNA and protein synthesis (Shors et al., 1999). These genes spur a complicated assembly and maturation process. This involves fabrication of membranes with material sourced from the host ER (Maruri-Avidal et al., 2013). These membrane bound particles will then expand, incorporating a number of viral proteins and becoming a viral replication factory

(Greseth and Traktman, 2022). Once the genome has been replicated, the virions can be transported to the host cell membrane, where fusion will allow them to be released (Payne and Kristenson, 1979) (Horsington et al., 2013).

### 3.4 VACV persistence in infected hosts

67. There currently exists no evidence that VACV is able to persist in a latent state within an infected host. Its large genome and particle size enables efficient detection and clearance by phagocytic cells of the immune system (Buller and Palumbo, 1991). Further evidence of this is that, despite several viral variants being passaged for decades as part of global vaccination efforts, no immune evading variants of VACV have arisen.

### 3.5 Pathology of VACV

68. The majority of VACV variants and strains are mild pathogens in people and animals. As described in paragraph 64, the receptor binding proteins harboured by VACV are numerous. It is perhaps because of this that no single cell surface molecule has been conclusively demonstrated to be necessary for VACV virus infection. The range of cell types that VACV can infect in culture is vast but despite this, in humans, VACV primarily target antigen presenting cells and activated but not resting T cells (Chahroudi et al., 2005).

69. When contracted zoonotically, such as in the case of bovine vaccinia (BV) from milking cows or buffalos, VACV manifests as ulcerative lesions at the primary site of infection (Medeiros-Silva et al., 2010). Secondary lesions following autoinoculation have also been observed (Oliveira et al., 2017c; Tack et al., 2013). These lesions or pustules are usually accompanied by one or more of symptoms such as fever, malaise, headache, nausea and muscle aches (Medeiros-Silva et al., 2010). The lesions typically take 21 days to fully heal whilst the other general symptoms usually resolve in 3 days (José da Silva Domingos et al., 2021). When administered as a vaccine, VACV infection normally induces a single lesion at the site of exposure around 3-4 days post vaccination, which generally resolves over 2-3 weeks (Fulginiti et al., 2003a). This is often accompanied by flu-like symptoms, as described above, and swelling and tenderness of the draining lymph node (Cono et al., 2003; Public Health Agency of Canada, 2011). In healthy people, these reactions resolve spontaneously and require only observation and symptomatic treatment (Cono et al., 2003; Fulginiti et al., 2003).

70. Serious adverse reactions associated with VACV such as *post-vaccinia encephalitis* (PVE) or death are rare, strain dependent, and particularly affect those with underlying risk factors such as atopic dermatitis, or those who are immunocompromised, as in the case of AIDS (Cono et al., 2003). More information regarding severe adverse reactions specifically relating to VACV as a vaccine can be found in Section 3.10 of this Chapter.

### 3.6 Transmission and shedding

71. The RARMPs for <u>DIR 140</u>, <u>DIR 170</u> and <u>DIR-179</u> describe transmission (both between humans, as well as to and between animals) and shedding of VACV in detail. Importantly, transmission can occur via direct physical contact with lesions or the vaccine inoculation site or contact with a contaminated objects (e.g. bandages, clothing, sheets and towels). An infected person may also spread VACV from the initial infection site by touching other body parts or people with contaminated hands, or through every day activities such as shaving (Cono et al., 2003; Oliveira et al., 2014; Tack et al., 2013; Webber et al., 2014). Oral transmission via drinking contaminated cow milk has been observed in humans (Damaso et al., 2000). Aerosol transmission of VACV has never been clearly documented in people when used as a vaccine (Lane and Fulginiti, 2003) and is considered unlikely.

72. In the context of smallpox vaccination, VACV can be shed from the primary lesion shortly post injection and can continue for up to at least 42 days in some vaccine recipients (Pittman et al., 2015). Shedding appears to peak at between 10 and 16 days post injection and drops off dramatically in about 90% of vaccinees by day 28 (Cooney et al., 1991; Cummings et al., 2008; Wharton et al., 2003).

VACV shedding has also been well demonstrated in non-human hosts, such as in the milk and faeces of experimentally infected dairy cattle, and the urine and faeces of mice (Ferreira et al., 2008; Matos et al., 2018).

73. The minimum infectious dose of VACV is unknown. In the context of vaccination, the dose required to illicit a robust immune response to VACV varies by strain but can be achieved with as little as  $2.5 \times 10^5$  pfu (Jacobs et al., 2009; Miller et al., 2008). For those that do receive an effective dose of vaccine, their propensity to transmit active virus to close contacts is uncertain but presumed to be relatively low (Lane and Fulginiti, 2003; Neff et al., 2002; Sepkowitz, 2003; Wertheimer et al., 2012). Over 2 million vaccinations were delivered in the U.S between 2002 and 2011, and of these, only 115 cases of secondary transmission were documented (Wertheimer et al., 2012). This rate of ~5 secondary transmissions per 100,000 vaccinations almost entirely represented close physical or intimate contacts (90%) and has been corroborated by smaller, more well controlled studies (Tack et al., 2013). It is also noteworthy that the vast majority of these infections were mild in presentation and only one life threatening event was documented. In hospital settings which contain high proportions of patients with compromised immune function and other vulnerabilities, fomite-based transmission has been observed, but not well quantified empirically (Sepkowitz, 2003).

74. In humans, viremia (viral presence in the blood) and viruria (viral presence in urine) is uncommon, although does occur in patients with progressive vaccinia and eczema vaccinatum (Lane and Fulginiti, 2003).

### 3.7 Recombination

75. Recombinants between Orthopoxviruses (OPVs) in cell cultures in a laboratory setting are easily produced and have been described in the RARMP for <u>DIR 170</u>.

76. Although replicating poxviruses can recombine very efficiently under certain circumstances, there are physical constraints within a cell that limit recombination between co-infecting viruses. VAVC transcription, translation and replication takes place in the cytoplasm but within membrane-bound cytoplasmic structures known as viral factories or virosomes (Katsafanas and Moss, 2007; Lin and Evans, 2010; Paszkowski et al., 2016), thus, compartmentalising and preventing the mixing of their nucleic acid from other viruses in the same cell (Paszkowski et al., 2016). For recombination to occur therefore, two viral factories of separate viruses in a super-infected cell would have to fuse during DNA replication.

77. For these reasons, it had long been thought that poxviruses seldom recombine naturally. However, genomic evidence suggests that recombination and horizontal gene transfer are major drivers of poxvirus evolution (Bratke and McLysaght, 2008; Brennan et al., 2023; Sprygin et al., 2022). In some instances, horizontal gene transfers appear to have conferred pox viruses with survival advantages (Bratke and McLysaght, 2008). Several poxvirus genes are thought to have been acquired and repurposed for functions such as immune evasion (Vallée et al., 2021).

78. In the case of the recent outbreak of mpox, it was demonstrated that recombination between viral isolates occurred, although the impact this had on driving the spread of the virus is unknown (Yeh et al., 2022).

79. Despite the genomic and laboratory evidence of their occurrence and possibility, concrete examples of natural poxvirus recombination are rare. On the whole, there still exists great uncertainty as to the frequency, mechanisms, and significance of gene flow and recombination events within poxviruses (Sprygin et al., 2022).

### 3.8 Environmental stability and methods of decontamination for VACV

80. Poxviruses are well known for their ability to persist in the environment, and they are more resistant to drying and increased temperature than other enveloped viruses. VACV stability is

determined by temperature, relative humidity and the materials on which VACV is introduced into the environment (fomites) (Wood et al., 2013).

81. VACV survival decreases at high temperatures or high humidity and is greater at lower temperatures. Dried VACV can be kept for more than 35 weeks at 4°C with no loss of infectivity (Rheinbaben et al., 2007). When frozen (-20°C), 1 in 1000 virus particles remained viable after 15 years (Essbauer et al., 2007; Rheinbaben et al., 2007). Samples (15 mL) of VACV at 10<sup>7.5</sup> TCID<sub>50</sub>/mL (median Tissue Culture Infectious Dose 50%; equivalent to approximately 0.5 x 10<sup>8</sup> pfu) can also remain viable for more than two weeks on food samples in the fridge (4°C) or close to 6 months in storm water at 4.5°C. However, the presence of soil in stormwater decreased survival time of the VACV sample to 6 days at 4.5°C or 3 days at 21.5°C (Essbauer et al., 2007). Murine faeces exposed to environmental conditions retained infectious VACV particles for at least 20 days (Abrahão et al., 2009).

82. Purified samples of VACV are inactivated within 1 minute by a range of common chemical disinfectants including 0.5% sodium hypochlorite and 40% ethanol (Chambers et al., 2009).

83. VACV is susceptible to UV irradiation, but a small proportion (up to 10%) of virions appear to remain active when dried onto UV exposed surfaces (Sagripanti and Lytle, 2011). VACV is inactivated by dry heat at 95°C for 2 hours (Sauerbrei and Wutzler, 2009) and by autoclaving (Espy et al., 2002). Appropriate hand hygiene after contact with items that may be contaminated with VACV includes washing with antimicrobial soap and water or alcohol-based hand-rub containing 60% ethanol or more (Wharton et al., 2003).

### 3.9 VACV as a treatment

84. Currently, VACV is considered well-suited as a viral vector to create a new generation of safer GM vaccines and cancer treatments, such as the ones proposed within this application (Chaurasiya et al., 2020; Dyer et al., 2019; Guo et al., 2019; Harrington et al., 2019; Nagata et al., 2018). Some of the features of VACV viral vectors that make them suitable for GM cancer treatment applications are:

- thermostability, which allows for a cold-chain independent distribution capacity
- large DNA genome capable of accepting inserts of up to 25 kb
- ability to grow to high titres in vitro
- ability to elicit strong humoral and cell-mediated immune responses that enhance the immune response to the target antigens
- absence of oncogenic potential or evidence of integration into the host genome, and
- wide host range.

### 3.10 Adverse reactions to VACV infections

85. Although smallpox vaccination using VACV during the eradication campaign was generally safe and effective, serious adverse reactions have been well documented. Most complications occurred as a result of vaccination, but serious sometimes fatal, reactions occurred following transmission to unvaccinated individuals. Several types of adverse events have occurred in healthy people, while other events have been associated with specific risk factors or underlying conditions (Cono et al., 2003; Fulginiti et al., 2003b; Lane and Goldstein, 2003a; Lane and Goldstein, 2003b; Maurer et al., 2003; Neff et al., 2002; Wittek, 2006).

86. As previously mentioned, secondary transmissions are a rare occurrence. This may be in part because accidental transmission would likely generate a very limited or low infectious dose. This in turn would reduce the rate of viral progression and the proportion of cells which produce infectious virions (Howell et al., 2024).

87. Various adverse reactions may result from VACV infection including Generalised vaccinia (GV), post-vaccinal encephalopathy (PVE), progressive vaccinia (PV), foetal vaccinia (FV), and eczema vaccinatum (EV). The most severe of these (GV, PV and PVE) are rare but can result in high levels of morbidity and even death (Kretzschmar et al., 2006; Maurer et al., 2003). FV is a rare complication,

with only 50 cases reported in the literature (Cono et al., 2003). It results from maternal exposure to VACV during pregnancy or shortly before conception and often leads to stillbirth or neonatal death. Due to its rarity, specific risk factors have not been determined. No other specific risks to foetuses or pregnant women have been identified. EV events are slightly more common, especially in those with a pre-existing history of eczema, but are unlikely to be fatal (Cono et al., 2003; Fulginiti et al., 2003b). Details of these adverse reactions are described in detail in <u>DIR-179</u>.

### 3.11 Treatment of adverse reactions

88. Vaccinia immunoglobulin (VIG) is made from the plasma of recently vaccinated people and has been successfully used to treat certain complications of VACV infection. It is recommended for treating severe cases of accidental implantation, severe generalised vaccinia, eczema vaccinatum and severe progressive vaccinia. It is not recommended for mild instances of accidental implantation, mild or limited generalised vaccinia, and post-vaccinial CNS disease. VIG is contraindicated in patients with vaccinia keratitis (Centers for Disease Control and Prevention, 2020; Cono et al., 2003; Enserink, 2002; Maurer et al., 2003). In the USA, VIG has been approved as a drug for adverse reactions to the smallpox (vaccinia) vaccine. The applicant has stated that VIG will be kept in supply and made available to all involved in the trial if required.

89. Cidofovir is another drug which may be considered as a second line treatment for adverse reactions to VACV. It has a broad spectrum anti-viral activity against most DNA viruses (De Clercq, 2002). While Cidofovir has shown anti-poxviral activity *in vitro* and in mice, there is limited data on its use in humans as a treatment for vaccinia-related adverse events (Centers for Disease Control and Prevention, 2020; Maurer et al., 2003; Wittek, 2006). Cidofovir can also have severe side effects, including irreversible renal toxicity (Centers for Disease Control and Prevention, 2020; Enserink, 2002). The applicant has indicated that they would maintain a supply of Cidofovir on hand during the trial. Cidofovir is available in Australia but is not approved for the treatment of vaccinia-related complications; off-label use would thus be required.

### 3.12 Risk group of VACV

90. The Australian Standard 2243.3:2022 *Safety in Laboratories Part 3: Microbiological safety and containment* (Standards Australia/New Zealand, 2010, 2022) classifies VACV as a risk group 2 organism, and the Australian Immunisation Handbook recommends vaccination of people working with a repeated risk of exposure to, or working with large quantities or concentrations of, *Vaccinia virus* cultures (Australian Technical Advisory Group on Immunisation (ATAGI), 2018, 2024).

### Section 4 The GMO – nature and effect of the genetic modification

91. Oncolytic viruses (OVs) are capable of replicating in cancer cells whilst having minimal impact on healthy cells. In addition to this, these viruses can be genetically modified to stimulate the hosts immune system to further recognise the cancerous cells they replicate in and subsequently mark those cells for destruction (Harrington et al., 2019). Modifications can also be made to abrogate the OVs ability to evade the host immune system and/or enhance their ability to replicate in and kill cancer cells. These properties in combination with their safety profile make OVs a promising treatment for refractory cancers (Zhang and Liu, 2020).

92. Advances in genetic-engineering and molecular virology have enabled progress in the use of OVs in cancer therapy in the last two decades. The 2015 FDA approval of the clinical use of Talimogene laherparepvec (T-VEC), a GM herpes virus for the treatment of melanoma in humans, was the first approval of its kind globally (Ferrucci et al., 2021; Johnson et al., 2015). A number of other OV treatments have demonstrated strong safety profiles in clinical studies (Chaurasiya et al., 2020).

93. Due to its desirable properties such as safety and payload capacity, multiple clinical trials utilising GM VACV as an OV backbone have already been undertaken (Guo et al., 2019). For example, a clinical trial in 16 patients with advanced solid tumours was well-tolerated in patients, resulted in

enriched replication in tumours, and demonstrated antitumor activity (Zeh et al., 2015). Another oncolytic VACV, known as Pexa-Vec, has had acceptable safety profiles when administered in Phase 1 and 2 clinical trials in human patients (Heo et al., 2013; Park et al., 2015). And GL-ONC1, a GM VACV based on the LIVP strain was well tolerated when administered into the peritoneal cavity of patients with advanced stage peritoneal carcinomatosis (Lauer et al., 2018).

94. The sponsor of this trial, Viromissile Inc, has developed IDOV-Immune, an oncolytic viral immunotherapy, for the treatment of patients with advanced solid tumours. IDOV-Immune is based on a vaccinia virus backbone whose derivative strain has been declared as CCI.

### 4.1 Genetic modifications to VACV

95. Viromissile Inc employed a strategy of homologous recombination (Falkner and Moss, 1990) to generate the GM VACV used in this trial. The resulting virus has 3 deleted virulence genes in addition to 3 inserted immunomodulatory genes. Disruption of these non-essential virulence genes and expression of the foreign genes not only attenuate the virus but also enhance its tumour-specific replication and boost anti-tumour immune responses.

96. The 3 inserted immunomodulatory genes are of human origin. The inserted genes activate the innate and adaptive immune response of the host to aid in eliminating the tumour cells that harbour the virus. Further information regarding the deleted and inserted genes has been declared as CCI.

97. Although cancerous tumours invoke some level of innate and adaptive immune response within their microenvironment, they ultimately manage to evade critical components of the anti-tumour immune response. This immune evasion appears to be facilitated by an array of mechanisms, including the modulation of immune suppressive mediators and suppression of regulatory T-cells (Vinay et al., 2015). This observation underlies the rationale for the treatment of such tumours with an OV which can infect them and reactivate the hosts anti-tumour immune response.

98. The immune response induced by these transgenes may additionally generate a long-term immunological memory, which is capable of tumour control and prevention of recurrence.

99. There have been multiple clinical studies utilising GM VACV as an oncolytic therapy which have employed strategies incorporating deletion of virulence genes in combination with insertion of immunomodulatory genes. These studies, some of which are mentioned in paragraph 93, have largely demonstrated the safety of this approach.

### 4.2 Shedding and safety of the GMO

100. Viromissile Inc has conducted a number of pre-clinical studies in cell lines and a model organism aimed at determining the dynamics of viral replication, toxicity, shedding and tissue tropism of the GMO.

101. Data provided by the applicant suggests that shedding of the GMO is minimal. Biodistribution data suggest that the GMO replicates efficiently in tumour cells and poorly in other cells. It should be noted that the number of animals tested is low. Also of note, is that results observed in murine models do not always translate in humans. However, similar clinical trials with VACV based OVs have shown excellent safety profiles in the past. The applicant's pre-clinical results support these observations. Details of these studies have been declared as CCI. Further details around uncertainty are presented in Chapter 2, Section 3.

### 4.3 Stability in the environment and decontamination

102. The modifications made to the VACV are not expected to have any impact on its host range, persistence in the environment, or ability to be destroyed by decontaminants.

### Section 5 The receiving environment

103. The receiving environment forms part of the context for assessing risks associated with dealings with GMOs. It informs the consideration of potential exposure pathways, including the likelihood of the GMOs spreading or persisting outside the site of release.

### 5.1 Clinical trial sites

104. The intended primary receiving environment would be solid tumours within the clinical trial participants. As stated in Chapter 1, Section 2.3.2 each patient would receive a single dose of the GM *Vaccinia virus* as a treatment and be monitored over a period of 28 days. Administration would be via i.v. infusion.

105. The secondary receiving environment would be the hospitals and clinics where the GMO would be dispensed, administered and waste disposed of. These exact sites are yet to be identified. All clinical sites involved in the study would be equipped to handle infectious agents and procedures would be conducted in accordance with the *Australian Guidelines for the Prevention and Control of Infection in Healthcare* (National Health and Medical Research Council, 2019).

106. The principal route by which the GM VACV as a treatment could enter the wider environment is by shedding from inoculated trial participants once they leave the hospital and return home. The tertiary receiving environment includes the trial participant's homes and any places they visit during the period when the therapeutic GM VACV is replicating and shedding.

### 5.2 Relevant environmental factors

107. Environmental factors relevant to the potential persistence or spread of the GMO, or the harm it may cause, include the presence of susceptible hosts and any physical conditions that may aid or restrict transmission to these hosts.

108. The ability of the GMO to physically persist in the environment on surfaces as well as its ability to be decontaminated would be unchanged from the parent organism. These features are described in Section 3.8.

109. The parent organism, VACV, was used worldwide as a vaccine to protect against smallpox infection. The smallpox vaccination program is no longer ongoing, but the majority of people over forty years of age in Australia are likely to have been vaccinated. As a result, a proportion of the population has already been exposed to the vaccinia virus. People vaccinated may be less susceptible to VACV infection, or infection may be asymptomatic or produce less severe symptoms (Cohen, 2001; Hatakeyama et al., 2005).

110. It is widely acknowledged that people for whom smallpox vaccination is contraindicated are more prevalent in the population today than during the era of mass smallpox vaccination. For example, approximately 17% of the Australian population have a history of atopic dermatitis (Chidwick et al., 2020). There are also likely to be significant numbers taking immunosuppressive drugs for disease control (e.g. for autoimmune inflammatory conditions), organ transplant recipients and people with HIV-AIDS.

111. Animals that can be infected with the GMO may be present in environments where it could be shed by trial participants (e.g. patient's homes). Such animals are most likely to include domestic pets and, potentially, livestock.

### 5.3 Related viral species in the receiving environment

112. Although limited in their abundance and distribution, there are some examples of related pox viruses which exist in the Australian environment, which are discussed below.

113. *Molluscum contagiosum virus* (MCV) is one example of widely abundant, human adapted pox virus present in Australia (Konya and Thompson, 1999). Despite being a member of the poxviridae

family, *Molluscum contagiosum* has no close relatives, and is the only member in its genus (Senkevich et al., 1997). The infections it causes are typically confined to children and benign. However, it is more severe and persistent in immunosuppressed patients, particularly in those with HIV/AIDS (Healthdirect Australia 2024).

114. Mpox is another member of the pox virus family present in the environment. However, the level of circulation in the Australian community has remained quite low since the outbreak in 2022 (Healthdirect Australia, 2024). Symptoms of mpox disease resemble that of other pox virus infections including fever, lymphadenopathy and pustule formation (Altindis et al., 2022). It is spread through skin to skin contact as well as contact with contaminated fomites (Alakunle et al., 2020). Although clinical presentation of mpox can be severe, it is very unlikely to result in death (Farahat et al., 2022).

115. Other more distally related and poorly characterised poxviruses are known to infect many native Australian animals, including mammals, birds and reptiles (Wildlife Health Australia, 2019). A recent example was identified in green sea turtles (Sarker et al., 2021). There have been no reports of such viruses infecting humans.

116. Similarly, the deliberately introduced myxoma virus (family Poxviridae, genus Leporipoxvirus) is not known to infect humans but is specific to rabbits and hares, causing lethal disease in some species (Águeda-Pinto et al., 2022; Kerr et al., 2022).

### 5.4 Presence of the introduced genes and encoded proteins in the environment

117. All 3 of the introduced genes are derived from the human genome. Therefore, humans have already been exposed to the proteins that would be produced by these genes in the GMO. However, the gene sequences have been optimised and modified for the purposes of creating the GMO.

### Section 6 Relevant Australian and international approvals

### 6.1 Australian approvals

118. The Regulator has not previously approved any DIR or DNIR licences for dealings with the proposed GMO.

119. The Regulator has issued limited and controlled DIR licences (DIR-140 and DIR-179) utilising VACV for clinical trials in humans. The purpose of DIR-140 is to evaluate the efficacy of GM VACV for treatment of liver, kidney and prostate cancer. The Regulator has also issued a limited and controlled DIR licence (DIR-179) utilising GM VACV for the treatment of solid cancerous tumours.

### 6.2 International approvals

120. There are currently no other international approvals for this GMO.

### Chapter 2 Risk assessment

### Section 1 Introduction

121. The risk assessment identifies and characterises risks to the health and safety of people or to the environment from dealings with GMOs, posed by or as the result of gene technology (The risk assessment process). Risks are identified within the established risk assessment context (Figure 2), taking into account current scientific and technical knowledge. A consideration of uncertainty, in particular knowledge gaps, occurs throughout the risk assessment process.



### Figure 2. The risk assessment process

122. The Regulator uses a number of techniques to identify risks, including checklists, brainstorming, reported international experience and consultation (OGTR, 2013).

123. Risk scenarios examined in RARMPs prepared for licence applications for the same or similar GMOs, are also considered.

124. Risk identification first considers a wide range of circumstances in which the GMO, or the introduced genetic material, could come into contact with people or the environment. This leads to postulating plausible causal pathways that may give rise to harm for people or the environment from dealings with a GMO. These are called risk scenarios.

125. Risk scenarios are screened to identify substantive risks, which are risk scenarios that are considered to have some reasonable chance of causing harm. Risk scenarios that could not plausibly occur, or do not

lead to harm in the short and long term, do not advance in the risk assessment process (Figure 3), i.e. the risk is considered to be no greater than negligible.

126. Risk scenarios identified as substantive risks are further characterised in terms of the potential seriousness of harm (consequence assessment) and the likelihood of harm (likelihood assessment). The consequence and likelihood assessments are combined to estimate the level of risk and determine whether risk treatment measures are required. The potential for interactions between risks is also considered.

### Section 2 Risk identification

127. Postulated risk scenarios are comprised of three components (Figure 3):

- i. the source of potential harm (risk source)
- ii. a plausible causal linkage to potential harm (causal pathway)
- iii. potential harm to people or the environment.



### Figure 3. Components of a risk scenario

128. When postulating relevant risk scenarios, the risk context is taken into account, including the following factors detailed in Section 2:

- the proposed dealings
- the proposed limits including the extent and scale of the proposed dealings
- the proposed controls to limit the spread and persistence of the GMO and
- the characteristics of the parent organism(s).

### 2.1 Risk source

129. The parent organism of the GMO is VACV. Details on the pathogenicity and transmissibility of VACV is provided in Chapter 1 (Section 3). Vaccination with VACV tends to produce a pustule at the inoculation site. Transmission of VACV from the vaccinee to other people and susceptible hosts, such as domestic pets, could occur from this site.

130. The sources of potential harms can be intended novel GM traits associated with one or more introduced genetic elements, or unintended effects/traits arising from the use of gene technology.

131. As discussed in Chapter 1 (Section 4), the GMO has been modified by the introduction of 3 human genes and deletion of 3 viral genes, intended to produce an oncolytic trait. These modifications are considered further as sources of potential harm.

132. The expression of the introduced genes is controlled by poxviral regulatory sequences. Regulatory sequences are naturally present in all organisms and the introduced/endogenous sequences are expected to operate in similar ways to endogenous sequences. The regulatory sequences are DNA that is not expressed as a protein; they are poxvirus specific and do not present a risk in the absence of poxvirus cellular machinery. Hence, potential harms from the regulatory sequences will not be further assessed for this application.

133. The genetic modifications involving introduction of genes have the potential to cause unintended changes to viral characteristics due to insertional effects such as interruptions, deletions, duplications or rearrangements of the genome. Pathways to any unintended effects in poxviruses have been already considered in the RARMP for <u>DIR 116</u>, and found to be negligible. Their likelihood will be minimised by the

proposed limits and controls. These include the requirement for any unintended effects to be reported to the Regulator immediately. Therefore, the potential for the processes of genetic modification to result in unintended effects will not be considered further.

134. Infection with VACV does not result in latent infection or integration into the host genome, and this will not be considered further.

### 2.2 Causal pathway

135. The following factors are taken into account when postulating plausible causal pathways to potential harm:

- proposed dealings
- proposed limits including extent and scale of the proposed dealings
- proposed controls to limit the spread and persistence of the GMOs
- routes of exposure to the GMOs, the introduced gene(s) and gene product(s)
- potential effects of the introduced gene(s) and gene product(s) on the properties of the organism
- potential exposure of other organisms to the GMOs in the environment
- the environment at the site(s) of release
- spread and persistence of the GMOs (e.g. dispersal pathways and establishment potential)
- tolerance to abiotic conditions (e.g. temperature, UV irradiation and humidity)
- gene transfer by horizontal gene transfer (HGT)
- unauthorised activities, and
- practices during and after administration of the GMOs.

136. Although all of these factors are taken into account, some are not included in the risk scenarios below as they may have been considered in previous RARMPs and a plausible pathway to harm could not be identified.

137. As discussed in Chapter 1(Section 1.1), the TGA, the trial sponsor, the Investigators and HREC all have roles in ensuring the safety of trial participants under the *Therapeutic Goods Act 1989*, and human clinical trials must be conducted in accordance with the *National Statement on Ethical Conduct in Human Research* (National Health and Medical Research Council et al., 2023). Therefore, risk scenarios in the current assessment focus primarily on risks posed to people other than those participants in the trial, and to the environment.

138. *Vaccinia virus* is transmitted through direct contact. Aerosol transmission is not considered as a viable route of infection for the GMO (see Paragraph 71). Therefore, aerosol transmission will not be considered further.

139. The GMOs and samples containing the GMOs are proposed to be transported and stored in line with the Regulator's *Guidelines for the Transport, Storage and Disposal of GMOs*. These are standard protocols for the handling of GMOs to minimise exposure to the GMOs, so risks associated with such transport will not be further assessed.

140. The Act provides for substantial penalties for unauthorised dealings with GMOs or noncompliance with licence conditions, and also requires the Regulator to have regard to the suitability of an applicant to hold a licence prior to the issuing of the licence. These legislative provisions are considered sufficient to minimise risks from unauthorised activities. Therefore, unauthorised activities will not be considered further.

### 2.3 Potential harm

141. In addition, the following factors are taken into account when postulating relevant risk scenarios for this licence application:

 harm to the health of people or desirable organisms, including disease in humans or animals or adverse immune response • the potential for establishment of a novel virus in the environment.

### 2.4 Postulated risk scenarios

142. Three risk scenarios were postulated and screened to identify any substantive risks. These scenarios are summarised in Table 1 and examined in detail in Sections 2.4.1 - 2.4.3 (this Chapter).

143. In the context of the activities proposed by the applicant and considering both the short and long term, none of the three risk scenarios were considering to be substantive.

Risk scenario	Risk source	Causal Pathway	Potential harm	Substantive risk?	Reasons
1	GMO	<ul> <li>i. Exposure of people undertaking dealings in clinical trial facilities to GMO via:</li> <li>needle stick/ sharps injury/ eye splash during GMO preparation, administration or sample analysis</li> <li>GMO contact with abraded skin</li> <li>contact with GMO contaminated materials</li> <li>ii. Transduction of cells</li> <li>iii. Replication of the GMO and expression of immunomodulatory transgenes</li> <li>iv. Further transmission to people or animals</li> </ul>	Adverse immune response Vaccinia-like disease, including serious adverse reactions	No	<ul> <li>Only trained and experienced personnel would prepare, administer and handle the GMO. These personnel would also be trained and experienced in the use and disposal of sharps.</li> <li>Use of PPE (e.g. gown, gloves, mask and eye protection) minimises the potential for exposure to staff handling the GMO.</li> <li>High-risk personnel are excluded from handling the GMO.</li> <li>Sample testing would be conducted by qualified personnel in pathology or other testing laboratories.</li> <li>The GMO is designed for enriched replication in cancer cells and less efficient replication in healthy cells. It is expected to be rapidly cleared by the immune response in healthy cells.</li> <li>Accidental exposure would only involve a small dose of GMO and the person would receive medical attention and would be monitored for symptoms.</li> <li>Exposed personnel would be instructed to cover pustules should they occur and avoid contact with high-risk groups and animals.</li> <li>Inadvertent exposures with wild-type VACV in healthy people documented to date did not lead to clinically significant symptoms or did not require treatment beyond first aid and observation.</li> </ul>
2	GMO	<ul> <li>i. Trial participant injected/infused with the GMO</li> <li> ■ ii. The GMO is shed at the infusion sites or in body fluids </li> </ul>	Adverse immune response Vaccinia-like disease, including serious	No	<ul> <li>In addition to the reasons described in Risk scenario 1:</li> <li>High-risk trial participants, including immunocompromised persons and pregnant people, would be excluded.</li> <li>Residual inoculum GMO is unlikely to be present at the site of administration as the line would be flushed and the infusion site covered</li> </ul>

Table 1. Summary of risk scenarios from the proposed dealings with the GMOs

Risk scenario	Risk source	Causal Pathway	Potential harm	Substantive risk?	Reasons
		<ul> <li>iii. The GMO is released to the environment, exposing other people or animals to the GMO</li> <li>iv. Infection of host cells</li> <li>v. Replication of the GMO and expression of the immuno-modulatory transgenes</li> <li>vi. Further transmission to people or animals</li> </ul>	adverse reactions		<ul> <li>with an occlusive dressing. The dressing would be changed every few days and disposed of in a biohazard bin.</li> <li>The site of administration will be monitored for infection by participants.</li> <li>Trial participants would be educated on the proper handling of wound dressings as well as towels and clothing which might come into contact with the site of administration.</li> <li>All trial participants would be using barrier contraception to prevent pregnancy and transmission of the GMO for at least 60 days after the last GMO treatment.</li> <li>If the trial participant develops vaccinia-related lesions or pustules, they would be instructed to launder sheets and clothing separately to other laundry in soapy water.</li> <li>Participants will have access to a pustule kit to appropriately clean and cover the lesions.</li> <li>Bandages, dressings and other materials used to care for vaccinia-related lesions would be disposed in a biohazard bin. The biohazard bin would be returned to the clinical trial site for disposal.</li> <li>The limited number of clinical trial participants and education on transmission pathways is likely to be at a low dose and unlikely result in infection. The transmission rate from people who have received a VACV vaccine to other people in more recent vaccination programmes is low, as described in paragraph 73.</li> </ul>
3	GMO	<ul> <li>i. Trial participant injected/infused with the GMO</li> <li>Ii. Trial participant becomes or is already infected with another compatible virus</li> <li>Iii. The GMO recombines with</li> </ul>	Novel disease in humans Establishment of novel virus with unknown pathogenicity in the environment	No	<ul> <li>Prior treatment with a poxvirus- based treatment or vaccine is an exclusion criteria. This reduces the likelihood of similar genomic sequences that are available for recombination.</li> <li>Patients with evidence of any active systemic infection will also be excluded from the trial.</li> <li>There is no reservoir of VACV in the Australian environment and limited opportunity for the GMO to come</li> </ul>

Risk scenario	Risk source	Causal Pathway	Potential harm	Substantive risk?	Reasons
		<ul> <li>another virus in the host</li> <li>iv. Produces a replication competent recombinant virus</li> <li>v. Recombinant virus is shed</li> <li>vi. Recombinant virus infects new hosts and is more virulent or pathogenic.</li> </ul>			<ul> <li>into contact with other related poxviruses.</li> <li>For recombination to occur, the GMO and another poxvirus need to be present in the same cell at the same time.</li> <li>Recombination is also limited by the size of VACV and viral factory compartmentalisation. The large 25 kb deleted region, housing multiple <i>Vaccinia</i> genes would need to be acquired for the GMOs to regain its replicative ability in healthy cells.</li> </ul>

### 2.4.1 Risk scenario 1

Risk source	GMO		
	<ul> <li>i. Exposure of people undertaking dealings in clinical trial facilities to GMO via:</li> <li>needle stick/ sharps injury/ eye splash during GMO preparation, administration, collection or</li> </ul>		
Causal pathway	<ul> <li>analysis of samples</li> <li>GMO contact with abraded skin</li> <li>contact with GMO contaminated materials</li> </ul>		
	ii. Transduction of cells		
	<ul> <li>Expression of immunomodulatory transgenes</li> </ul>		
	iv. Further transmission to people or animals		
Potontial harm	Adverse immune response		
Fotentiai naim	Vaccinia-like disease, including serious adverse reactions		

### **Risk source**

144. The source of potential harm for this postulated risk scenario is the GMO as a treatment.

### **Causal Pathway**

145. There are a number of ways that people may be exposed to the GMOs while undertaking the dealings as part of this trial. The GMO treatment would be prepared and infused into clinical trial patients with solid tumours. Biological samples, including blood, anal swabs, and urine would be collected throughout the trial. During these dealings, there is a potential risk of exposure to people involved in the trial via needle stick, sharps injury and/or eye splash.

146. Exposure via needle stick may occur during the dilution of the GMO in which a syringe and needle is required to extract the GMO from the vial. It might also occur whilst administering the GMO into the i.v

bag. An eye splash might occur if either a vial containing the GMO or the i.v infusion bag used for administration is perforated or dropped.

147. Blood samples will be collected by trained staff at the clinical trial site. These samples may be prepared and analysed at the trial site or shipped to pathology laboratories. Controls proposed by the applicant, including appropriate training and the use of containment equipment, would minimise this risk. Use of PPE (e.g. gown, gloves, mask and eye protection) would minimise the potential for exposure of staff handling the GMO within the trial site. Analyses of samples outside the trial site would be conducted by qualified personnel in pathology or other testing laboratories, which are required to adhere to national standards for handling of infectious substances and are considered sufficient to prevent the personnel exposure to the GMO. Additionally, appropriate decontamination and disposal practices would prevent persistence and spread of the GMO.

148. In the event of an exposure, personnel would receive immediate medical attention by trained staff. The exposed individuals would then be monitored for any developing symptoms. The staff would be instructed to cover pustules should they occur and to avoid contact with high-risk groups and animals. This measure would minimise the potential transmission of the GMO to other people and animals.

149. The applicant has stated that treatments for VACV infection such as VIG will be kept on hand during dealings with the GMO.

150. The above-mentioned limits and controls would minimise the potential exposure of people to the GMOs via needle stick, sharps injury and/or eye splash.

### Exposure via contact of abraded skin with infusion site

151. As mentioned in Chapter 1, Section 3.6, VACV is transmitted through close physical contact between infected and non-infected people or animals. If people in clinical trial facilities come in contact with the infusion site after patient treatment, they could be exposed to the GMO.

152. Transmission of VACV from the treated trial participant to another person would require close contact with abraded skin. The applicant has stated that the i.v. line would be flushed with sterile normal saline after administration and that the infusion site would be coved by an occlusive dressing. Together, these measures would limit the dissemination of the GMO from the infusion site.

### Exposure by contact with contaminated materials

153. If people in clinical trial facilities come in contact with GMO contaminated materials, they could be exposed to the GMO. As discussed in Chapter 1, Section 3.8, VACV can remain viable for extended periods under certain circumstances. The applicant has stated that GMO waste and materials contaminated with the GMO would be disposed according to infectious medical waste management procedures (Chapter 1, Section 2.3.8). This would minimise any potential exposure to the GMO via contaminated waste.

### **Potential harm**

154. If people undertaking dealings in clinical trial facilities are exposed to the GMOs via needle stick, sharps injury, eye splash or via close contact to an unhealed infusion site, or via GMO waste they could suffer symptoms of VACV infection, and on rare occasions an adverse immune response such as those described in Section 3.1.

155. Although the vaccinia strain modified to create the GMOs is capable of replicating in human cells, and cause illness in humans (Chapter 1, Section 3), the GMO has been modified to be have poor replication efficiency in healthy cells and be replication competent in cancer cells (Chapter 1, Section 4). It is expected to be cleared by the immune system if it transduces healthy cells. The three genes introduced into the GMO all stimulate the host immune response, whilst the three deleted genes limit replicative function in healthy cells. The combination of these changes increases the ability of the host to clear the virus in healthy cells. Thus, any exposure of people undertaking dealings in clinical trial facilities is unlikely to result in viral infection/disease, and depending on the level of exposure, would likely only result in an acute reaction. Inadvertent exposures with wild-type VACV in healthy people documented to date did not lead to clinically

significant symptoms or require treatment beyond first aid and observation (Cono et al., 2003; Fulginiti et al., 2003b; Maurer et al., 2003).

156. Those who are immunocompromised, who are more likely to suffer a severe adverse reaction upon exposure, would be excluded from preparing or administering the GMO.

157. Although the persistent over-expression of all three immunoregulatory proteins could result in an adverse reaction, the rapid clearance of the virus would greatly reduce the risk of this occurring. Further mitigation of this risk comes in the form of the 3 deleted genes in the GMO, all of which would aid the host in clearing the virus from healthy tissues. Furthermore, the transgenes expressed by the GMO are all of human origin and therefore unlikely to be allergenic to exposed persons.

158. In addition to the points raised above, the applicant has provided some pre-clinical data which suggests that the rate of shedding of the GMO is minimal in a model organism. It also appears to have enriched replication in tumour cells. This data and its interpretation has been declared as CCI.

### Conclusion

159. Risk scenario 1 is not identified as a substantive risk because potential exposure would be limited by the proposed limits and controls, and the GMO is designed to replicate less efficiently in healthy cells and be enriched in cancer cells. Any infection caused by the GMO is likely to be cleared by the host immune response without significant complication. Therefore, the potential for an unintentional exposure of people undertaking dealings at clinical trial sites to the GMO resulting in ill health in humans and animals is not identified as a risk that could be greater than negligible and does not warrant further detailed assessment.

### 2.4.2 Risk scenario 2

Risk source	GMO		
	i. Trial participant injected/infused with the GMO ↓		
	ii. The GMO is shed at the infusion site or in body fluids ↓		
	iii. The GMO is released to the environment, exposing other people or animals to the GMO.		
Causal pathway	•		
cuusui putiiway	iv. Infection of host cells		
	+		
	v. Replication of the GMO and expression of the immuno-modulatory transgenes		
	•		
	i. Further transmission to people or animals		
Potential harm	Adverse immune response		
Fotentiai naim	Vaccinia-like disease, including serious adverse reactions		

### **Risk source**

160. The source of potential harm for this postulated risk scenario is the GMO as a treatment.

### **Causal Pathway**

161. Following administration, the GMO could be shed from the patients at infusion site or in bodily fluids. It could subsequently be transmitted to a person who came into close contact with either the patient, or a patient-generated fomite.

162. The clinical trial would involve i.v. infusion as the mode of administration, leading to the presence of the GMO in the blood in the hours following administration. However, the GMO is expected to be quickly cleared by the immune system due to the modifications to the GMO, which stimulate the immune response and limit viral replication outside of cancer cells.

163. Transmissible VACV of both high and low pathogenic strains can shed into the faeces and urine of experimentally infected mice. When VACV is administered as a vaccine, viremia (viral presence in the blood) and viruria (viral presence in urine) in humans is uncommon. It does occur in patients with progressive vaccinia and eczema vaccinatum (Lane and Fulginiti, 2003). These conditions are most likely to be manifested in persons who are immunocompromised and have a history of severe skin disease, respectively. Such persons are excluded from participating in the clinical trial.

164. The applicant has proposed that all trial participants would be using barrier contraception to prevent pregnancy and transmission of the GMOs. As mentioned in Chapter 1, Section 3.6, VACV is usually transmitted through physical contact with lesions or pustules, or contact with virus contaminated materials. The use of barrier contraception would prevent incidental transmission through physical contact, and pregnancy.

165. The applicant has provided some pre-clinical data which suggests that the rate of shedding of the GMO is minimal in a model organism. It also appears to replicate poorly in healthy cells and is enriched in tumour cells. This data is declared as CCI.

166. The GMO could be transmitted from the trial participants should they develop vaccinia-related skin or oral pustules. Trial participants would be informed about risks associated with the GMO and instructed to follow appropriate hygiene protocols and disposal of contaminated dressings.

167. In addition, all trial participants will be provided with a pustule management kit and those who develop lesions would be instructed to follow the pustule management plan as described in paragraph 24. The management plan would be explained to prospective participants during initial screening and anyone unwilling or unable to comply would not be enrolled in the trial. Trial participants would also be expected to seal contaminated disposable items in a provided primary container (e.g. biohazard bag) and then place this into a provided secondary container (biohazard bin). At each visit, trial participants would return the biohazard bin to the clinical trial site for disposal as clinical waste. Participants would also be advised to launder contaminated fabrics in hot soapy water. The trial participant would also be instructed to change the dressing privately, unless they require assistance from a caregiver, and limit access to any pets (particularly dogs), other animals, or higher-risk individuals (see paragraph 33). If another person or animal develops a suspicious rash, this would be reported and may be examined by the clinical trial investigator. Together, these measures would minimise potential transmission of the GMO to other people and animals.

168. The secondary transmission rate from people who have received a VACV as part of vaccination programmes is low, as described in paragraph 73. Further indication of the relatively low risk of viral dissemination is evident in a 2006 study testing a range of environmental samples that could have been in contact with a bandaged pustule from VACV infected persons. These samples were found to be negative for live virus as determined by plaque infectivity assay (Stark et al., 2006). The limited number of clinical trial participants and education on transmission pathways is likely to reduce transmission.

### **Potential harm**

169. If people or animals (pets, wildlife, livestock) are exposed to the GMO, a range of outcomes are possible. The potential harm to people exposed to the GMO are adverse immune responses and vaccinia-like diseases and reactions as described in Risk scenario 1. If a susceptible animal is exposed to the GMO, it could lead to infection, shedding of the GMO in the environment via faeces or urine and exposure of other animals and people. This form of exposure is unlikely to cause harm because in addition to involving a low number of viral particles, the GMO was designed to preferentially replicate in cancer cells and it is expected to be cleared by the immune system if it infects healthy cells.

### Conclusion

170. Risk scenario 2 is not identified as a substantive risk because potential exposure would be limited by the proposed limits and controls (including bandaging of pustules), and the GMO are designed to have enriched replication in cancer cells and poorer replication efficiency in healthy cells. Therefore, this risk could not be greater than negligible and does not warrant further detailed assessment.

### 2.4.3 Risk scenario 3

Risk source	GMO		
	i. Trial participant injected/infused with the GMO		
	<ul> <li>Trial participant becomes or is already infected with another virus</li> </ul>		
Causal pathway	iii. The GMO recombines with another virus in the host		
,	iv. Produces a replication competent recombinant virus		
	v. Recombinant virus shed		
	vi. Recombinant virus infects new hosts and is more virulent or pathogenic.		
	Novel disease in humans or animals		
Potential harm	Establishment of novel virus with unknown pathogenicity in the environment		

### **Risk source**

171. The source of potential harm for this postulated risk scenario is the GMO as a treatment.

### **Causal Pathway**

172. Should the trial participant be infected by other viruses (either incidentally or through vaccination or treatment), recombination can occur between viral types if they simultaneously infect the same cell. Similarly, another host could become infected with the GMOs through accidental exposure, and either have an existing viral infection or acquire one while the GMO is present. If recombination occurs, the introduced genes could potentially restore the replication competence of the GMO in healthy cells. The resulting novel recombinant virus could then be more readily shed from its host.

173. Even though poxviruses can infect a wide range of organisms, specific viruses have a variable host range, and some are restricted to a single host (Oliveira et al., 2017b). While recombination between different classes of virus can occur, the GM virus is more likely to recombine with another poxvirus than with an unrelated virus (see RARMP for <u>DIR 116</u>).

174. As described in the RARMPs for <u>DIR 140</u>, <u>DIR 170</u>, and <u>DIR-179</u>, *Molluscum contagiosum* virus (MCV) is likely to be present in the Australian population and MCV infection is more prevalent in school-aged children, adolescents, and young adults than in older adults (Konya and Thompson, 1999). As described in the RARMP for <u>DIR 140</u>, there are no reports on the ability of MCV to recombine with other poxviruses, MCV has co-existed with variola virus (the causative agent for smallpox) for thousands of years, and with VACV for over 150 years, without evidence of recombinants forming and persisting in the human population. These observations provide strong evidence that the propensity for VACV recombination with related viruses in the environment is very low.

175. In the case of mpox, whilst it is known to recombine with variants within its own strain (Yeh et al., 2022), there are no reports of mpox recombining with other pox viruses. During the first 3 quarters of 2024, approximately 700 cases of mpox have been detected Australia (Australian Government Department of Health, 2024). The symptomatic nature of most mpox infections and the fact that it is a notifiable disease, is likely to limit the degree of undetected transmission (Altindis et al., 2022; Ježek et al., 1987).

176. Although present in the Australian environment, myxoma virus only infects rabbits and hares, and is therefore unlikely to come into contact with a trial participant.

177. Although there have been reports of VACV reservoirs in Latin America (José da Silva Domingos et al., 2021), there currently exists no evidence that this has entered the Australian environment. Thus, recombination between GM and wild-type VACV because of the proposed clinical trial is highly unlikely.

178. As discussed in Risk scenario 1, the GMO is designed to be have poorer replication efficiency in healthy cells and is expected to localise to cancer cells and be cleared by the immune system in healthy cells. This reduces the likelihood that co-infection of the same cells with another pox virus would occur and constitutes a barrier for potential recombination.

179. Finally, in order for a recombination event to occur with mpox, MCV or another similar poxvirus, both viruses would have to co-infect the same cells of the same person. Once this event occurred, the two viral replication factories would have to be involved in DNA replication at the same time and would have to fuse (Katsafanas and Moss, 2007; Lin and Evans, 2010; Paszkowski et al., 2016). Once the two viral replication factories had fused, compatible regions of each genome would have to be exchanged (Paszkowski et al., 2016). For this highly unlikely series of events to lead to harm, the swapped regions of the genome would have to confer increased virulence or pathogenicity. Given that the widespread use of VACV vaccines across multiple centuries has never resulted in an outcome such as the one described, the likelihood of this occurring during this trial is considered highly unlikely.

### **Potential harm**

180. If the GMO recombines with another poxvirus, it could lead to a novel replication competent virus, it could then be shed from the host and cause disease in humans or animals, or lead to the establishment of a novel virus in the environment.

181. Any virus formed in this way, would result in progeny having any permutation of genomic segments of the two parent strains. Even in the unlikely scenario that recombination with a co-infecting poxvirus was able to generate a new replication competent poxvirus, it is not expected that recombination would lead to a virus that is more pathogenic or virulent than the wild type circulating poxvirus. This is because the deleted regions of the GMO are designed to reduce pathogenicity and the inserted genes are designed to enhance the immune response, which would likely result in enhanced viral clearance.

### Conclusion

182. Risk scenario 3 is not identified as a substantive risk because recombination is unlikely due to the natural barriers to recombination, and the lack of history of recombination in poxviruses leading to harm. Therefore, this risk could not be greater than negligible and does not warrant further detailed assessment.

### Section 3 Uncertainty

183. Uncertainty is an intrinsic part of risk analysis and is present in all aspects of risk analysis. This is discussed in detail in the Regulator's <u>Risk Analysis Framework</u> document.

184. Uncertainty is addressed by approaches such as balance of evidence, conservative assumptions, and applying risk management measures that reduce the potential for risk scenarios involving uncertainty to lead to harm. If there is residual uncertainty that is important to estimating the level of risk, the Regulator will take this uncertainty into account in making decisions.

185. As clinical trials are designed to gather data, there are generally data gaps when assessing the risks of a clinical trial application involving GMOs. However, clinical trial applications are required to be limited and

controlled. Even if there is uncertainty about the characteristics of a GMO, limits and controls restrict exposure to the GMO, and thus decrease the likelihood of harm.

186. For DIR-208, uncertainty is noted in relation to:

- the enriched replication of the GMO in cancer cells over healthy cells
- shedding of the GMO in humans
- the effect of the transgenes in vivo.

187. As this is a first in human trial, ViroMissile Inc has conducted several non-clinical studies in a model species. These data are declared as CCI. Given that the applicant's pre-clinical viral shedding studies were conducted in a model organism, it is possible that the results observed would not be replicated in human. There is uncertainty as to whether data gathered in the model species would be transferrable to humans.

188. While some uncertainty remains, it is unlikely that the GMOs would behave very differently compared to the similar stains of VACV, which have been used in humans during the smallpox eradication programme. The exact parental strain of VACV being used in this trial has not been tested before in humans and so there is a small degree of uncertainty regarding its behaviour following infection. That being said, the deletion of vaccinia genes and introduction of genes, which enhance the immune response (Chapter 1, Section 4), are designed to increase the safety profile of the GMO should it be inadvertently exposed to people other than trial participants.

189. Overall, the level of uncertainty in this risk assessment is considered low and does not impact on the overall estimate of risk.

190. Additional data, including information to address these uncertainties, may be required to assess possible future applications with reduced limits and controls, such as a larger scale trial or the commercial release of these GMOs.

191. Chapter 3, Section 4, discusses information that may be required for future release.

### Section 4 Risk evaluation

192. Risk is evaluated against the objective of protecting the health and safety of people and the environment to determine the level of concern and, subsequently, the need for controls to mitigate or reduce risk. Risk evaluation may also aid consideration of whether the proposed dealings should be authorised, need further assessment, or require collection of additional information.

193. Factors used to determine which risks need treatment may include:

- risk criteria
- level of risk
- uncertainty associated with risk characterisation
- interactions between substantive risks.

194. Three risk scenarios were postulated whereby the proposed dealings might give rise to harm to people or the environment. In the context of the limits and controls proposed by the applicant, and considering both the short and long term, none of these scenarios were identified as substantive risks. The principal reasons for these conclusions are summarised in Table 1 and include:

- the GMO has been designed to have enriched replication in cancer cells
- limited ability and opportunity for the GMOs to transfer the introduced genes through recombination
- suitability of limits and controls proposed by the applicant.

195. Therefore, risks to the health and safety of people, or the environment, from the proposed release of the GMO into the environment are considered to be negligible. The *Risk Analysis Framework* (OGTR, 2013), which guides the risk assessment and risk management process, defines negligible risks as risks of no

discernible concern with no present need to invoke actions for mitigation. Therefore, no additional controls are required to treat these negligible risks. Hence, the Regulator considers that the dealings involved in this proposed release do not pose a significant risk to either people or the environment.

### Chapter 3 Risk management plan

### Section 1 Background

196. Risk management is used to protect the health and safety of people and to protect the environment by controlling or mitigating risk. The risk management plan addresses risks evaluated as requiring treatment and considers limits and controls proposed by the applicant, as well as general risk management measures. The risk management plan informs the Regulator's decision-making process and is given effect through licence conditions.

197. Under Section 56 of the Act, the Regulator must not issue a licence unless satisfied that any risks posed by the dealings proposed to be authorised by the licence can be managed in a way that protects the health and safety of people and the environment.

198. All licences are subject to three conditions prescribed in the Act. Section 63 of the Act requires that each licence holder inform relevant people of their obligations under the licence. The other statutory conditions allow the Regulator to maintain oversight of licensed dealings: Section 64 requires the licence holder to provide access to premises to OGTR inspectors and Section 65 requires the licence holder to report any information about risks or unintended effects of the dealing to the Regulator on becoming aware of them. Matters related to the ongoing suitability of the licence holder must also be reported to the Regulator.

199. The licence is also subject to any conditions imposed by the Regulator. Examples of the matters to which conditions may relate are listed in Section 62 of the Act. Licence conditions can be imposed to limit and control the scope of the dealings and to manage risk to people or the environment. In addition, the Regulator has extensive powers to monitor compliance with licence conditions under Section 152 of the Act.

### Section 2 Risk treatment measures for substantive risks

200. The risk assessment of risk scenarios listed in Chapter 2 concluded that there are negligible risks to people and the environment from the proposed clinical trial of GMO. These risk scenarios were considered in the context of the scale of the proposed clinical trial (Chapter 1, Section 2.1), the proposed controls (Chapter 1, Section 2.2), and the receiving environment (Chapter 1, Section 5), and considering both the short and the long term. The risk evaluation concluded that no specific risk treatment measures are required to treat these negligible risks. Limits and controls proposed by the applicant and other general risk management measures are discussed below.

### Section 3 General risk management

201. The limits and controls proposed in the application were important in establishing the context for the risk assessment and in reaching the conclusion that the risks posed to people and the environment are negligible. Therefore, to maintain the risk context, licence conditions have been imposed to limit the number of trial participants, the types of sites where the GMO can be prepared and administered and the duration of the trial, as well as a range of controls to restrict the spread and persistence of the GMOs and their genetic material in the environment. The conditions are discussed and summarised in this Chapter and listed in detail in the licence.

### 3.1 Limits and controls on the clinical trial

202. Sections 2.1 and 2.2 in Chapter 1, list the limits and controls proposed by Novotech. Many of these are discussed in the three risk scenarios considered in Chapter 2. The appropriateness of the limits and controls is considered further in the following sections.

### 3.1.1 Consideration of limits and controls proposed by Novotech

203. The proposed clinical trial would involve a maximum of 40 participants within Australia, and most dealings with the GMOs would take place in medical facilities such as clinical trial units, hospitals and analytical laboratory facilities. Activities that would occur outside of medical facilities include transport, storage and disposal of the GMOs. The applicant has proposed to complete the study within 5 years of commencement. Conditions maintaining the risk context and proposed limits of the trial, such as the maximum number of trial participants and duration of the study, have been included in the licence.

204. The applicant advised that import and transport of the GMO and waste containing the GMO would be in accordance with IATA shipping classification UN 3373 [Category B] and/or the Regulator's *Guidelines for the Transport, Storage and Disposal of GMOs.* These are standard protocols for the handling and minimising exposure to the GMOs. Once at the clinical trial site, access to the GMO would be restricted to appropriately trained personnel. These proposed transport conditions are suitable for the GMO. Therefore, the licence details the minimum requirements for packaging and labelling the GMO and waste contaminated with the GMO for transport and storage within a clinical trial site, as well as transport of the GMO for export. These measures would limit the exposure of people and the environment to the GMOs.

205. There is an ongoing multi-country outbreak of monkeypox virus and local transmission of the virus has been reported. Therefore, as a precaution, the licence requires the licence holder to provide a written methodology to reliably detect the GMO, or the presence of the genetic modifications described in this licence in a person. The written methodology must be provided to the Regulator at least 14 days prior to first administering the GMO.

206. There are proposed inclusion and exclusion criteria for both trial participants and staff as listed in Chapter 1, Section 2.3.4 and paragraph 44. The inclusion and exclusion criteria for trial participants would be subject to approval by a HREC, who would consider the safety of the individuals involved in the trial. There is limited data regarding exposure of pregnant women, young children and immunocompromised individuals to VACV. While some studies suggest that VACV vaccination does not increase the overall risk of negative pregnancy outcomes (Badell et al., 2015), the CDC advises that VACV vaccines should not be administered to pregnant women in the absence of smallpox exposure (Center for Disease Control and Prevention, 2024). Additionally, severe adverse events are strain dependent and more common in immunocompromised individuals, children under 12 months of age and those with skin disease; such groups are also excluded from VACV vaccination. Therefore, as a precaution, the licence requires that trial participants who are immunocompromised, suffer from severe skin disease, and women who are pregnant or breastfeeding are excluded. This also serves to minimise the potential for spread and persistence of the GMO as people in these groups are more likely to experience a serious adverse reaction. For example, the pathophysiology of skin conditions such as atopic dermatitis can result in a defective skin barrier function, epidermal hyperplasia, and abnormal immune responses, this can enable systemic spread of poxvirus infection (Reed et al., 2012). When VACV is used to vaccinate against smallpox, potential pustule formation is likely to occur within seven days. Given this, the licence conditions would exclude clinical trial staff who are suffering recurrent or active skin disease or immunocompromised from providing direct care for at least seven days after each infusion treatment or if pustules are present.

207. Staff who are immunocompromised and/or suffer from severe skin disease are also excluded from conducting dealings with the GMOs. In addition, as it is a first in human clinical trial, a condition is included in the licence that the licence holder should inform persons preparing and administering the GMOs of the risks associated with the GMO.

208. The applicant advised that the GMO would be administered to trial participants via i.v. infusion by clinical staff at clinical trial sites. The applicant has also proposed that clinical staff would wear PPE including gown, gloves and eye protection. These practices would minimise exposure of people handling and administering the GMOs (Risk scenario 1) and have been included in the licence conditions.

209. The applicant proposed to prepare the GMO in a BSC II and that staff preparing and administering the GMO wear face masks. As discussed in paragraph 71 and paragraph 138, aerosols are not considered as a plausible pathway for exposure to the GMO. Therefore, the use of a BSC II and face masks have not been included in the licence conditions.

210. Conditions are included in the licence requiring the licence holder to ensure that all GMOs, including material or waste that has been in contact with the GMO, within the clinical trial site, are decontaminated by autoclaving, chemical treatment or by high-temperature incineration. If external service providers are used for waste destruction this must be through a clinical waste stream.

211. The Industry Code of Practice for the Management of Clinical and Related Wastes details requirements for clinical waste including waste segregation, packaging, labelling, storage, transport and accountability (Biohazard Waste Industry, 2010). The clinical waste stream typically involves destruction of infectious waste by incineration or autoclaving, which are considered appropriate for disposal of the GMO. Given that VACV can persist in the environment (Chapter 1, Section 3.8) and compatible hosts such as rodents, marsupials and others as listed in paragraph 61 would be present in the Australian environment, disposal measures such as burial or maceration would not ensure containment. Therefore, the licence also requires waste disposal by external service providers to be by autoclaving or high-temperature incineration. These measures would limit the exposure of people or other animals to the GMOs.

212. The applicant has proposed to provide patients with instructions should suspicious skin pustules develop, and provide instructions to patients of good hand hygiene and cough etiquette practices. They will also provide trial participants with a pustule kit and biohazard bin, as described in paragraph 24. Together, these instructions, pustule kit and biohazard bin would limit the exposure of people or other animals to the GMOs. The applicant has also stated that the trial participant would be instructed to launder sheets and clothing in a 10% bleach solution, and separately to other laundry, should pustule(s) develop. Given that VACV can be inactivated with antimicrobial soap and water or bleach with 0.5% sodium hypochlorite (Section 3.8), the requirement for 10% bleach has not been imposed in the licence conditions.

213. Part of the pustule management plan is for the trial participants to avoid high-risk individuals (paragraph 24). When VACV is used to vaccinate against smallpox, if pustule formation is to occur, it is likely to occur within seven days. Given this, licence conditions are proposed to exclude clinical trial staff for whom exclusions apply from engaging in the care of trial participants for at least seven days after each GMO administration or any time pustules are present. Children under the age of 12 months are also likely to develop serious adverse reactions should they become infected with VACV. As such, licence conditions include that trial participants would avoid direct physical contact with children under 12 months of age, with recurrent or active skin conditions, and persons who are pregnant or breastfeeding, for 7 days from the time of each treatment with the GMO or any time lesions are present.

214. Although there are natural barriers which limit the frequency and likelihood of poxvirus recombination, there still exists a possibility of the GMO recombining if there is co-infection in the same cell with another poxvirus. This may occur if a participant is vaccinated or treated with another poxvirus shortly before or after to the proposed trial. To limit the risk of this occurrence, a licence condition has been imposed which prevents trial participants from receiving treatment with a poxvirus-based vaccine or treatment for 30 days prior to the trial and 30 days after the last treatment with the GMO. For over 80% of VACV vaccinations, shedding is undetectable after 28 days (Pittman et al., 2015).

215. The applicant proposed that trial participants should refrain from donating blood, organs, sperm and eggs during the clinical trial and for at least 60 days after the last treatment dose. Given the health status of the trial participants, it is unlikely that they would be considered suitable donors' however, there is limited data regarding persistence, biodistribution and shedding of the GMO after treatment. Therefore, a condition has been included in the licence to reflect this.

216. The applicant proposed that trial participants would be instructed to use contraceptives to avoid pregnancy during the clinical trial treatment for at least 90 days after the last treatment dose. As mentioned above vaccinia virus shedding is mostly undetectable 28 days after inoculation. However, given the potential for harm if a pregnancy were impacted by infection with the GMO, a licence condition stipulating the use of barrier contraception for 60 days following the final treatment has been included. There is limited data regarding the shedding of the GMO in body fluids. Therefore, barrier contraception is recommended to avoid exposure to the GMO via shedding in body fluids such as semen and vaginal secretions, in addition to prevention of pregnancy.

217. A standard condition is included in the licence requiring the licence holder to ensure that dealings are conducted so as to ensure containment of the GMO, not compromise the health and safety of people and minimise unintentional exposure to the GMO. A note written under the condition explains that compliance may be achieved by only engaging persons who are required to adhere to appropriate standards to conduct the dealings.

218. Other conditions included in the licence are standard conditions that state that only people authorised by the licence holder are covered by the licence, and that the licence holder must inform all people dealing with the GMOs, other than external service providers, of applicable licence conditions.

219. Further conditions to be implemented in the licence are to ensure that a compliance management plan is in place for each clinical trial site before administration of the GMO commences at that site. The compliance management plan must detail how the licence holder intends to comply with the licence conditions, including listing persons responsible for site management, proposed reporting structures, staff training procedures and transport and disposal processes.

### 3.1.2 Summary of licence conditions to be implemented to limit and control the clinical trial

220. A number of licence conditions have been included to limit and control the proposed clinical trial, based on the above considerations. These include requirements to:

- limit the trial to 40 trial participants, which are to be conducted at clinical trial sites
- restrict access to the GMO
- ensure personnel involved in the trial are appropriately trained and follow appropriate behavioural requirements
- ensure appropriate PPE is used
- restrict personnel permitted to administer the GMO
- requiring decontamination of the GMO and materials and equipment that have been in contact with the GMO at clinical trial sites using effective disinfectants or disposal using a certified waste contractor in accordance with standard clinical waste disposal practices, as required by the relevant Australian and state legislation
- transport and store the GMO and samples from GMO-treated participants in accordance with IATA shipping classification UN 3373 [Category B] and/or the minimum requirements for packaging, and labelling as detailed in the licence.
- clinical waste stream to be used by external service providers to destroy GMO-related waste.

### 3.2 Other risk management considerations

221. All DIR licences issued by the Regulator contain a number of conditions that relate to general risk management. These include conditions relating to:

- applicant suitability
- contingency plans
- identification of the persons or classes of persons covered by the licence
- reporting requirements
- access for the purpose of monitoring for compliance.

### 3.2.1 Applicant suitability

222. In making a decision whether or not to issue a licence, the Regulator must have regard to the suitability of the applicant to hold a licence. Under Section 58 of the Act, matters that the Regulator must take into account include:

- any relevant convictions of the applicant
- any revocation or suspension of a relevant licence or permit held by the applicant under a law of the Commonwealth, a State or a foreign country
- the capacity of the applicant to meet the conditions of the licence.

223. The conditions includes a requirement for the licence holder to inform the Regulator of any information that would affect their suitability.

224. In addition, the applicant organisation must have access to an IBC and be an accredited organisation under the Act.

### 3.2.2 Contingency plans

225. Novotech is required to submit a contingency plan to the Regulator before commencing dealings with the GMO. This plan will detail measures to be undertaken in the event of:

- the unintended release of the GMO, including spills
- exposure of, or transmission to persons other than trial participants
- a person exposed to the GMO developing a serious adverse response.

### 3.2.3 Identification of the persons or classes of persons covered by the licence

226. Persons covered by the licence are the licence holder and employees, agents or contractors of the licence holder and other persons who are, or have been, engaged or otherwise authorised by the licence holder to undertake any activity in connection with the dealings authorised by the licence. Prior to dealings with the GMO, Novotech is required to provide a list of people and organisations that are covered by the licence, or the function or position where names are not known at the time.

### 3.2.4 Reporting requirements

227. The licence requires the licence holder to immediately report any of the following to the Regulator:

- any additional information regarding risks to the health and safety of people or the environment associated with the dealings
- any contraventions of the licence by persons covered by the licence
- any unintended effects of the clinical trial.

228. A number of written notices are also required under the licence regarding dealings with the GMO, to assist the Regulator in designing and implementing a monitoring program for all licensed dealings. The notices include:

- identification of the clinical trial sites where administration of the GMO to trial participants would take place
- expected date of administration with the GMO for each clinical trial site
- cease of administration with the GMO for each clinical trial site.

### 3.2.5 Monitoring for compliance

229. The Act stipulates, as a condition of every licence, that a person who is authorised by the licence to deal with a GMO, and who is required to comply with a condition of the licence, must allow inspectors and other persons authorised by the Regulator to enter premises where a dealing is being undertaken for the purpose of monitoring or auditing the dealing.

230. If monitoring activities identify changes in the risks associated with the authorised dealings, the Regulator may also vary licence conditions, or if necessary, suspend or cancel the licence.

231. In cases of non-compliance with licence conditions, the Regulator may instigate an investigation to determine the nature and extent of non-compliance. The Act provides for criminal sanctions of large fines and/or imprisonment for failing to abide by the legislation, conditions of the licence or directions from the Regulator, especially where significant damage to the health and safety of people or the environment could result.

### Section 4 Issues to be addressed for future releases

232. Additional information has been identified that may be required to assess an application for a commercial release of the GMO, or to justify a reduction in limits and controls. This includes:

• information and data that would address the uncertainties noted in Chapter 2, Section 3. Specifically, information obtained from testing for selective replication competency in cancer cells, tissue tropism and shedding of the GMOs in inoculated trial participants.

### Section 5 Conclusions of the consultation RARMP

233. The risk assessment concludes that the proposed clinical trial of the GMO poses negligible risks to the health and safety of people or the environment as a result of gene technology. These negligible risks do not require specific risk treatment measures.

234. Conditions are imposed to limit the trial to the proposed scale, location and duration, and to restrict the spread and persistence of the GMO and its genetic material in the environment, as these were important considerations in establishing the context for assessing the risks.

### References

Abrahão, J.S., de Souza Trindade, G., Siqueira Ferreira, J.M., Campos, R.K., Bonjardim, C.A., Peregrino Ferreira, P.C., and Kroon, E.G. (2009). Long-lasting stability of *Vaccinia virus* strains in murine feces: implications for virus circulation and environmental maintenance. Archives of Virology *154*, 1551-1553.

Abrahão, J.S., Silva-Fernandes, A.T., Lima, L.S., Campos, R.K., Guedes, M.I.M.C., Cota, M.M.G., Assis, F.L., *et al.* (2010). *Vaccinia virus* infection in monkeys, Brazilian Amazon. Emerging Infectious Diseases *16*, 976-979.

Adams, M.M., Rice, A.D., and Moyer, R.W. (2007). *Rabbitpox virus* and *Vaccinia virus* infection of rabbits as a model for human smallpox. Journal of Virology *81*, 11084-11095.

Águeda-Pinto, A., Kraberger, S., Everts, A., Gutierrez-Jensen, A., Glenn, H.L., Dalton, K.P., Podadera, A., *et al.* (2022). Identification of a Novel Myxoma Virus C7-Like Host Range Factor That Enabled a Species Leap from Rabbits to Hares. mBio *13*, e0346121.

Alakunle, E., Moens, U., Nchinda, G., and Okeke, M.I. (2020). Monkeypox Virus in Nigeria: Infection Biology, Epidemiology, and Evolution. Viruses *12*, 1257.

Altindis, M., Puca, E., and Shapo, L. (2022). Diagnosis of monkeypox virus – An overview. Travel Medicine and Infectious Disease *50*, 102459.

Artois, M., Charlton, K.M., Tolson, N.D., Casey, G.A., Knowles, M.K., and Campbell, J.B. (1990). Vaccinia recombinant virus expressing the rabies virus glycoprotein: safety and efficacy trials in Canadian wildlife. Canadian Journal of Veterinary Research *54*, 504-507.

Australian Government Department of Health (2024). National Notifiable Disease Surveillance System. Accessed: 19 November. <u>https://nindss.health.gov.au/pbi-dashboard/</u>.

Australian Technical Advisory Group on Immunisation (ATAGI) (2018). Australian Immunisation Handbook, Vol 1, 10 edn (Australian Government Department of Health, Canberra, Australia: Australian Government National Health and Medical Research Council).

Australian Technical Advisory Group on Immunisation (ATAGI) (2024). Australian Immunisation Handbook (Canberra: Australian Government Department of Health and Aged Care).

Badell, M.L., Meaney-Delman, D., Tuuli, M.G., Rasmussen, S.A., Petersen, B.W., Sheffield, J.S., Beigi, R.H., *et al.* (2015). Risks Associated With Smallpox Vaccination in Pregnancy: A Systematic Review and Metaanalysis. Obstet Gynecol *125*, 1439-1451.

Baldick, C.J., Jr., and Moss, B. (1993). Characterization and temporal regulation of mRNAs encoded by vaccinia virus intermediate-stage genes. J Virol *67*, 3515-3527.

Bennett, M., Gaskell, R.M., Gaskell, C.J., Baxby, D., and Kelly, D.F. (1989). Studies on poxvirus infection in cats. Archives of Virology *104*, 19-33.

Biohazard Waste Industry (2010). Industry Code of Practice for the Management of Clinical and Related Wastes, 6 edn.

Bratke, K.A., and McLysaght, A. (2008). Identification of multiple independent horizontal gene transfers into poxviruses using a comparative genomics approach. BMC Evolutionary Biology *8*, 67.

Brennan, G., Stoian, A.M.M., Yu, H., Rahman, M.J., Banerjee, S., Stroup, J.N., Park, C., et al. (2023). Molecular Mechanisms of Poxvirus Evolution. mBio 14, e0152622. Brochier, B., Blancou, J., Thomas, I., Languet, B., Artois, M., Kieny, M.P., Lecocq, J.P., *et al.* (1989). Use of recombinant vaccinia-rabies glycoprotein virus for oral vaccination of wildlife against rabies: innocuity to several non-target bait consuming species. Journal of Wildlife Diseases *25*, 540-547.

Buller, R.M., and Palumbo, G.J. (1991). Poxvirus pathogenesis. Microbiol Rev 55, 80-122.

Center for Disease Control and Prevention (2024). Guidelines for Vaccinating Pregnant Persons. (United States of America government) Accessed: 20/11/2024. <u>https://www.cdc.gov/vaccines-pregnancy/hcp/vaccination-guidelines/index.html</u>.

Centers for Disease Control and Prevention (2020). Medical Management of Adverse Reactions to Vaccinia Vaccination. (available online <u>https://www.cdc.gov/smallpox/clinicians/vaccine-medical-management6.html</u>) Accessed: 03/11/2020. <u>https://www.cdc.gov/smallpox/clinicians/vaccine-medical-management6.html</u>.

Chahroudi, A., Chavan, R., Koyzr, N., Waller, E.K., Silvestri, G., and Feinberg, M.B. (2005). *Vaccinia virus* tropism for primary hematolymphoid cells is determined by restricted expression of a unique virus receptor. Journal of Virology *79*, 10397-10407.

Chambers, A.E., Dixon, M.M., and Harvey, S.P. (2009). Studies of the suitability of fowlpox as a decontamination and thermal stability simulant for variola major. International Journal of Microbiology *2009*, 158749.

Chaurasiya, S., Fong, Y., and Warner, S.G. (2020). Optimizing oncolytic viral design to enhance antitumor efficacy: Progress and challenges. Cancers *12*, 1699.

Chidwick, K., Busingye, D., Pollack, A., Osman, R., Yoo, J., Blogg, S., Rubel, D., *et al.* (2020). Prevalence, incidence and management of atopic dermatitis in Australian general practice using routinely collected data from MedicineInsight. Australasian Journal of Dermatology *61*, e319-e327.

Cohen, J. (2001). Smallpox vaccinations: How much protection remains? Science 294, 985-985.

Cono, J., Casey, C.G., and Bell, D.M. (2003). Smallpox vaccination and adverse reactions. Guidance for clinicians. MMWR Recommendations and Reports *52*, 1-28.

Cooney, E.L., Collier, A.C., Greenberg, P.D., Coombs, R.W., Zarling, J., Arditti, D.E., Hoffman, M.C., *et al.* (1991). Safety of and immunological response to a recombinant *Vaccinia virus* vaccine expressing HIV envelope glycoprotein. The Lancet *337*, 567-572.

Cummings, J.F., Polhemus, M.E., Hawkes, C., Klote, M., Ludwig, G.V., and Wortmann, G. (2008). Persistence of vaccinia at the site of smallpox vaccination. Clinical Infectious Diseases *46*, 101-102.

Damaso, C.R.A., Esposito, J.J., Condit, R.C., and Moussatche, N. (2000). An emergent poxvirus from humans and cattle in Rio de Janeiro State: *Cantagalo Virus* may derive from Brazilian Smallpox vaccine. Virology *277*, 439-449.

De Clercq, E. (2002). Cidofovir in the treatment of poxvirus infections. Antiviral Res 55, 1-13.

Downie, A.W. (1939). A study of the lesions produced experimentally by cowpox virus. The Journal of Pathology and Bacteriology *48*, 361-379.

Dumbell, K., and Richardson, M. (1993). Virological investigations of specimens from buffaloes affected by buffalopox in Maharashtra State, India between 1985 and 1987. Archives of Virology *128*, 257-267.

Dyer, A., Baugh, R., Chia, S.L., Frost, S., Iris, Jacobus, E.J., Khalique, H., *et al.* (2019). Turning cold tumours hot: oncolytic virotherapy gets up close and personal with other therapeutics at the 11th Oncolytic Virus Conference. Cancer Gene Therapy *26*, 59-73.

Enserink, M. (2002). Treating vaccine reactions: Two lifelines, but no guarantees. Science 298, 2313-2313.

Esparza, J., Schrick, L., Damaso, C.R., and Nitsche, A. (2017). Equination (inoculation of horsepox): An early alternative to vaccination (inoculation of cowpox) and the potential role of horsepox virus in the origin of the smallpox vaccine. Vaccine *35*, 7222-7230.

Espy, M.J., Uhl, J.R., Sloan, L.M., Rosenblatt, J.E., Cockerill, F.R., and Smith, T.F. (2002). Detection of vaccinia virus, herpes simplex virus, varicella-zoster virus, and *Bacillus anthracis* DNA by LightCycler polymerase chain reaction after autoclaving: Implications for biosafety of bioterrorism agents. Mayo Clinic Proceedings 77, 624-628.

Essbauer, S., Meyer, H., Porsch-Ozcurumez, M., and Pfeffer, M. (2007). Long-lasting stability of *Vaccinia virus* (orthopoxvirus) in food and environmental samples. Zoonoses and Public Health *54*, 118-124.

Falkner, F.G., and Moss, B. (1990). Transient dominant selection of recombinant vaccinia viruses. J Virol 64, 3108-3111.

Farahat, R.A., Ali, I., Al- Ahdal, T., Benmelouka, A.Y., Albakri, K., El-Sakka, A.A., Abdelaal, A., *et al.* (2022). Monkeypox and human transmission: Are we on the verge of another pandemic? Travel Medicine and Infectious Disease *49*, 102387.

Felipetto Cargnelutti, J., Schmidt, C., Masuda, E.K., Braum, L.D., Weiblen, R., and Furtado Flores, E. (2012). *Vaccinia viruses* isolated from cutaneous disease in horses are highly virulent for rabbits. Microbial Pathogenesis *52*, 192-199.

Ferreira, J.M., Abrahao, J.S., Drumond, B.P., Oliveira, F.M., Alves, P.A., Pascoal-Xavier, M.A., Lobato, Z.I., *et al.* (2008). *Vaccinia virus*: shedding and horizontal transmission in a murine model. J Gen Virol *89*, 2986-2991.

Ferrucci, P.F., Pala, L., Conforti, F., and Cocorocchio, E. (2021). Talimogene Laherparepvec (T-VEC): An Intralesional Cancer Immunotherapy for Advanced Melanoma. Cancers (Basel) *13*.

Fulginiti, V.A., Papier, A., Lane, J.M., Neff, J.M., and Henderson, D.A. (2003a). Smallpox vaccination: A review, part I. Background, vaccination technique, normal vaccination and revaccination, and expected normal reactions. Clinical Infectious Diseases *37*, 241-250.

Fulginiti, V.A., Papier, A., Lane, J.M., Neff, J.M., and Henderson, D.A. (2003b). Smallpox vaccination: A review, part II. Adverse events. Clinical Infectious Diseases *37*, 251-271.

Greseth, M.D., and Traktman, P. (2022). The Life Cycle of the Vaccinia Virus Genome. Annu Rev Virol 9, 239-259.

Guo, Z.S., Lu, B., Guo, Z., Giehl, E., Feist, M., Dai, E., Liu, W., *et al.* (2019). Vaccinia virus-mediated cancer immunotherapy: cancer vaccines and oncolytics. Journal for ImmunoTherapy of Cancer 7, 6.

Hammerschlag, Y., MacLeod, G., Papadakis, G., Adan Sanchez, A., Druce, J., Taiaroa, G., Savic, I., *et al.* (2022). Monkeypox infection presenting as genital rash, Australia, May 2022. Euro Surveill *27*.

Harrington, K., Freeman, D.J., Kelly, B., Harper, J., and Soria, J.-C. (2019). Optimizing oncolytic virotherapy in cancer treatment. Nature Reviews Drug Discovery *18*, 689-706.

Hatakeyama, S., Moriya, K., Saijo, M., Morisawa, Y., Kurane, I., Koike, K., Kimura, S., *et al.* (2005). Persisting humoral antiviral immunity within the Japanese population after the discontinuation in 1976 of routine smallpox vaccinations. Clinical and Diagnostic Laboratory Immunology *12*, 520-524.

Healthdirect Australia (2024). Molluscum contagiosum. Accessed: 19 November. https://www.healthdirect.gov.au/molluscum-contagiosum.

Healthdirect Australia (2024). Mpox (Monkeypox). (Australian Government) Accessed: 06 November. <u>https://www.healthdirect.gov.au/monkeypox</u>.

Heo, J., Reid, T., Ruo, L., Breitbach, C.J., Rose, S., Bloomston, M., Cho, M., *et al.* (2013). Randomized dose-finding clinical trial of oncolytic immunotherapeutic vaccinia JX-594 in liver cancer. Nature Medicine *19*, 329-336.

Horsington, J., Lynn, H., Turnbull, L., Cheng, D., Braet, F., Diefenbach, R.J., Whitchurch, C.B., *et al.* (2013). A36-dependent Actin Filament Nucleation Promotes Release of Vaccinia Virus. PLOS Pathogens *9*, e1003239.

Howell, L.M., Gracie, N.P., and Newsome, T.P. (2024). Single-cell analysis of VACV infection reveals pathogen-driven timing of early and late phases and host-limited dynamics of virus production. PLOS Pathogens *20*, e1012423.

ICH (2016). Integrated addendum to ICH E6(R1): Guideline for good clinical practice. Report No. E6(R2).

Jacobs, B.L., Langland, J.O., Kibler, K.V., Denzler, K.L., White, S.D., Holechek, S.A., Wong, S., et al. (2009). *Vaccinia virus* vaccines: past, present and future. Antiviral Research *84*, 1-13.

Ježek, Z., Szczeniowski, M., Paluku, K.M., and Mutombo, M. (1987). Human Monkeypox: Clinical Features of 282 Patients. The Journal of Infectious Diseases *156*, 293-298.

Johnson, D.B., Puzanov, I., and Kelley, M.C. (2015). Talimogene laherparepvec (T-VEC) for the treatment of advanced melanoma. Immunotherapy 7, 611-619.

José da Silva Domingos, I., Silva de Oliveira, J., Lorene Soares Rocha, K., Bretas de Oliveira, D., Geessien Kroon, E., Barbosa Costa, G., and de Souza Trindade, G. (2021). Twenty Years after Bovine Vaccinia in Brazil: Where We Are and Where Are We Going? Pathogens *10*.

Katsafanas, G.C., and Moss, B. (2007). Colocalization of transcription and translation within cytoplasmic poxvirus factories coordinates viral expression and subjugates host functions. Cell Host & Microbe 2, 221-228.

Kerr, P.J., Cattadori, I.M., Sim, D., Liu, J., Holmes, E.C., and Read, A.F. (2022). Divergent Evolutionary Pathways of Myxoma Virus in Australia: Virulence Phenotypes in Susceptible and Partially Resistant Rabbits Indicate Possible Selection for Transmissibility. J Virol *96*, e0088622.

Konya, J., and Thompson, C.H. (1999). *Molluscum contagiosum virus*: antibody responses in persons with clinical lesions and seroepidemiology in a representative Australian population. The Journal of Infectious Diseases *179*, 701-704.

Kretzschmar, M., Wallinga, J., Teunis, P., Xing, S., and Mikolajczyk, R. (2006). Frequency of adverse events after vaccination with different *Vaccinia* strains. PLOS Medicine *3*, e272.

Laliberte, J.P., Weisberg, A.S., and Moss, B. (2011). The membrane fusion step of vaccinia virus entry is cooperatively mediated by multiple viral proteins and host cell components. PLoS Pathog 7, e1002446.

Lane, J.M., and Fulginiti, V.A. (2003). Transmission of *Vaccinia virus* and rationale for measures for prevention. Clinical Infectious Diseases *37*, 281-284.

Lane, J.M., and Goldstein, J. (2003a). Adverse events occurring after smallpox vaccination. Seminars in Pediatric Infectious Diseases *14*, 189-195.

Lane, J.M., and Goldstein, J. (2003b). Evaluation of 21st-century risks of smallpox vaccination and policy options. Annals of Internal Medicine *138*, 488-493.

Lauer, U.M., Schell, M., Beil, J., Berchtold, S., Koppenhöfer, U., Glatzle, J., Königsrainer, A., *et al.* (2018). Phase I Study of Oncolytic Vaccinia Virus GL-ONC1 in Patients with Peritoneal Carcinomatosis. Clin Cancer Res *24*, 4388-4398.

Li, G., Chen, N., Feng, Z., Buller, R.M., Osborne, J., Harms, T., Damon, I., *et al.* (2006). Genomic sequence and analysis of a vaccinia virus isolate from a patient with a smallpox vaccine-related complication. Virol J *3*, 88.

Lin, Y.-C.J., and Evans, D.H. (2010). *Vaccinia virus* particles mix inefficiently, and in a way that would restrict viral recombination, in coinfected cells. Journal of Virology *84*, 2432-2443.

Liu, L., Cooper, T., Howley, P.M., and Hayball, J.D. (2014). From crescent to mature virion: vaccinia virus assembly and maturation. Viruses *6*, 3787-3808.

Maruri-Avidal, L., Weisberg, A.S., and Moss, B. (2013). Direct formation of vaccinia virus membranes from the endoplasmic reticulum in the absence of the newly characterized L2-interacting protein A30.5. J Virol *87*, 12313-12326.

Matos, A.C.D., Rehfeld, I.S., Guedes, M., and Lobato, Z.I.P. (2018). Bovine Vaccinia: Insights into the Disease in Cattle. Viruses *10*.

Maurer, D.M., Harrington, B., and Lane, J.M. (2003). Smallpox vaccine: contraindications, administration, and adverse reactions. American Family Physician *68*, 889-896.

McLysaght, A., Baldi, P.F., and Gaut, B.S. (2003). Extensive gene gain associated with adaptive evolution of poxviruses. Proc Natl Acad Sci U S A *100*, 15655-15660.

Medeiros-Silva, D.C., dos Santos Moreira-Silva, E.A., de Assis Silva Gomes, J., da Fonseca, F.G., and Correa-Oliveira, R. (2010). Clinical signs, diagnosis, and case reports of Vaccinia virus infections. The Brazilian Journal of Infectious Diseases *14*, 129-134.

Mercer, J., Knébel, S., Schmidt, F.I., Crouse, J., Burkard, C., and Helenius, A. (2010). Vaccinia virus strains use distinct forms of macropinocytosis for host-cell entry. Proc Natl Acad Sci U S A *107*, 9346-9351.

Middaugh, N., Petersen, B., McCollum, A.M., and Smelser, C. (2016). Notes from the field: Adverse reaction after *Vaccinia virus* vaccination - New Mexico, 2016. Morbidity and Mortality Weekly Report *65*, 1351-1352.

Miller, J.D., van der Most, R.G., Akondy, R.S., Glidewell, J.T., Albott, S., Masopust, D., Murali-Krishna, K., *et al.* (2008). Human Effector and Memory CD8<sup>+</sup> T Cell Responses to&#xa0;Smallpox and Yellow Fever Vaccines. Immunity *28*, 710-722.

Miranda, J.B., Borges, I.A., Campos, S.P.S., Vieira, F.N., de Azara, T.M.F., Marques, F.A., Costa, G.B., *et al.* (2017). Serologic and molecular evidence of *Vaccinia virus* circulation among small mammals from different biomes, Brazil. Emerging Infectious Diseases *23*, 931-938.

Nagata, L.P., Irwin, C.R., Hu, W.G., and Evans, D.H. (2018). Vaccinia-based vaccines to biothreat and emerging viruses. Biotechnology and Genetic Engineering Reviews *34*, 107-121.

National Health and Medical Research Council (2019). Australian Guidelines for the Prevention and Control of Infection in Healthcare. (Canberra: Commonwealth of Australia, available online <a href="https://www.nhmrc.gov.au/about-us/publications/australian-guidelines-prevention-and-control-infection-healthcare-2019#block-views-block-file-attachments-content-block-1">https://www.nhmrc.gov.au/about-us/publications/australian-guidelines-prevention-and-control-infection-healthcare-2019#block-views-block-file-attachments-content-block-1</a>).

National Health and Medical Research Council, Australian Research Council, and Universities Australia (2023). National Statement on Ethical Conduct in Human Research 2023. (Canberra).

Neff, J.M., Lane, J.M., Fulginiti, V.A., and Henderson, D.A. (2002). Contact vaccinia--transmission of vaccinia from smallpox vaccination. JAMA 288, 1901-1905.

OGTR (2013). Risk Analysis Framework 2013, 4th edn (Canberra: Office of the Gene Technology Regulator, available online: <u>https://www.ogtr.gov.au/resources/publications/risk-analysis-framework-2013</u>).

Oliveira, G., Assis, F., Almeida, G., Albarnaz, J., Lima, M., Andrade, A.C., Calixto, R., *et al.* (2015). From lesions to viral clones: biological and molecular diversity amongst autochthonous Brazilian *Vaccinia virus*. Viruses *7*, 1218-1237.

Oliveira, G.P., Rodrigues, R.A.L., Lima, M.T., Drumond, B.P., and Abrahão, J.S. (2017a). Poxvirus Host Range Genes and Virus-Host Spectrum: A Critical Review. Viruses *9*.

Oliveira, G.P., Rodrigues, R.A.L., Lima, M.T., Drumond, B.P., and Abrahão, J.S. (2017b). Poxvirus host range genes and virus-host spectrum: A critical review. Viruses *9*, 331.

Oliveira, G.P., Silva-Fernandes, A.T., Assis, F.L., Alves, P.A., Franco-Luiz, A.P.M., Figueiredo, L.B., de Almeida, C.M.C., *et al.* (2014). Intrafamilial transmission of *Vaccinia virus* during a bovine vaccinia outbreak in Brazil: a new insight in viral transmission chain. American Journal of Tropical Medicine and Hygeine *90*, 1021-1023.

Oliveira, J.S.d., Figueiredo, P.D.O., Costa, G.B., Assis, F.L.d., Drumond, B.P., Da Fonseca, F.G., Nogueira, M.L., *et al.* (2017c). Vaccinia Virus Natural Infections in Brazil: The Good, the Bad, and the Ugly. Viruses *9*, 340.

Park, S.H., Breitbach, C.J., Lee, J., Park, J.O., Lim, H.Y., Kang, W.K., Moon, A., *et al.* (2015). Phase 1b trial of biweekly intravenous Pexa-Vec (JX-594), an oncolytic and immunotherapeutic *Vaccinia virus* in colorectal cancer. Molecular Therapy *23*, 1532-1540.

Paszkowski, P., Noyce, R.S., and Evans, D.H. (2016). Live-cell imaging of *Vaccinia virus* recombination. PLOS Pathogens *12*, e1005824.

Payne, L.G., and Kristenson, K. (1979). Mechanism of vaccinia virus release and its specific inhibition by N1-isonicotinoyl-N2-3-methyl-4-chlorobenzoylhydrazine. J Virol *32*, 614-622.

Pittman, P.R., Garman, P.M., Kim, S.-H., Schmader, T.J., Nieding, W.J., Pike, J.G., Knight, R., *et al.* (2015). Smallpox vaccine, ACAM2000: Sites and duration of viral shedding and effect of povidone iodine on scarification site shedding and immune response. Vaccine *33*, 2990-2996.

Public Health Agency of Canada (2011). Pathogen Safety Data Sheets: Infectious Substances – Vaccinia Virus. (available online, <u>https://www.canada.ca/en/public-health/services/laboratory-biosafety-biosecurity/pathogen-safety-data-sheets-risk-assessment/vaccinia-virus.html</u>) Accessed: 19/11/19. <u>https://www.canada.ca/en/public-health/services/laboratory-biosafety-biosecurity/pathogen-safety-data-sheets-risk-assessment/vaccinia-virus.html</u>.

Qin, L., Favis, N., Famulski, J., and Evans David, H. (2015). Evolution of and Evolutionary Relationships between Extant Vaccinia Virus Strains. Journal of Virology *89*, 1809-1824.

Reed, J.L., Scott, D.E., and Bray, M. (2012). Eczema Vaccinatum. Clinical Infectious Diseases 54, 832-840.

Rheinbaben, F.v., Gebel, J., Exner, M., and Schmidt, A. (2007). Environmental resistance, disinfection, and sterilization of poxviruses. In Poxviruses, A.A. Mercer, A. Schmidt, and O. Weber, eds. (Basel: Birkhäuser Basel), pp. 397-405.

Riyesh, T., Karuppusamy, S., Bera, B.C., Barua, S., Virmani, N., Yadav, S., Vaid, R.K., *et al.* (2014). Laboratoryacquired buffalopox virus infection, India. Emerging Infectious Diseases *20*, 324-326.

Robinson, A.J., and Mercer, A.A. (1988). Orf virus and vaccinia virus do not cross-protect sheep. Archives of Virology *101*, 255-259.

Sagripanti, J.L., and Lytle, C.D. (2011). Sensitivity to ultraviolet radiation of Lassa, vaccinia, and Ebola viruses dried on surfaces. Archives of Virology *156*, 489-494.

Sarker, S., Hannon, C., Athukorala, A., and Bielefeldt-Ohmann, H. (2021). Emergence of a Novel Pathogenic Poxvirus Infection in the Endangered Green Sea Turtle (Chelonia mydas) Highlights a Key Threatening Process. Viruses *13*.

Sauerbrei, A., and Wutzler, P. (2009). Testing thermal resistance of viruses. Archives of Virology 154, 115-119.

Schmidt, Florian I., Bleck, Christopher Karl E., Reh, L., Novy, K., Wollscheid, B., Helenius, A., Stahlberg, H., *et al.* (2013). Vaccinia Virus Entry Is Followed by Core Activation and Proteasome-Mediated Release of the Immunomodulatory Effector VH1 from Lateral Bodies. Cell Reports *4*, 464-476.

Schramm, B., and Locker, J.K. (2005). Cytoplasmic organization of poxvirus DNA replication. Traffic *6*, 839-846.

Senkevich, T.G., Koonin, E.V., Bugert, J.J., Darai, G., and Moss, B. (1997). The genome of molluscum contagiosum virus: analysis and comparison with other poxviruses 1. Virology *233*, 19-42.

Sepkowitz, K.A. (2003). How contagious is vaccinia? New England Journal of Medicine 348, 439-446.

Shors, T., Keck, J.G., and Moss, B. (1999). Down regulation of gene expression by the *Vaccinia virus* D10 protein. Journal of Virology *73*, 791-796.

Sprygin, A., Mazloum, A., van Schalkwyk, A., and Babiuk, S. (2022). Capripoxviruses, leporipoxviruses, and orthopoxviruses: Occurrences of recombination. Frontiers in Microbiology 13.

Standards Australia/New Zealand (2010). Safety in laboratories Part 3: Microbiological safety and containment AS/NZS 2243.3:2010, 6 edn (SAI Global Limited).

Standards Australia/New Zealand (2022). AS/NZS 2243.3:2022 Safety in laboratories, Part 3: Microbiological safety and containment.

Stark, J.H., Frey, S.E., Blum, P.S., and Monath, T.P. (2006). Lack of transmission of vaccinia virus. Emerging Infectious Diseases *12*, 698-700.

Tack, D.M., Karem, K.L., Montgomery, J.R., Collins, L., Bryant-Genevier, M.G., Tiernan, R., Cano, M., *et al.* (2013). Unintentional transfer of vaccinia virus associated with smallpox vaccines. Human Vaccines & Immunotherapeutics *9*, 1489-1496.

Tulman, E.R., Delhon, G., Afonso, C.L., Lu, Z., Zsak, L., Sandybaev, N.T., Kerembekova, U.Z., *et al.* (2006). Genome of Horsepox Virus. Journal of Virology *80*, 9244-9258.

Vallée, G., Norris, P., Paszkowski, P., Noyce, R.S., and Evans, D.H. (2021). Vaccinia Virus Gene Acquisition through Nonhomologous Recombination. Journal of Virology *95*, 10.1128/jvi.00318-00321.

Vinay, D.S., Ryan, E.P., Pawelec, G., Talib, W.H., Stagg, J., Elkord, E., Lichtor, T., *et al.* (2015). Immune evasion in cancer: Mechanistic basis and therapeutic strategies. Seminars in Cancer Biology *35*, S185-S198.

Webber, B.J., Montgomery, J.R., Markelz, A.E., Allen, K.C., Hunninghake, J.C., Ritchie, S.A., Pawlak, M.T., *et al.* (2014). Spread of vaccinia virus through shaving during military training, Joint Base San Antonio-Lackland, TX, June 2014. Medical Surveillance Monthly Report *21*, 2-6.

Wertheimer, E.R., Olive, D.S., Brundage, J.F., and Clark, L.L. (2012). Contact transmission of *Vaccinia virus* from smallpox vaccinees in the United States, 2003-2011. Vaccine *30*, 985-988.

Wharton, M., Strikas, R.A., Harpaz, R., Rotz, L.D., Schwartz, B., Casey, C.G., Pearson, M.L., *et al.* (2003). Recommendations for using smallpox vaccine in a pre-event vaccination program. Supplemental recommendations of the Advisory Committee on Immunization Practices (ACIP) and the Healthcare Infection Control Practices Advisory Committee (HICPAC). MMWR Recommendations and Reports *52*, 1-16.

Wildlife Health Australia (2019). Poxviruses and Australian mammals. Fact Sheet October 2019 2.1.

Wittek, R. (2006). Vaccinia immune globulin: current policies, preparedness, and product safety and efficacy. International Journal of Infectious Diseases *10*, 193-201.

Wood, J.P., Choi, Y.W., Wendling, M.Q., Rogers, J.V., and Chappie, D.J. (2013). Environmental persistence of *Vaccinia virus* on materials. Letters in Applied Microbiology *57*, 399-404.

Yang, Z., Reynolds, S.E., Martens, C.A., Bruno, D.P., Porcella, S.F., and Moss, B. (2011). Expression profiling of the intermediate and late stages of poxvirus replication. Journal of Virology *85*, 9899-9908.

Yeh, T.-Y., Hsieh, Z.-Y., Feehley, M.C., Feehley, P.J., Contreras, G.P., Su, Y.-C., Hsieh, S.-L., *et al.* (2022). Recombination shapes the 2022 monkeypox (mpox) outbreak. Med *3*, 824-826.

Zeh, H.J., Downs-Canner, S., McCart, J.A., Guo, Z.S., Rao, U.N.M., Ramalingam, L., Thorne, S.H., *et al.* (2015). First-in-man study of western reserve strain oncolytic *Vaccinia virus*: Safety, systemic spread, and antitumor activity. Molecular Therapy *23*, 202-214.

Zhang, Q., and Liu, F. (2020). Advances and potential pitfalls of oncolytic viruses expressing immunomodulatory transgene therapy for malignant gliomas. Cell Death & Disease *11*, 485.

# Appendix A: Summary of submissions from prescribed experts, agencies and authorities on the consultation RARMP

Submissions	Summary of issues raised	Comment	
1	Agrees that the proposed clinical trial poses negligible risk to human health and safety and the environment.	Noted.	
	Acknowledged considerations addressed in the RARMP and noted that:	Noted.	
	<ul> <li>remaining uncertainty considered in Section 3 of Chapter 2 in the RARMP and considered that given the proposed licence conditions the level of uncertainty in this risk assessment is low and does not impact on the overall estimate of risk.</li> </ul>		
	<ul> <li>the limits and controls outlined in the RARMP for this clinical trial are appropriate and the trial represents negligible risk to human health and the environment.</li> </ul>	Noted.	
2	Has no concerns regarding the trial and noted that:	Noted.	
	<ul> <li>the proposed trial poses no significant risk to the population, and. it is a very small scale trial in Australia.</li> </ul>		
	<ul> <li>this is an innovative and potentially valuable trial and cancer treatment that is well worth trying.</li> </ul>		
3	Noted that if the trial is conducted in the State of Western Australia, the applicant is required to obtain a Prohibited Matter Permit from DPIRD.	This information was included in the Summary of the RAMRP but has been added to Section 1.1 of the RARMP <i>"Interface with other regulatory agencies" for clarity</i> .	
	Considered that the risks scenarios in the RARMP seemed appropriate and well measured and agreed with the overall conclusion of the RARMP that the proposed trial poses negligible risk of harm to human health and safety and the environment	Noted.	
4	Raised concerns regarding the delivery of the treatment communities with limited medical capacity and support. Considered that this issue would likely be considered by the local Council and the state Health Department.	Licence conditions require the GMO to be administered within a clinical trial site. As discussed in Section 1.1 of the RARMP, medical facilities regulated by State and Territory governments and are considered suitable for the proposed dealings.	
5	Agreed that the proposed trial poses negligible risks the health and safety of people and the environment.	A condition has been added to licence (LC29(d)) requiring the licence holder must obtain written agreement from the	
	Noted that the current PCR testing used to identify mpox infections is likely to cross-react with the	trial participant that they will inform the medical practitioner of their participation in the trial if seeking medical attention	

Submissions	Summary of issues raised	Comment
	GMO and trial participants and/close contacts exposed to the GMO suspected of mpox infection would have inconclusive test results. Suggest that applicants should be required to inform trial participants and close household contacts of this so that they can inform their physician or pathology laboratory of their participation in the trial if they are also testing for mpox.	outside of the clinical trial In addition, a condition requires the licence holder to provide medical attention to any person exposed to the GMO (including close contacts) and to provide information about the GMO to the attending medical practitioner
6	<ul> <li>Recommended that the Regulator should consider:</li> <li>rewording text around selective replication in tumours</li> </ul>	The wording throughout the ramp has been modified to reflect the likelihood that some replication of the GMO may occur in healthy cells, although this is likely to be less efficient than the replication in tumour cells.
	<ul> <li>broadening controls associated with exposure to seminal and vaginal fluids.</li> </ul>	Risk scenario 2 has been amended for clarity. In addition, a note has been added to LC29 specifying that the barrier is intended to minimise skin contact with pustules as well as bodily fluids.
	<ul> <li>risks to lab staff handling biospecimens from the participants</li> </ul>	This risk scenario considers potential harm to people conducting dealings with the GMO. It has been amended to include collection of samples as a pathway to exposure, in addition to personnel preparing and administering the GMO as well as those analysing samples. In addition, paragraph 147 of the RARMP has been amended to clarify that biological samples may also be collected or analysed at pathology laboratories. These facilities adhere to national standards for handling of infectious substances and are considered sufficient to prevent the personnel exposure to the GMO.
	Agreed with the overall conclusion of the RARMP	Noted.

1

The exact origin of VACV is unknown and the

Appendix B: Summary of submissions from the public on the				
consultation RARMP				
Submission	Summary of issues raised	Comment		

Raised concerns regarding the origin of the

VACV and the strain used to generate the GMO as it is not detailed in the RARMP	specific strain of to generate the GM VACV being used in the trial has been declared as Commercial Confidential Information which the OGTR cannot disclose to the public. This information was available to the relevant agencies and experts who were consulted on the RARMP.
The OGTR should consider risks of systemic infection of pathogens other than the GMO to trial participants as a result of i.v infusion	The GMO will be manufactured under Good Manufacturing Practice (GMP) conditions, which involves quality control testing. Additionally, as discussed in Chapter 1, Section 1.1, patient safety and the quality and efficacy of the treatment with the GMO are regulated under the <i>Therapeutic Goods</i> <i>Act 1989.</i> Clinical trials of therapeutic products that are experimental and under development are regulated by the Therapeutic Goods Administration (TGA) through the Clinical Trial Approval (CTA) scheme or the Clinical Trial Notification (CTN) scheme. Approval by a Human Research Ethics Committee (HREC) is also required prior to conducting the trial.
Considered that control measures such as PPE and the exclusion of high-risk personnel does not eliminate or minimise risk due to the unknown minimum infectious dose of the VACV.	Work practices and PPE included in the licence conditions aim to minimise the potential for exposure of personnel to the GMO. The risk of harm to personnel conducting dealings as a result of exposure to the GMO is discussed in Risk Scenario 1 of the RARMP and considered as negligible.
Considered that the Australian Guidelines for the Prevention and Control of Infection in Healthcare (2024) are not appropriate for clinical trials because they are developed for an acute care setting. Suggested that the OGTR should consider further limitations to the clinical trial.	The Australian Guidelines for the Prevention and Control of Infection in Healthcare provide evidence-based recommendations that minimise the risk of transmission of infectious agents in the healthcare setting. These measures are considered suitable to minimise risks of personnel exposure to the GMOs within clinical trial site. In addition, licence conditions are imposed to ensure that appropriate training and use of PPE. As mentioned above, the risk of harm to personnel conducting dealings as a result of exposure to the GMO is discussed in Risk Scenario 1 of the RARMP and considered as negligible.