

March 2025

# Risk Assessment and Risk Management Plan (consultation version) for

# **DIR 215**

# Limited and controlled release of canola genetically modified for dairy protein production

Applicant: Miruku Australia Pty Ltd

# This RARMP is open for consultation until 13 May 2025.

Written comments on the risks to human health and safety and the environment posed by this proposed release are invited. You may make your submission

via the consultation hub: https://consultations.health.gov.au/ (search for DIR 215)

via email to: <a href="mailto:ogtr@health.gov.au">ogtr@health.gov.au</a>

or via mail to: The Office of the Gene Technology Regulator

MDP 54, GPO Box 9848, Canberra ACT 2601.

Please note that issues regarding food safety and labelling, the use of agricultural chemicals, and marketing and trade implications do **not** fall within the scope of these evaluations as they are the responsibilities of other agencies and authorities.

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# Summary of the Risk Assessment and Risk Management Plan (Consultation Version)

# for

# **Licence Application No. DIR 215**

#### Introduction

The Gene Technology Regulator (the Regulator) has received a licence application for the intentional release of a genetically modified organism (GMO) into the environment. It qualifies as a limited and controlled release application under the *Gene Technology Act 2000* (the Act). The Regulator has prepared a draft Risk Assessment and Risk Management Plan (RARMP) for this application, which concludes that the proposed field trial poses negligible risks to the health and safety of people and the environment. Licence conditions have been drafted for the proposed field trial. The Regulator invites submissions on the RARMP, including draft licence conditions, to inform the decision on whether or not to issue a licence.

# The application

Applicant	Miruku Australia Pty Ltd (Miruku)
Project title	Limited and controlled release of canola genetically modified for dairy protein production <sup>1</sup>
Parent organism	Canola (Brassica napus L.)
Introduced genes	Introduced gene <sup>2</sup> producing dairy protein:
	<ul> <li>modified β-casein gene based on the gene from cattle (Bos taurus) for dairy protein production</li> </ul>
	Introduced marker gene:
	bar gene from bacterium Streptomyces hygroscopicus for tolerance to the herbicide glufosinate
Genetic modification method	Agrobacterium-mediated transformation
Number of lines	Up to 50 lines
Previous releases	None in Australia or overseas
Proposed locations	Up to 2 sites per year in 2025, 5 in 2026, 10 in 2027, 15 in 2028 and 20 in 2029. Sites to be selected from 135 possible local government areas in New South Wales, Victoria, Western Australia and South Australia
Proposed release size	Up to 1 ha in 2025, 5 ha in 2026, 25 ha in 2027, 105 ha in 2028, and 300 ha in 2029, with a maximum of 436 ha over the period of release
Proposed period of release	From May 2025 until December 2029
Principal purpose	To produce dairy protein in GM canola under field conditions

<sup>&</sup>lt;sup>1</sup> The title of the project as supplied by the applicant is "Limited and controlled release of Canola genetically modified for dairy protein and fat composition".

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<sup>&</sup>lt;sup>2</sup> Confidential Commercial Information (CCI): Some details about the introduced genetic elements in GM canola have been declared as CCI under section 185 of the Act. This information will be made available to the prescribed experts and agencies that will be consulted on this application. CCI is not available to the public.

#### Risk assessment

The risk assessment process considers how the genetic modification and proposed activities conducted with the GMOs might lead to harm to people or the environment. Risks are characterised in relation to both the seriousness and likelihood of harm, taking into account current scientific/technical knowledge, information in the application (including proposed limits and controls) and relevant previous approvals. Both the short- and long-term impacts are considered.

Credible pathways to potential harm that were considered included exposure of people or other desirable organisms to the GM plant material, potential for persistence or dispersal of the GMOs, transfer of the introduced genetic material to non-GM canola plants and potential for fitness advantages to pest organisms. Potential harms associated with these pathways included toxicity and allergenicity to people, toxicity to desirable animals, and environmental harms due to weediness.

The risk assessment concludes that risks to the health and safety of people or the environment from the proposed dealings are negligible. No specific risk treatment measures are required to manage these negligible risks. The principal reasons for the conclusion of negligible risks are that the proposed limits and controls, such as not using GM plant material in commercial human food or animal feed, will effectively minimise exposure to the GMOs. In addition, there is no evidence to suggest the introduced genetic modifications would lead to harm to people or the environment.

# Risk management

The risk management plan describes measures to protect the health and safety of people and to protect the environment by controlling or mitigating risk. The risk management plan is given effect through licence conditions. Draft licence conditions are detailed in Chapter 4 of the RARMP.

As the level of risk is considered negligible, specific risk treatment is not required. However, since this is a limited and controlled release, the licence includes limits on the size, location and duration of the release, as well as controls to prohibit the use of GM plant material in commercial human food and animal feed, to minimise dispersal of the GMOs or GM pollen from the trial site, to transport GMOs in accordance with the Regulator's guidelines, to destroy GMOs at the end of the trial and to conduct post-harvest monitoring at the trial sites to ensure the GMOs are destroyed.

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# **Abbreviations**

AICIS	Australian Industrial Chemicals Introduction Scheme	
APVMA	Australian Pesticides and Veterinary Medicines Authority	
CCI	Confidential Commercial Information	
СМА	Cow's milk allergy	
DAFF	Department of Agriculture, Fisheries and Forestry	
DIR	Dealings involving Intentional Release	
FSANZ	Food Standards Australia New Zealand	
GM(O)	Genetically modified (organism)	
ha	Hectare(s)	
HGT	Horizontal gene transfer	
IBC	Institutional Biosafety Committee	
LGA	Local Government Area	
m	Metre(s)	
NLRD	Notifiable Low Risks Dealings	
NSW	New South Wales	
OGTR	Office of the Gene Technology Regulator	
PAT	Phosphinothricin acetyltransferase	
RARMP	Risk Assessment and Risk Management Plan	
Regulations	Gene Technology Regulations 2001	
Regulator	Gene Technology Regulator	
SA	South Australia	
TGA	Therapeutic Goods Administration	
the Act	The Gene Technology Act 2000	
UTR	Untranslated region	
Vic	Victoria	
WA	Western Australia	

Abbreviations V

# Chapter 1 Risk assessment context

# Section 1 Background

- 1. An application has been made under the *Gene Technology Act 2000* (the Act) for Dealings involving the Intentional Release (DIR) of genetically modified organisms (GMOs) into the Australian environment.
- 2. The Act and the Gene Technology Regulations 2001 (the Regulations), together with corresponding State and Territory legislation, comprise Australia's national regulatory system for gene technology. Its objective is to protect the health and safety of people, and to protect the environment, by identifying risks posed by or as a result of gene technology, and by managing those risks through regulating certain dealings with GMOs.
- 3. Section 50 of the Act requires that the Gene Technology Regulator (the Regulator) must prepare a Risk Assessment and Risk Management Plan (RARMP) in response to an application for release of GMOs into the Australian environment. Sections 50, 50A and 51 of the Act and sections 9 and 10 of the Regulations outline the matters which the Regulator must take into account and who must be consulted when preparing the RARMP.
- 4. The *Risk Analysis Framework* (OGTR, 2013) explains the Regulator's approach to the preparation of RARMPs in accordance with the Act and the Regulations. The Regulator has also developed operational policies and guidelines that are relevant to DIR licences. These documents are available from the Office of the Gene Technology Regulator (OGTR) <u>website</u>.
- 5. Figure 1 shows the information that is considered, within the regulatory framework, in establishing the risk assessment context. This information is specific for each application. Potential risks to the health and safety of people or the environment posed by the proposed release are assessed within this context. Chapter 1 provides the specific information for establishing the risk assessment context for this application.

#### **RISK ASSESSMENT CONTEXT**

The GMO Proposed GMO dealings

Modified genes Activities
Novel traits Limits
Controls

Parent organism (comparator)

Origin and taxonomy

Cultivation and use

Biology

Previous releases

Australian approvals

International approvals

**Receiving environment** 

Environmental conditions: abiotic and biotic factors

Production practices Related organisms

Similar genes and proteins

Figure 1. Summary of parameters used to establish the risk assessment context, within the legislative requirements, operational policies and guidelines of the OGTR, and the Risk Analysis Framework

6. In accordance with section 50A of the Act, this application is considered to be a limited and controlled release application, as the Regulator was satisfied that it meets the criteria prescribed by the Act. Therefore, the Regulator was not required to consult with prescribed experts, agencies and authorities before preparation of the RARMP.

#### 1.1 Interface with other regulatory schemes

- 7. Gene technology legislation operates in conjunction with other regulatory schemes in Australia. The GMOs and any proposed dealings may also be subject to regulation by other Australian government agencies that regulate GMOs or GM products, including Food Standards Australia New Zealand (FSANZ), the Australian Pesticides and Veterinary Medicines Authority (APVMA), the Therapeutic Goods Administration (TGA), the Australian Industrial Chemicals Introduction Scheme (AICIS) and the Department of Agriculture, Fisheries and Forestry (DAFF). These dealings may also be subject to the operation of State legislation recognising an area as designated for the purpose of preserving the identity of GM crops, non-GM crops, or both GM crops and non-GM crops, for marketing purposes.
- 8. To avoid duplication of regulatory oversight, risks that have been considered by other regulatory agencies will not be re-assessed by the Regulator.

# Section 2 The proposed dealings

- 9. Miruku Australia Pty Ltd (Miruku, the applicant) proposes to release several GM canola lines into the environment under limited and controlled conditions. The GM plants have been genetically modified for dairy protein production.
- 10. The purpose of the release is to assess dairy protein production in GM canola under field conditions. The applicant will also evaluate agronomic performance of the GM canola lines in the field.
- 11. The dealings involved in the proposed intentional release are to:
  - conduct experiments with the GMOs
  - breed the GMOs
  - propagate the GMOs
  - use the GMOs in the course of manufacture of a thing that is not the GMOs
  - grow the GMOs
  - transport the GMOs
  - dispose of the GMOs

and the possession, supply or use the GMOs in the course of any of these dealings.

- 12. Initial transformation of the GMOs will occur in Australia under a Notifiable Low Risks Dealings (NLRD) authorisation.
- 13. The GM plant material would not be used for commercial human food or animal feed.
- 14. The GM seeds will be processed to release the protein components to use in food products that may only be used in human sensory testing to assess their feel, smell, taste and appearance. They will not be used for commercial food or feed. Sensory testing would result in negligible consumption of the components from the GM seeds as the products are not intended to be swallowed during testing. These trials would only occur if Miruku obtains the appropriate approvals for each trial in accordance with the National Statement on Ethical Conduct in Human Research.

### 2.1 The proposed limits of the dealings (duration, size, location and people)

- 15. The release is proposed to take place between May 2025 and December 2029. Planting would occur primarily during the winter cropping season, but occasionally a summer crop cycle may also be used.
- 16. GM canola is proposed to be grown at up to 52 trial sites over the period of release. The proposed maximum number of sites, planting area per site, combined total planting area for each year, and cumulative maximum total planting area are detailed Table 1.

Table 1. Proposed duration and maximum number of sites and planting area per year

Year	Maximum number of sites per year	Maximum area (ha) per site	Maximum combined area (ha) per year	Cumulative maximum total area (ha)
2025	2	0.5	1	1
2026	5	1	5	6
2027	10	2.5	25	31
2028	15	7	105	136
2029	20	15	300	436

17. Sites for trial release would be selected from 135 possible local government areas (LGAs) in New South Wales (NSW), Victoria (Vic), Western Australia (WA) and South Australia (SA) (Table 2). The field trials would occur on research stations or private land in rural areas where persons other than those conducting dealings would not have access to the field trial sites.

Table 2. LGAs where GM canola trial sites may be located

New South Wales	Victoria	Western Australia	South Australia	
Berrigan	igan Ararat		Adelaide Plains	
Bland Ballarat		Beverley	Barossa	
Blayney	Benalla	Boddington	Light	
Cabonne	Buloke	Boyup Brook	Wakefield	
Coolamon	Campaspe	Bridgetown-Greenbushes		
Coonamble	Central Goldfields	Brookton		
Cootamundra-Gundagai	Colac Otway	Broomehill-Tambellup		
Cowra	Corangamite	Carnamah		
Dubbo	Gannawarra	Coorow		
Edward River	Glenelg	Corrigin		
Federation	Golden Plains	Cranbrook		
Forbes	Greater Bendigo	Cuballing		
Gilgandra	Greater Geelong	Cunderdin		
Greater Hume	Greater Shepparton	Dalwallinu		
Griffith				
Gunnedah Hindmarsh Donnybrook-Balir		Donnybrook-Balingup		
Gwydir	Horsham	Dowerin		
Hay	Indigo Dumbleyung			
Hilltops Loddon Espe		Esperance		
Inverell Macedon Ranges Gnowangerup		Gnowangerup		
Junee	Mildura	Goomalling		
Leeton	Mitchell	Greater Geraldton		
Liverpool Plains	Moira	Jerramungup		
Lockhart	Moorabool	Katanning		
Mid-Western	Mount Alexander	Kent		
Moree Plains	Moyne	Kojonup		
Murray River	Northern Grampians	Manjimup		
Murrumbidgee	Pyrenees	Merredin		
Muswellbrook	Southern Grampians	Mingenew		
Narrabri	Strathbogie	Moora		
Narrandera	Swan Hill	Morawa		
Narromine	Wangaratta	Nannup		
Orange	West Wimmera	Narrogin		

New South Wales	Victoria	Western Australia	South Australia
Parkes	Wodonga	Northam	
Snowy Valleys	Wyndham	Perenjori	
Tamworth	Yarriambiack	Pingelly	
Temora		Plantagenet	
Upper Hunter		Quairading	
Wagga Wagga		Ravensthorpe	
Walgett		Tammim	
Warren		Three Springs	
Warrumbungle		Toodyay	
Weddin		Victoria Plains	
		Wagin	
		Wandering	
		West Arthur	
		Wickepin	
		Williams	
		Wongan-Ballidu	
		Woodanilling	
		Wyalkatchem	
		York	

18. Only trained and authorised persons would be permitted to deal with the GM canola.

## 2.2 The proposed controls to restrict the spread and persistence of the GMOs in the environment

- 19. The applicant has proposed a number of controls to restrict the spread and persistence of the GM canola and the introduced genetic material in the environment. These include:
- locating each trial site at least 50 m away from the nearest natural waterway
- surrounding each planting area with either a 50 m monitoring zone, a 35 m monitoring zone and 15 m pollen trap, or a 10 m monitoring zone and covering the planting area with an insect-proof tent (see Figure 2)
- inspecting monitoring zones from 14 days prior to flowering until harvest, to identify and destroy volunteer canola or related species
- surrounding each monitoring zone with an isolation zone of 950 m, or 350 m with use of a pollen trap, or 390 m with use of an insect-proof tent, where no canola or sexually compatible species are grown (see Figure 2)
- treating non-GM canola plants grown in planting areas and pollen traps as if they are GMOs
- cleaning equipment and clothing after use on trial sites
- bagging or tenting GM canola
- using seeding or harvesting methods or equipment that minimise dispersal of GM plant material
- controlling rodents on trial sites
- restricting access to trial sites to authorised persons
- cleaning of planting areas post-harvest
- tilling and irrigating each planting area during the post-harvest monitoring period
- post-harvest monitoring of the trial sites for 24 months to identify any volunteer canola, and destroy volunteers before they reach flowering
- destroying all GMOs not required for further experimentation
- transporting and storing GMOs in accordance with the current Regulator's Guidelines for the

# Transport, Storage and Disposal of GMOs

not allowing the GMOs or GM products to be used for commercial human food or animal feed.

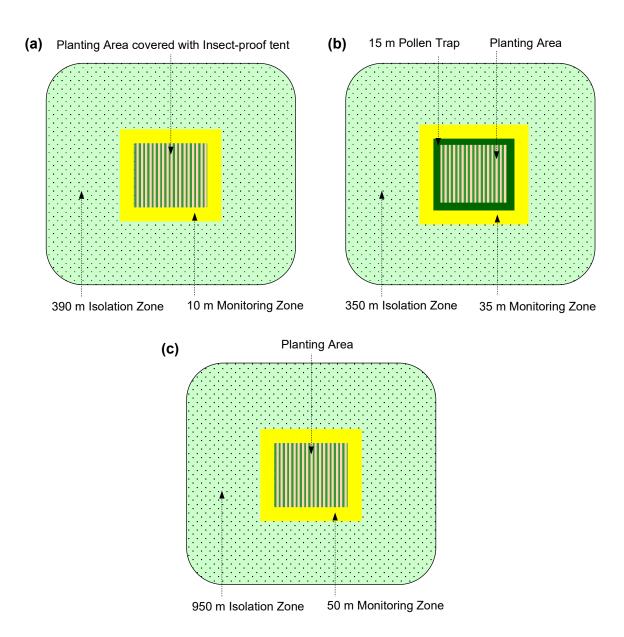


Figure 2. Diagrams (not to scale) showing the relationships between planting area, pollen trap, monitoring zone and isolation zone. Site layout (a) with insect-proof tent, (b) without insect-proof tent and with pollen trap, and (c) without insect-proof tent or pollen trap.

20. The proposed limits and controls are taken into account in the risk assessment (Chapter 2) and their suitability for containing the release will be evaluated in the risk management plan (Chapter 3).

# Section 3 The parent organism

- 21. The parent organism is *Brassica napus* L., commonly known as canola, rapeseed or oilseed rape. *B. napus* is exotic to Australia.
- 22. Canola is the third-most widely grown crop in Australia. It is grown mainly in WA, NSW, Vic and SA (ABARES, 2024a). Canola oil is used as food and the canola meal remaining after oil extraction is used as animal feed.

- B. napus is naturalised in Australia. In agricultural areas where it is grown, it can be a weed in subsequent crops. There are isolated reports of B. napus as an environmental weed in WA and Vic (Randall, 2017). However, the most recent Western Australian state government environmental weed risk assessment gives B. napus a weed risk rating of negligible to low (Moore and Nazeri, 2022), and the most recent Victorian state government environmental weed list gives B. napus a risk ranking score of zero and classified as 'lower risk' (White et al., 2022).
- Detailed information about the parent organism is contained in the document The Biology of Brassica napus L. (canola) and Brassica juncea (L.) Czern. & Coss. (Indian mustard) (OGTR, 2024), which was produced to inform the risk analysis process and is available from the Resources page on the OGTR website. Baseline information from this document will be used and referred to throughout the RARMP.
- While non-GM canola is not generally regarded as allergenic or toxic to humans or animals, it does produce some toxins and anti-nutritional factors such as erucic acid and glucosinolates, and some cases of canola food, pollen and dust allergies have also been reported (OGTR, 2024).

#### The GMOs, nature and effect of the genetic modification Section 4

The applicant proposes to release 5 groups of up to 10 canola lines each, genetically modified for dairy protein production.

### The genetic modifications in the GMOs proposed for release

- 27. The specific parental canola variety for the GMOs is 'Oscar'.
- 28. All GM canola lines will contain one of 2 variants (denoted BCN2 and BCN3) of the gene encoding βcasein (CSN2) from domestic cattle, fused to a gene derived from common plant species. Each line also contains a sequence to target the expressed  $\beta$ -casein fusion proteins to the cell wall or vacuoles and a glufosinate herbicide tolerance marker gene. Genes introduced into the GM canola lines are summarised in Table 3. Regulatory and localisation sequences are summarised in Table 4. Each GM canola line contains only a subset of the genetic elements listed in Table 3 and Table 4.
- The identities of some the genetic elements, and the arrangement of genetic elements in the vectors introduced into GM canola have been declared Confidential Commercial Information (CCI). Under section 185 of the Act, the confidential information is made available to the prescribed agencies and experts that are consulted on the RARMP for this application.

**Table 3. Introduced genes** 

Genetic element	Source organism	Encoded protein	Intended function
bar	Streptomyces hygroscopicus	Phosphinothricin acetyltransferase	Herbicide resistance
BCN2	Bos taurus	Miruku β-casein (CSN2) variant 2	β-casein production
BCN3	B. taurus	Miruku β-casein (CSN2) variant 3	β-casein production
CCI gene	CCI (common plant species)	CCI	ССІ

#### 4.1.1 **BCN2** and BCN3

- The introduced BCN2 and BCN3 constructs encode variants to A2 β-casein from cattle (Bos taurus). The purpose of the introduction of BCN2 and BCN3 is to produce the β-casein protein variants in seeds of GM canola. Expression is driven by a seed-specific promoter, either alone, or in combination with organelle localisation sequences.
- Caseins are the primary proteins in bovine milk, comprising 80% of total protein content (Hassanin et al., 2022). β-casein accounts for approximately 35% of bovine milk protein (Daniloski et al., 2022), and is

highly polymorphic, with at least 15 known variants (Sebastiani et al., 2020). The A1 and A2 variants are the most common, which differ by a single amino acid at residue 67 (Farrell et al., 2004).  $\beta$ -casein has a highly charged and hydrophilic N-terminal region and a primarily hydrophobic C-terminus (Dauphas et al., 2005), which is primarily due to the presence of 5 phosphorylated serine residues (Creamer et al., 1981; McCarthy et al., 2013). This amphiphilic nature enables  $\beta$ -casein to act as an emulsifier, and partial dephosphorylation of these serine residues results in reduced ability of  $\beta$ -casein to stabilise emulsions and the accumulation of  $\beta$ -casein in larger globules (Cassiano and Areas, 2003; McCarthy et al., 2013).

- 32. The *BCN2* and *BCN3* coding sequences are identical to A2  $\beta$ -casein sequence, except *BCN2* replaces serine residues at positions 17, 19 and 35 with aspartic acid, while *BCN3* replaces serine residues at positions 15, 18 and 35 with aspartic acid and serine residues at positions 17 and 19 with glutamic acid. These modifications are expected to result in partial dephosphorylation of  $\beta$ -casein and alter its emulsification properties.
- 33. *BCN2* and *BCN3* coding sequences are fused with a gene derived from common plant species and are expressed as a chimeric fusion protein using a seed-specific promoter.

#### 4.1.2 bar

- 34. The bar (bialaphos resistance) gene is derived from the bacterium Streptomyces hygroscopicus (Thompson et al., 1987). The bar gene encodes a phosphinothricin acetyltransferase (PAT) enzyme that confers tolerance to glufosinate herbicide. PAT acetylates glufosinate, converting it to N-acetyl-L-glufosinate, which is not toxic to plants (OECD, 2002).
- 35. Expression of *bar* is controlled by a 35S promoter from the Cauliflower mosaic virus (*CaMV35S*), which drives constitutive expression of *bar* in all plant tissues (Kay et al., 1987).
- 36. The Regulator has previously assessed and approved GM crops containing the *bar* gene for commercial release in Australia, most recently under licence <u>DIR-190</u>, and for field trials, most recently under <u>DIR 204</u>.

#### 4.1.3 Regulatory and localisation sequences

37. The GM canola lines will also contain introduced regulatory sequences and localisation signals to control expression of the inserted genes (Table 4). These include promoters to drive gene expression, terminators, and localisation signals to spatially restrict protein localisation. The identities of some of the promoters and 3'UTR sequences introduced into GM canola have been declared CCI by the Regulator. Under section 185 of the Act, the CCI has been made available to the prescribed agencies and experts that are consulted on the RARMP for this application.

Table 4. Introduced regulatory elements and localisation sequences

Genetic element	Source	Intended function
AtCel1 cell wall signal	Cell wall signal from Arabidopsis thaliana CEL1 gene	Cell wall targeting sequence
CaMV35S promoter	Cauliflower mosaic virus	Constitutive promoter
CCI 3'UTR Glycine max		Terminator and polyadenylation signal
CCI promoter G. max		Seed-specific promoter
C-terminal beta-conglycinin vacuolar sequence from Glycine max		Terminator
ER Retention C-terminal H/KDEL tag sequence from A. thaliana		Endoplasmic reticulum retention
GmCVS vacuolar signal Vacuole signal from G. max beta-conglycinin gene		Vacuole targeting sequence

Genetic element	Source	Intended function
HvAVSP vacuolar signal	Vacuole signal from <i>Hordeum vulgare</i> aleurain gene	Vacuole targeting sequence
Prr	Nicotiana tabacum	Cell wall targeting sequence
nos 3'UTR	Agrobacterium tumefaciens	Terminator and polyadenylation signal

# 4.2 Method of genetic modification

- 38. The GM canola lines were generated by *Agrobacterium*—mediated transformation. This method has been widely used in Australia and overseas for introducing genes into plants. Information about this method can be found in the document <u>Methods of plant genetic modification</u>, available from the OGTR Risk Assessment References page.
- 39. Canola breeding line 'Oscar' was transformed using the methodology described by Zhang et al. (2005). Transformants were selected on media containing the herbicide glufosinate. Selection agents were also used to eliminate *Agrobacterium* during *in vitro* selection of the transformed canola plants. *Agrobacterium* is not normally transmitted from one generation to the next via seed, therefore selected GM canola plants were propagated by single seed descent.
- 40. Parental canola lines were transformed with one of 5 binary plasmid vectors classified into 2 categories, which are described in Table 5. The arrangement of the genetic elements in the 5 vectors introduced into GM canola have been declared CCI.

Table 5. Categories of binary vector transformed into GM canola

Category	Vector number	Description
1	1	β-casein version 2 fusion (BCN2) fusion protein with glufosinate resistance
2	2-5	β-casein version 3 fusion (BCN3) fusion protein with various cell wall or vacuole targeting sequences and glufosinate resistance

### 4.3 Toxicity/allergenicity of the proteins associated with the introduced genes

- 41. As the GMOs are at an early stage of development, no toxicity or allergenicity studies have been conducted on the GM canola plants or purified protein produced by the introduced genetic elements. The genetic element components of the  $\beta$ -casein fusion protein and their encoded proteins have also not been assessed by authorities in any countries for toxicity and allergenicity.
- 42. Discussion of the toxicity/allergenicity of some the introduced genes that have been declared CCI is made available to the prescribed agencies and experts that are consulted on the RARMP for this application as required under section 185 of the Act.

#### 4.3.1 β-casein

- 43. A recent population-based study reported cow's milk allergy (CMA) in 1.3% of 1-year-old Australian infants (Soriano et al., 2023). Allergic reactions to cow's milk proteins are characterised by asthma, atopic dermatitis, urticaria (hives), rhinitis, gastrointestinal disorders, and anaphylaxis (Docena et al., 1996). Tolerance to cow's milk proteins usually develops as children mature, and consequently is present in less than 0.5% of adults (Fiocchi et al., 2010).
- 44. Caseins are known food allergens. In one study of 80 patients with known CMA, 100% of patients produced immunoglobulin-E (IgE) antibodies against casein proteins, indicating an immune reaction (Docena et al., 1996), while in another study of CMA patients, 75.3% produced IgE antibodies against caseins (Shoormasti et al., 2011).

- 45. There is little information was found regarding potential toxicity or allergenicity of  $\beta$ -casein specifically. Upon digestion, proteolysis of the A1 variant of  $\beta$ -casein produces the peptide  $\beta$ -casomorphin-7 (BCM-7), which is implicated in adverse gastrointestinal effects (Giribaldi et al., 2022). However, the applicant proposes to introduce sequence variations to the A2 variant in GM canola, which does not produce BCM-7 and consequently is not associated with digestive intolerance. Nonetheless, given the high rate of allergenicity to caseins among CMA patients, (Daniloski et al., 2022), allergenicity to A2  $\beta$ -casein variants is possible.
- 46. Allergenicity to caseins via dermal contact is rare and often requires existing damage to the skin barrier to cause immune reaction (Jensen et al., 2022). One study describes a single patient who developed dermatitis and rhinitis in response to casein protein present in microbiology laboratory culture media (Nakonechna et al., 2019). In another study, a single patient developed rhinitis and asthma following exposure to casein in a dermatological formulation associated with their occupation (Bonadonna et al., 2003). No information was found in the literature on toxicity or allergenicity associated with dermal contact with  $\beta$ -casein specifically.

## 4.3.2 PAT protein

- 47. The *bar* gene and its encoded PAT protein have been extensively assessed in previous RARMPs for commercial release of GM crops including canola (<u>DIR 021/2002</u>, <u>DIR 108</u>, <u>DIR 138</u>, <u>DIR 175</u>), cotton (<u>DIR 062/2005</u>, <u>DIR 143</u>, <u>DIR 145</u>, <u>DIR 173</u>) and a limited and controlled release of GM wheat (<u>DIR 204</u>). The PAT protein has been assessed to lack toxicity to humans or animals, or allergenicity in humans on the following basis:
  - the *bar* gene was derived from the common soil bacterium *S. hygroscopicus*, which is not considered a pathogen of humans or other animals;
  - no sequence homology has been found between PAT and any known toxic or allergenic proteins;
  - the PAT protein does not possess any of the characteristics associated with food allergens;
  - the PAT protein is inactivated by heat, e.g. through cooking, and by low pH, e.g. in the human stomach;
  - the PAT protein is rapidly degraded in simulated gastric or intestinal fluid; and
  - purified PAT protein was not toxic to mice and rats when administered at high doses in acute toxicity studies.
- 48. FSANZ has approved food derived from a number of GM crops expressing PAT protein as safe for human consumption. This includes GM canola (ANZFA, 2001; FSANZ, 2017), cotton (FSANZ, 2005a, 2010a, b, 2013), corn (FSANZ, 2005b) and rice (FSANZ, 2008).

#### 4.4 Characterisation of the GMOs

49. The applicant has stated that the GM canola lines proposed for release are still in development. Initial observations of the GM canola grown in controlled glasshouse conditions indicated no phenotypic differences compared to non-GM canola. Both GM and non-GM canola exhibited similar growth patterns, morphology, germination rates, time to flowering, plant height and seed count per plant. No quantitative data were provided by the applicant. Further data on the agronomic performance of the GM canola lines will be collected during the field trials.

# Section 5 The receiving environment

50. The receiving environment forms part of the context in which the risks associated with dealings involving the GMOs are assessed. Relevant information about the receiving environment includes abiotic and biotic interactions of the crop with the environment where the release would occur; agronomic practices for the crop; presence of plants that are sexually compatible with the GMOs; and background presence of the gene(s) used in the genetic modification (OGTR, 2013).

51. Detailed information about the commercial cultivation and distribution of canola in Australia is presented in the document *The Biology of Brassica napus L. (canola) and Brassica juncea (L.) Czern. & Coss. (Indian mustard)* (OGTR, 2024).

#### 5.1 Relevant abiotic factors

52. The geographical distribution of commercial canola cultivation in Australia is limited by several abiotic factors, the most important being water availability. Canola is generally grown as a winter crop in winter-dominant medium and high rainfall environments that receive annual rainfall of more than 350 mm (GRDC, 2009; OGTR, 2024). Germination of seed will only occur if there is sufficient soil moisture, and drought stress after anthesis can significantly reduce yield due to abortion of seed and reduced pod numbers. Canola is also sensitive to waterlogging (GRDC, 2009; OGTR, 2024). Other abiotic stresses that can reduce canola yields include frost, particularly during early pod development, and heat stress (GRDC, 2009).

#### **5.2** Relevant biotic factors

- 53. A number of diseases have the potential to significantly reduce the yield of canola. Blackleg disease caused by the fungal pathogen *Leptosphaeria maculans* is the most serious disease affecting commercial canola production in Australia (GRDC, 2009; OGTR, 2024). Other damaging diseases of canola include stem rot caused by the fungus *Sclerotinia sclerotiorum* and damping-off, caused mainly by the fungus *Rhizoctonia solani* (GRDC, 2009, 2015).
- 54. Canola is most susceptible to insect pests during establishment of the crop, particularly from redlegged earth mite (*Halotydeus destructor*), blue oat mites (*Penthaleus major*, *P. falcatus* and *P. tectus* sp. n.), lucerne fleas (*Sminthurus viridis*), cutworms (*Agrotis* spp.) and aphids (*Brevicoryne brassicae*, *Myzus persicae*, *Lipaphis pseudobrassicae* and *Aphis craccivora*, which may also act as viral vectors) (GRDC, 2009). From flowering to crop maturity, severe damage can be caused by aphids, Rutherglen bugs (*Nysius vinitor*), diamondback moth caterpillars (*Plutella xylostella*) and heliothis caterpillars (family *Noctuidae*).
- 55. Canola is highly susceptible to weed competition during the early stages of growth (GRDC, 2009, 2015). Hybrid canola varieties have greater seedling vigour than open-pollinated canola and so are more competitive with weeds (GRDC, 2015, 2017). Common weeds of Australian canola crops include grassy weeds (such as rigid ryegrass, vulpia and wild oat), volunteer cereals, and weeds from the Brassicaceae family including wild radish (*Raphanus raphanistrum*), Indian hedge mustard (*Sisymbrium orientale*), shepherds purse (*Capsella bursa-pastoris*), Asian mustard (*Brassica tournefortii*), charlock (*Sinapis arvensis*), turnip weed (*Rapistrum rugosum*) and Buchan weed (*Hirschfeldia incana*) (GRDC, 2015, 2017).

### 5.3 Relevant agricultural practices

- 56. The applicant specifies that GM canola seeds would be planted in trial sites during the winter cropping season, but a summer cropping cycle may also be used occasionally. Non-GM canola lines planted at the trial sites for comparative purposes would be treated as if it were GM canola.
- 57. GM and non-GM canola crops would be maintained in a similar manner to commercial canola crops (see Section 2.2). Standard cultivation practices for canola in Australia are discussed in *GRDC Canola GrowNotes* (GRDC, 2015, 2017).
- 58. The applicant proposes to only use the glufosinate tolerance conferred by the introduced *bar* gene as a selectable marker during transformation. Glufosinate herbicide is not intended to be applied to plants growing in the field trial. Glufosinate is not routinely used to control volunteer canola (AOF, 2019).
- 59. The applicant specifies that the GM canola would be grown at field sites either as an irrigated or dryland crop. Seeds would be planted in row plots with typical row spacing for canola, for example 30-40 cm, in plots spaced 1-2 m apart, although other configurations may be used. Small areas would be handplanted or planted with a small plot cone-seeder, while larger areas would be planted with commercial equipment.
- 60. Nitrogen fertiliser would be deep injected pre-plant or at planting. Land would be cultivated once or twice after planting to control weeds, aerate soil and allow efficient irrigation. Furrow or flood irrigation

would be used where necessary, and pre-irrigation may be conducted to store soil moisture and reduce salt levels in the soil.

- 61. Pest monitoring would be conducted once or twice per week by field technicians.
- 62. Canola seeds would be harvested by hand for small plantings or with commercial equipment for larger plantings, when seed moisture reaches 5-8%.
- 63. Planting areas will be left fallow after harvest and site cleaning, to facilitate the germination and monitoring of volunteers. Sites may be replanted with GM canola in subsequent years or would be planted with rotation crops such as cereals or pulses.
- 64. Additional agricultural practices proposed by the applicant are discussed further in Chapter 3, Section 3.1.

## 5.4 Presence of related plants in the receiving environment

- 65. Canola is primarily self-pollinating, but approximately 30% of seeds are produced by cross-pollination (Hüsken and Dietz-Pfeilstetter, 2007). Cross-pollination can be mediated by insects, wind or physical contact (OGTR, 2024).
- 66. Canola has been reported to outcross in the field with the following species: *Brassica carinata*, *B. napus*, *B. juncea*, *B. oleracea*, *B. rapa*, *Hirschfeldia incana* (Buchan weed), *R. raphanistrum* (wild radish) and *S. arvensis* (charlock) (Ford et al., 2006; Warwick et al., 2009). All of these species are known to be present in Australia, with the exception of *B. carinata* (Atlas of Living Australia, accessed 17 January 2025).
- 67. Of the *Brassica* species in Australia, canola may potentially hybridise under natural conditions with sexually compatible species that include: other *B. napus* groups or subspecies (including vegetables such as swedes, rutabaga and kale), *B. juncea*, *B. rapa* (wild turnip; includes vegetables such as turnip, Chinese cabbage and pak choi) and *B. oleracea* (wild cabbage; includes vegetables such as cauliflower, Brussels sprouts, kale and cabbage) (Salisbury, 2002). However, hybrids between *B. napus* and *B. oleracea* have been shown to be difficult to obtain (Ford et al., 2006).
- 68. Under open pollination conditions, naturally occurring hybrids between *B. napus* and the related weedy species *R. raphanistrum*, *H. incana* and *S. arvensis* have been reported at very low frequencies (Darmency and Fleury, 2000; Darmency et al., 1998; Salisbury, 2002), and are generally sterile or predominantly sterile (Salisbury, 2002).
- 69. Canola is widely grown as an oil seed crop in Australia, and the proposed trial sites are located in commercial canola growing regions. Commercial canola in these areas includes non-GM canola and GM canola authorised for commercial release. Most Australian canola crops are herbicide tolerant, with 4 herbicide tolerance traits available for commercial cultivation: triazine tolerance (non-GM), imidazolinone tolerance (non-GM), glyphosate tolerance (GM), or glufosinate tolerance (GM) (Brown, 2021; Matthews et al., 2021). Details of all GM canola varieties approved by the Regulator for commercial release in Australia are available from the OGTR website.

#### 5.5 Presence of similar genes and their products in the environment

- 70. *BCN2* and *BCN3* are modified sequences of the gene *CSN2* derived from cattle (*B. taurus*), which are present in the agricultural environment. The genetic sequence of the other part of the fusion protein constituent is derived from common plant species and these sequences naturally occur in all plants.
- 71. The *bar* gene is from the common soil bacterium *S. hygroscopicus*, which is widespread and prevalent in the environment.
- 72. Regulatory and localisation sequences are derived from common plants, a plant virus (CaMV) or a soil bacterium (*A. tumefaciens*) that are widespread in the environment. Although some of the regulatory sequences are derived from plant pathogens (*A. tumefaciens* and CaMV), they comprise only small parts of the total genomes and cannot of themselves cause disease.

# Section 6 Relevant Australian and international approvals

# 6.1 Australian approvals

# Approvals by the Regulator

73. The GM canola lines included in this application have not been previously approved for release in Australia.

# Approvals by other government agencies

74. The GM canola lines included in this application have not been previously approved by any other government agencies in Australia.

# 6.2 International approvals

75. The GM canola lines included in this application have not received any approvals from authorities in other countries.

# Chapter 2 Risk assessment

# Section 1 Introduction

76. The risk assessment identifies and characterises risks to the health and safety of people or to the environment from dealings with GMOs, posed by or as the result of gene technology (Figure 3). Risks are identified within the established risk assessment context (Chapter 1), taking into account current scientific and technical knowledge. A consideration of uncertainty, in particular knowledge gaps, occurs throughout the risk assessment process.

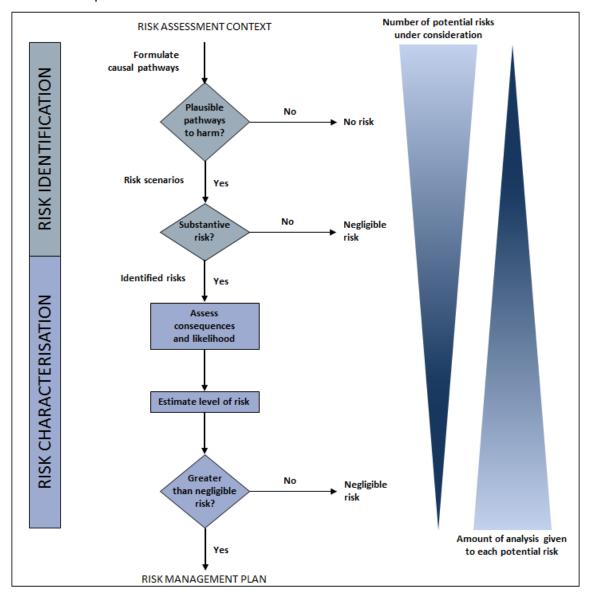


Figure 3. The risk assessment process

- 77. The Regulator uses a number of techniques to identify risks, including checklists, brainstorming, reported international experience and consultation (OGTR, 2013).
- 78. Risk identification first considers a wide range of circumstances in which the GMO, or the introduced genetic material, could come into contact with people or the environment. This leads to postulating plausible causal pathways that may give rise to harm for people or the environment from dealings with a GMO. These are called risk scenarios.

- 79. Risk scenarios are screened to identify substantive risks, which are risk scenarios that are considered to have some reasonable chance of causing harm. Risk scenarios that could not plausibly occur, or do not lead to harm in the short and long term, do not advance in the risk assessment process (Figure 4), i.e. the risk is considered to be no greater than negligible.
- 80. Risk scenarios identified as substantive risks are further characterised in terms of the potential seriousness of harm (consequence assessment) and the likelihood of harm (likelihood assessment). The consequence and likelihood assessments are combined to estimate the level of risk and determine whether risk treatment measures are required. The potential for interactions between risks is also considered.
- 81. A weed risk assessment approach is used to identify traits that may contribute to risks from GM plants, as this approach addresses the full range of potential adverse outcomes associated with plants. In particular, novel traits that may increase the potential of the GMO to spread and persist in the environment or increase the level of potential harm compared with the parental plant(s) are used to postulate risk scenarios (Keese et al., 2014). Risk scenarios postulated in previous RARMPs prepared for licence applications for the same or similar GMOs, are also considered.

## Section 2 Risk identification

- 82. Postulated risk scenarios are comprised of three components (Figure 4):
  - i. the source of potential harm (risk source)
  - ii. a plausible causal linkage to potential harm (causal pathway)
  - iii. potential harm to people or the environment.

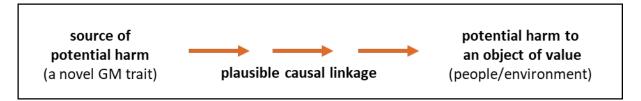


Figure 4. Components of a risk scenario

- 83. When postulating relevant risk scenarios, the risk context is taken into account, including the following factors detailed in Chapter 1:
  - the proposed dealings
  - the proposed limits including the extent and scale of the proposed dealings
  - the proposed controls to restrict the spread and persistence of the GMOs
  - the characteristics of the parent organism(s).

#### 2.1 Risk source

- 84. The sources of potential harms can be intended novel GM traits associated with one or more introduced genetic elements, or unintended effects/traits arising from the use of gene technology.
- 85. As discussed in Chapter 1, the GM canola lines have been modified by the introduction of fusion protein sequence comprised of variants of *CSN2* from cattle and a gene derived from common plant species. The introduced genes are intended to produce dairy protein in seed. These introduced genes are considered further as sources of potential harm.
- 86. The GM canola lines also contain the *bar* gene which confers glufosinate herbicide tolerance, and was used as selectable marker genes. The *bar* and its protein product PAT have been extensively characterised and assessed as posing negligible risk to human or animal health or to the environment by the Regulator, as well as by other regulatory agencies in Australia and overseas. As the *bar* gene has not

been found to pose a substantive risk to either people or the environment, its potential effects will not be further considered for this application.

- 87. The introduced genes are controlled by introduced regulatory sequences derived from various species (see Table 3). Regulatory sequences and introns are naturally present in all plants, and the introduced sequences are expected to operate in similar ways to endogenous sequences. The regulatory sequences are DNA that is not expressed as a protein, so exposure is to the DNA only and dietary DNA has no toxicity (Delaney et al., 2018). Hence, potential harms from the regulatory sequences and introns will not be considered further.
- 88. The genetic modifications involving introduction of genes have the potential to cause unintended effects in several ways. These include insertional effects such as interruptions, deletions, duplications or rearrangements of the genome, which can lead to altered expression of endogenous genes. There could also be increased metabolic burden due to expression of the introduced proteins, novel traits arising out of interactions with non-target proteins and secondary effects arising from altered substrate or product levels in biochemical pathways. However, these types of effects also occur spontaneously and in plants generated by conventional breeding. Accepted conventional breeding techniques such as hybridisation, mutagenesis and somaclonal variation can have a much larger impact on the plant genome than genetic engineering (Schnell et al., 2015). Plants generated by conventional breeding have a long history of safe use, and there are no documented cases where conventional breeding has resulted in the production of a novel toxin or allergen in a crop (Steiner et al., 2013). Therefore, the potential for the processes of genetic modification to result in unintended effects will not be considered further.

#### 2.2 Causal pathway

- 89. The following factors are taken into account when postulating plausible causal pathways to potential harm:
  - routes of exposure to the GMOs, the introduced gene(s) and gene product(s)
  - potential exposure to the introduced gene(s) and gene product(s) from other sources in the environment
  - the environment at the site(s) of release
  - agronomic management practices for the GMOs
  - spread and persistence of the GMOs (e.g. reproductive characteristics, dispersal pathways and establishment potential)
  - tolerance to abiotic conditions (e.g. climate, soil and rainfall patterns)
  - tolerance to biotic stressors (e.g. pests, pathogens and weeds)
  - tolerance to cultivation management practices
  - gene transfer to sexually compatible organisms
  - gene transfer by horizontal gene transfer (HGT)
  - unauthorised activities.
- 90. Although all of these factors are taken into account, some are not included in risk scenarios because they have been considered in previous RARMPs and are not expected to give rise to substantive risks.
- 91. The potential for horizontal gene transfer (HGT) from GMOs to species that are not sexually compatible, and any possible adverse outcomes, have been reviewed in the literature (Keese, 2008; Philips et al., 2022) and assessed in previous RARMPs. No risk greater than negligible was identified, due to the rarity of HGT events and because the gene sequences are already present in the environment and available for transfer via demonstrated natural mechanisms. Therefore, HGT will not be assessed further.
- 92. Previous RARMPs have considered the potential for unauthorised activities to lead to an adverse outcome. The Act provides for substantial penalties for non-compliance and unauthorised dealings with

GMOs. The Act also requires the Regulator to have regard to the suitability of the applicant to hold a licence prior to the issuing of the licence. These legislative provisions are considered sufficient to minimise risks from unauthorised activities, and no risk greater than negligible was identified in previous RARMPs. Therefore, unauthorised activities will not be considered further.

### 2.3 Potential harm

- 93. Potential harms from GM plants are based on those used to assess risk from weeds (Keese et al., 2014; Virtue, 2008) including:
  - harm to the health of people or desirable organisms, including toxicity/allergenicity
  - reduced biodiversity for nature conservation
  - reduced establishment or yield of desirable plants
  - reduced products or services from the land use
  - restricted movement of people, animals, vehicles, machinery and/or water
  - reduced quality of the biotic environment (e.g. providing food or shelter for pests or pathogens) or abiotic environment (e.g. negative effects on fire regimes, nutrient levels, soil salinity, soil stability or soil water table).
- 94. Judgements of what is considered harm depend on the management objectives of the land where the GM plant may be present. A plant species may have different weed risk potential in different land uses such as dryland cropping or nature conservation.

#### 2.4 Postulated risk scenarios

- 95. Four risk scenarios were postulated and screened to identify any substantive risks. These scenarios are summarised in Table 6 and examined in detail in Sections 2.4.1 2.4.4.
- 96. In the context of the activities proposed by the applicant and considering both the short and long term, none of the 4 risk scenarios gave rise to any substantive risks.

Table 6. Summary of risk scenarios from the proposed dealings with GM canola

Risk scenario	Risk source	Causal pathway	Potential harm	Substantive risk?	Reason
1	Introduced genes for dairy protein production	Cultivation of GM canola at trial sites  Exposure of people and desirable animals to products of the introduced genes	Increased toxicity or allergenicity for people OR increased toxicity to desirable animals	No	<ul> <li>The GM canola would not be used as commercial human food or animal feed</li> <li>The short duration and proposed controls for the field trial would restrict exposure of animals to the GM plants through contact or consumption</li> <li>The limits and controls of the field trial would restrict exposure of people to the GM plants</li> <li>β-casein fusion proteins are not expected to be toxic but could be allergenic. However, people will not consume GM canola seeds or products (other than a small number of people as part of the sensory tests), and animals are unlikely to consume a dose that would cause toxicity.</li> </ul>
2	Introduced genes for dairy protein production	Cultivation of GM canola at trial sites  Persistence of GM canola seed at trial sites or dispersal of GM seed outside trial limits  Establishment of populations of volunteer GM plants expressing the introduced genes in the environment	Increased toxicity or allergenicity for people OR increased toxicity to desirable animals OR reduced establishment or yield of desirable plants	No	<ul> <li>The limits and controls of the field trial would minimise dispersal or persistence of GM seeds</li> <li>GM canola is susceptible to standard weed management measures</li> <li>As discussed in Risk Scenario 1, no substantive risk was identified for increased adverse effects in people or toxicity to animals</li> <li>Canola has limited ability to compete with other plants and the genetic modifications are not expected to alter the dispersal characteristics of the GM canola.</li> </ul>

Risk scenario	Risk source	Causal pathway	Potential harm	Substantive risk?	Reason
3	Introduced genes for dairy protein production	Cultivation of GM canola at trial sites  Pollen from GM plants dispersed outside the trial sites  Outcrossing with sexually compatible plants  Establishment of populations of hybrid GM plants expressing the introduced genes in the environment	Increased toxicity or allergenicity for people OR increased toxicity to desirable animals OR reduced establishment or yield of desirable plants	No	<ul> <li>The controls of the field trial would minimise pollen flow to sexually compatible plants outside the trial sites</li> <li>As discussed in Risk Scenario 1, no substantive risk was identified for increased adverse effects in people or toxicity to animals</li> <li>As discussed in Risk Scenario 2, the genetic modifications are not expected to alter the dispersal characteristics of the GM canola.</li> </ul>
4	Introduced genes for dairy protein production	Cultivation of GM canola at trial sites  Consumption of GM canola seed by pest animals  Increased fitness of pest animals	Reduced establishment of yield of desirable plants OR Reduced biodiversity	No	<ul> <li>The limited scale and other proposed limits and controls minimise exposure of pests to the GM seeds</li> <li>GM canola seeds are unlikely to contribute a large proportion of the overall diet for pest species</li> <li>Consumption of GM canola seed containing β-casein fusion proteins is unlikely to provide a fitness advantage to pest species</li> <li>Pests are controlled by current pest management practices.</li> </ul>

#### 2.4.1 Risk scenario 1

Risk source	Introduced genes for dairy protein production
Causal pathway	Cultivation of GM canola at trial sites  Exposure of people and desirable animals to products of the introduced genes
Potential harm	Increased toxicity or allergenicity for people OR Increased toxicity to desirable animals

#### Risk source

97. The source of potential harm for this postulated risk scenario is the introduced genes for dairy protein production in GM canola plants.

### Causal pathway

- 98. The GM canola would be grown at the trial sites. As the introduced genes for dairy protein production are controlled by a seed-specific promoter, the encoded fusion proteins would be produced in seeds of the GM plants. A review of the literature corroborates seed-specific expression of the promoter. A study utilising the promoter to express an introduced gene in a common plant species showed expression in seeds but not in leaf tissue. Trial staff would be exposed to the seeds during harvesting. The applicant has not tested the seeds or any other tissues for levels of produced fusion proteins.
- 99. People involved in the breeding, cultivating, harvesting, transporting and processing of the GM canola may be exposed to expressed proteins through contact with the GMOs, including direct contact with GM plant material. This would be expected to primarily occur at the trial site, but could also occur anywhere the GM seeds are transported or used. The proposed limits and controls of the trial would minimise the likelihood that people or other organisms would be exposed to GM plant material. The GM canola is not proposed for use in commercial human food, and therefore, people are unlikely to be exposed to the introduced genetic elements or their products as a result of consuming GM canola seed. The applicant proposes that GM canola will only be handled by trained and authorised staff and all GM plant material would be transported in accordance with the Regulator's <u>Guidelines for the Transport, Storage and Disposal of GMOs</u>.
- 100. The applicant proposes human sensory testing of ingredients isolated from GM canola seed to assess the taste, smell and texture of the oil products and protein concentrates isolated from the seed. Although the products are not intended to be swallowed during testing, some ingestion of small amounts may occur. People participating in sensory evaluations could also be exposed to protein concentrates enriched in  $\beta$ -casein fusion proteins by dermal contact, contact with mucous membranes or inhalation.
- 101. The applicant does not propose use of the GM canola in commercial animal feed, and livestock would not be permitted to graze the trial sites. Therefore, livestock are not expected to be exposed to GM plants grown at the trial sites. Desirable wild animals, such as native mammals and birds, could enter the trial sites and consume GM plants including seeds. The limited size and duration of the field trial would restrict the number of desirable wild animals exposed to GM plants grown at the trial sites.

#### **Potential harm**

- 102. If the people or animals were exposed to the GM canola at the trial site, the potential harms are increased toxicity or allergenicity to people or increased toxicity to desirable animals.
- 103. Toxicity is the adverse effect(s) of exposure to a dose of a substance as a result of direct cellular or tissue injury, or through the inhibition of normal physiological processes (Felsot, 2000). Allergenicity is the

potential of a substance to elicit an immunological reaction following its ingestion, dermal contact or inhalation, which may lead to tissue inflammation and organ dysfunction (Arts et al., 2006).

- 104. As mentioned in Chapter 1 Section 3, while non-GM canola is not generally regarded as allergenic or toxic to humans or animals, it does produce some allergens, toxins and anti-nutritional factors. No allergic reactions to processed canola oil have been reported in the literature, though there are reports of food allergies to *B. napus* seed extracts (Poikonen et al., 2006) and flour (Alvarez et al., 2001). As discussed in Chapter 1, Section 4.1, the proteins encoded by the introduced genes result in dairy protein production. There is no reasonable expectation that the introduced genes expressed in the GM canola would affect the pathways producing endogenous toxins or allergens in canola, or lead to the production of novel toxins or allergens.
- 105. As discussed in Chapter 1, Section 4.3, the introduced fusion proteins and genetic constructs have not been assessed for toxicity and allergenicity by any authorities or animal feeding studies. However, both caseins and the second fusion protein constituent derived from common plant species are known food allergens. Allergy to milk proteins is most common in young children and decreases rapidly with age (Jensen et al., 2022). Allergenicity to casein proteins occurs via ingestion of the proteins. The GM canola seeds containing  $\beta$ -casein fusion proteins are not intended for human consumption and, as noted in paragraph 100, only negligible levels of ingestion are likely during sensory assessment. Furthermore, allergic reactions to casein proteins via dermal contact are rare, and often require a damaged skin barrier to cause an immune response (Jensen et al., 2022). In addition, the molecular weight of a compound must be less than 500 daltons in size to penetrate intact skin (Bos and Meinardi, 2000). Bioinformatic analysis of the amino acid sequence for the endogenous *CSN2* protein from which the BCN2 and BCN3 sequences are derived indicate a molecular weight (independent of the fusion protein) of 18.62 kDa, which is too large to penetrate the skin.
- 106. There are very few studies about toxicity of casein proteins in the literature and no reports related to humans were found. Canola seeds can be eaten by several animal species, including beetles, rodents and birds. Seed predation is greatest when seeds are buried at shallow depths, and mice can climb canola plants and feed on seed pods. Some reports indicate casein tolerance in laboratory animals. An early study suggested it is almost impossible to administer lethal amounts of casein orally to rats (Boyd et al., 1967). A high-casein diet was also shown to attenuate renal injury and inflammation in rats without adverse health effects (Shimada et al., 2020). Conversely, another study demonstrated that consumption of a high-casein diet for a 4-week period resulted in a twofold increase in colonic DNA damage in rats (Toden et al., 2005). However, it is unclear whether  $\beta$ -casein specifically is responsible for these genotoxic effects. Further, it is unlikely that rodents will consume GM canola seed at levels sufficient to mimic the amount of casein protein consumed in this study.
- 107. While studies have demonstrated allergenicity of milk proteins and caseins more broadly, no scientific evidence was identified to indicate allergenicity or toxicity of  $\beta$ -casein specifically in humans or animals. The genetic constructs proposed for insertion into GM canola are derived from the sequence for the A2 variant of  $\beta$ -casein, which is a common component of cow's milk consumed by humans and is not associated with gastrointestinal intolerance linked to the A1 variant.
- 108. Further information about the  $\beta$ -casein fusion protein relevant to consideration of this risk scenario has been declared CCI by the Regulator. Under section 185 of the Act, the CCI has been made available to the prescribed agencies and experts that are consulted on the RARMP for this application.

## Conclusion

109. Risk scenario 1 is not identified as a substantive risk because the GM plant material would not be used as commercial human food and animal feed, the GM diary fusion proteins are not expected to be toxic, and the proteins encoded by the introduced genes are too large to penetrate an intact skin barrier and cause allergenicity via dermal contact. Further, the limits and controls of the field trial would restrict exposure of people and desirable animals to the GM plants. Therefore, this risk could not be considered greater than negligible and does not warrant further detailed assessment.

#### 2.4.2 Risk scenario 2

Risk source	Introduced genes for dairy protein production	
Causal pathway	Cultivation of GM canola at trial sites  Persistence of GM canola seed at trial sites or dispersal of GM seed outside trial limits  Establishment of populations of volunteer GM plants expressing the introduced genes in the environment	
Potential harm	Increased toxicity or allergenicity for people  OR  Increased toxicity to desirable animals  OR  Reduced establishment or yield of desirable plants	

#### **Risk source**

110. The source of potential harm for this postulated risk scenario is the introduced genes for dairy protein production in GM canola plants.

#### **Causal pathway**

- 111. The GM canola would be grown at the trial sites. GM seeds could be physically dispersed outside the trial sites by human activity, animal activity, wind or water. GM seeds could also persist on trial sites after completion of the trial. These GM seeds could grow in the environment and establish populations of volunteer GM plants.
- 112. Viable GM canola seeds could be dispersed outside the trial sites by human activity, such as transport of seeds and movement of agricultural machinery. Inadvertent dispersal of seeds by people dealing with the GMOs would be minimised by cleaning of all equipment prior to removal from the trial sites. The applicant also proposes that areas used for equipment cleaning would be inspected for volunteers.
- 113. GM seeds could be dispersed outside the trial limits by animal activity. Canola seeds have no specific adaptions, such as burrs or hooks, for dispersal by animals (OGTR, 2024). Dispersal of canola seed via endozoochory (consumption and excretion of viable seed) by birds only occurs at very low levels (Twigg et al., 2008; Woodgate et al., 2011). Canola seeds could be transported short distances by hoarding animals, such as ants and mice. The applicant is also proposing to implement rodent control measures at trial sites.
- 114. Canola seeds lack specialised structures that would assist their dispersal by wind (OGTR, 2024). However, the GM canola may be windrowed prior to harvesting, and under strong wind conditions plant material containing seeds could disperse outside trial sites. The applicant proposes to minimise the likelihood of wind dispersal by implementing measures including ensuring high density of GMO plants prior to windrowing, using a windrow roller, or by appropriate site selection.
- 115. The GM canola seeds could be dispersed by water during flooding or heavy runoff, although seeds are unlikely to remain viable after prolonged exposure to water (OGTR, 2024). To minimise the potential for seed dispersal during flooding, the applicant proposes to locate the trial at sites that are not prone to flooding.
- 116. During harvest of the GM canola, a small percentage of the seeds are expected to remain on the trial sites. Persistence of GMOs at the trial sites after the field trial is finished could occur if seeds in the seed bank were dormant. Canola generally does not exhibit primary dormancy, but secondary dormancy has been described (OGTR, 2024). A study in western Canada revealed that secondary seed dormancy

prolonged persistence of volunteer canola plants (Gulden et al., 2003). Persisting canola seed banks have been shown to significantly contribute to the dynamics of feral canola populations (Pivard et al., 2008). A long-term monitoring study in Germany detected GM canola volunteers in arable fields for up to 15 years after the field trial concluded, but did not detect spatial dispersion (Belter, 2016). In Australia, volunteers can be found for up to 3 years after growing canola due to persistence in seed banks, though the majority of volunteer seedlings emerge the year following a canola crop (AOF, 2019).

- 117. To minimise persistence of GM seeds on the trial sites, the applicant proposes to promote seed germination by light post-harvest tillage and irrigation. During a post-harvest monitoring period, the applicant proposes to inspect the trial sites at least once every 35 days for at least 24 months, and destroy any canola volunteers, until the site is free from volunteers for at least 12 months. The suitability of the proposed controls to manage GM seed dispersal and persistence is discussed in detail in Chapter 3, Section 3.1. These control measures are expected to minimise persistence of viable GM canola seeds on the trial sites.
- 118. If GM canola seeds were dispersed outside trial limits, it is unlikely that they would establish ongoing volunteer populations. Even in environments without active weed management, volunteer canola populations along transportation routes rely on recurrent spillages to persist (Yoshimura et al., 2006) and volunteer canola dispersed into natural areas was reported to rapidly become extinct (Busi and Powles, 2016). Though the applicant does not anticipate that the introduced genetic modifications would affect the overall ability of GM canola volunteers to survive in the environment, this is an area of uncertainty.
- 119. In agricultural areas of Australia where canola is grown, volunteer populations are controlled by a range of weed management measures. Effective methods for control of canola volunteers include grazing, mowing, cultivation and application of a range of knockdown or selective herbicides (AOF, 2019). The introduced genetic modifications are not expected to affect the susceptibility of GM volunteers to standard weed management measures. Some of the canola lines will contain the *bar* gene and be tolerant to glufosinate herbicide. However, as discussed in Chapter 1, Section 5.3, glufosinate herbicide is not routinely used for controlling volunteer canola (AOF, 2019).
- 120. The applicant states that GM canola grown in controlled glasshouse conditions did not display any differences in growth patterns, morphology and fertility compared to non-GM canola. Both GM and non-GM canola exhibited similar germination rates, time to flowering, height, and seed count per plant. Therefore, the introduced genetic modifications are not expected to impact persistence or dispersal of GM canola seed.

#### **Potential harm**

- 121. If the GM canola entered the Australian environment, the potential harms are increased toxicity or allergenicity to people, increased toxicity to desirable animals, or reduced establishment or yield of desirable plants.
- 122. As discussed in Risk scenario 1, no substantive risk was identified for increased toxicity or allergenicity of the GM canola for people or increased toxicity to desirable animals. Although the  $\beta$ -casein fusion protein constituents may be associated with allergenicity, the GM plant material would not be used as commercial human food and animal feed, and the proteins encoded by the introduced genes are too large to penetrate an intact skin barrier to cause allergenicity via dermal contact. The limits and controls of the field trial would further restrict exposure to the GM canola and any associated toxicity or allergenicity.
- 123. Populations of volunteer GM canola could reduce establishment or yield of desirable plants. GM volunteers could directly compete with agricultural crops, pastures or native vegetation. GM volunteers could also reduce the yield of commercial canola crops by providing a reservoir for pathogens, such as the important fungal diseases blackleg and stem rot (see Chapter 1, Section 5.2). No information could be found to suggest that the introduced genetic modifications are likely to make the GM canola more susceptible to pathogens.
- 124. Canola is considered a less competitive crop species than wheat or barley (GRDC, 2011), which are the main crops grown in NSW, Vic, SA and WA (ABARES, 2024b). All domesticated crop species are

expected to be poor competitors with pasture species or established native vegetation. Therefore, canola volunteers have limited ability to compete with desirable plants. As discussed in Chapter 1, Section 4.4, the applicant did not identify any differences in morphological or growth characteristics in the GM canola plants compared to non-GM canola. Therefore, the GM canola plants are not expected to show increased spread or persistence in the environment. However, given the GM plants have not yet been grown in the field, it is uncertain whether the introduced genetic modifications would increase their overall competitiveness. In addition, no information could be found to suggest that the introduced genetic modifications would enable the GM canola to produce allelopathic substances which would negatively affect plant establishment around them. A standard condition of a licence for a field trial would be that the applicant immediately notify the OGTR of any unintended effects.

125. The genetic modifications are not expected to affect the susceptibility of GM volunteers to standard weed management measures. Although some of the GM canola lines will contain the *bar* gene and be tolerant to glufosinate herbicide, as discussed in Section 5.3, glufosinate herbicide will not be applied to the GM canola plants as part of this trial and other methods are available to manage GM canola volunteers.

#### Conclusion

126. Risk scenario 2 is not identified as a substantive risk because the proposed limits and controls of the field trial would minimise dispersal or persistence of GM seeds, the GM canola is susceptible to standard weed management measures, and the genetic modifications are not expected to increase the overall weediness or competitiveness of the GM canola with other plants. Therefore, this risk could not be considered greater than negligible and does not warrant further detailed assessment.

#### 2.4.3 Risk scenario 3

Risk source	Introduced genes for dairy protein production	
Causal pathway	Cultivation of GM canola at trial sites  Pollen from GM plants dispersed outside the trial sites  Outcrossing with sexually compatible plants  Establishment of populations of hybrid GM plants expressing the introduced genes in the environment	
Potential harm	Increased toxicity or allergenicity for people  OR  Increased toxicity to desirable animals  OR  Reduced establishment or yield of desirable plants	

#### **Risk source**

127. The source of potential harm for this postulated risk scenario is the introduced genes for dairy protein production in GM canola plants.

## **Causal pathway**

128. The GM canola would be grown at the trial sites. Pollen from the GM plants could be transported outside the trial site by wind or insect vectors and could fertilise sexually compatible plants. Hybrid seeds containing the introduced genes could be harvested or could grow as volunteers.

- 129. It should be noted that vertical gene flow *per se* is not considered an adverse outcome, but may be a link in a chain of events that may lead to an adverse outcome.
- 130. Canola is primarily self-pollinating, but approximately 30% of seeds are produced by cross pollination. Outcrossing decreases rapidly with distance, with the majority of cross-pollination occurring over distances less than 10 m (OGTR, 2024). The introduced genetic modifications are not expected to affect the pollen dispersal characteristics of the GM canola.
- 131. The GM canola could outcross with nearby canola crops or volunteers, if there is synchronicity of flowering. As discussed in Chapter 1, Section 5.4, canola can also occasionally hybridise with the related horticultural crops *B. juncea*, *B. oleracea* and *B. rapa* and the related weeds *H. incana*, *R. raphanistrum* and *S. arvensis*.
- 132. The applicant has proposed control measures to minimise pollen flow from GM plants growing on the trial sites to sexually compatible plants outside the trial sites (Chapter 1, Section 2.2). Each planting area would be surrounded by a monitoring zone which would be inspected for sexually compatible plants from before the GM canola flowers and until harvest. The applicant also proposes the planting area will be surrounded by a large isolation zone, or a smaller isolation zone combined with use of a either a pollen trap or an insect-proof tent. In addition, any GM volunteers growing on the trial sites after harvest would be destroyed prior to flowering. The suitability of the proposed controls to manage pollen flow is discussed in detail in Chapter 3, Section 3.1. These controls are expected to minimise pollen flow from the GM canola to sexually compatible non-GM plants outside the trial sites.
- 133. If pollen from GM plants fertilised plants in a commercial canola crop, hybrid GM seeds could be harvested for human food and animal feed, or be replanted in a crop. However, even in the complete absence of measures to restrict pollen flow, outcrossing rates between neighbouring commercial canola fields are less than 0.1% under Australian conditions (Rieger et al., 2002). Therefore, the planting seed described in this risk pathway could only contain a very low proportion of hybrid GM seed, so people and desirable animals could only be exposed to very low levels of the hybrid GMOs.
- 134. If pollen from GM plants fertilised sexually compatible plants growing as crops, volunteers or weeds, the hybrid GM seeds could grow as volunteers. Populations of hybrid GM volunteers could be consumed by desirable animals or could reduce the establishment or yield of desirable plants.
- 135. It is not expected that the intended modifications would change the pollination characteristics of the GM canola to increase the likelihood of pollination of non-GM canola or related species.

#### **Potential harm**

- 136. The potential harms from this risk scenario are adverse health effects in people and/or desirable organisms, or reduced establishment or yield of desirable plants.
- 137. As discussed in Risk scenario 1, no substantive risk was identified for increased toxicity or allergenicity of the GM canola for people or increased toxicity to desirable animals. Similarly, in the rare event of outcrossing between the GM canola and sexually compatible plants, the proteins encoded by the introduced genes are too large to penetrate an intact skin barrier to cause allergenicity via dermal contact.
- 138. As discussed in Risk scenario 2, the GM canola is not likely to be weedier or more competitive than non-GM canola. Similarly, in hybrids between the GM plants and sexually compatible plants, the genetic modifications are not expected to confer an overall increased ability to compete with other plants.

#### Conclusion

139. Risk scenario 3 is not identified as a substantive risk because the controls of the field trial would minimise pollen flow to sexually compatible plants outside the trial sites. GM hybrids are not likely to differ from the GM canola, for which Risk scenarios 1 and 2 did not identify adverse health effects in people, toxicity in animals or weediness as substantive risks. Therefore, this risk could not be considered greater than negligible and does not warrant further detailed assessment.

#### 2.4.4 Risk scenario 4

Risk source	Introduced genes for dairy protein production		
	Cultivation of GM canola at trial sites		
Causal pathway	Consumption of GM canola seed by pest animals		
	■ Increased fitness of pest animals		
	<b>↓</b>		
	Reduced establishment or yield of desirable plants		
Potential harm	OR		
	Reduced biodiversity		

#### **Risk source**

140. The source of potential harm for this postulated risk scenario is the introduced genes for dairy protein production in GM canola plants.

### **Causal pathway**

- 141. The GM canola would be grown at the trial sites. The GM canola produces seeds with dairy protein production.
- 142. Pest animals, such as rodents, large mammalian species, or birds may ingest the GM canola seed at the trial site and may have a fitness advantage as a result of consuming the GM canola seeds. Populations of these pests may then increase as a consequence of this increased fitness.
- 143. As discussed in Risk scenario 1, insects, rodents and birds may consume canola seeds. The applicant has proposed the use of insect proof tents as one option for the planting area. If this were used, access to GM canola by insects would be limited by the insect-proof tent. The applicant has also proposed use of pesticide treatment to control insects as required. As noted in Risk scenario 3, that applicant has proposed rodent control measures that would limit rodent access.
- 144. If animals do consume GM canola seeds, it is likely that the seed will only make up a subset of the animal's overall diet, and the GM seed will only be available for a short period of time before harvest and until any seed on the soil surface is buried by tilling.

#### **Potential harm**

- 145. The potential harms from this risk scenario are reduced establishment or yield of desirable plants or reduced biodiversity.
- 146. If pests consuming the GM canola seeds with expression of  $\beta$ -casein fusion proteins had a fitness advantage over those which did not consume the GM canola seeds, populations could increase to a greater extent than expected. In that case, they may have a greater negative effect on native or other desirable plants, or on desirable animals. This could occur via reduced establishment or yield of native or other desirable plants due to increased consumption by greater pest animal populations. In natural environments, this may result in the loss of biodiversity. In agricultural areas this may result in reduced crop yields. They might also increase competition with desirable animals for food sources or territory and thereby reduce animal biodiversity.
- 147. While there is a body of literature supporting caseins as a source of amino acids and their role in muscle growth in humans, there is limited evidence to show similar growth benefits in animals consuming caseins. A study in which dairy cows were provided 300 g casein per day for 2 weeks via infusion found that a casein-infused diet increased amino acid concentrations and milk production compared to cows fed grass silage alone (Vanhatalo et al., 2003). Another study in malnourished rats determined that re-feeding with a

casein-infused diet increased bone strength and body weight catch-up compared to a whey-infused diet, but was insufficient to return the rats to a normal body weight (Masarwi et al., 2016). However, no studies were found to suggest a casein-infused diet leads to enhanced fitness or competitiveness of animals.

- 148. Birds may consume the GM canola seeds containing  $\beta$ -casein fusion proteins, however any increased fitness advantage to birds consuming the seeds is expected to be very minor and would not be inherited by subsequent generations.
- 149. If enhanced fitness occurred in pest animals that consumed the GM canola seed producing  $\beta$ -casein, the improvement would be expected to be minor and transient, it would be isolated to the individual animal, and is unlikely to change the existing impact of known pest animals if consumed. Additionally, the limits and controls of the trial proposed by the applicant are proposed to limit the consumption GM canola seeds by pest species. The details of those control measures are discussed in Chapter 3. Based on these factors, it is considered that the increased levels of  $\beta$ -casein fusion proteins in the GM canola seeds is unlikely to provide a fitness advantage that would increase the existing impact of known pest animals, though this is an area of uncertainty.

#### Conclusion

150. Risk scenario 4 is not identified as a substantive risk due to the lack of evidence to suggest any fitness advantage to pest animals as a result of consuming higher levels of  $\beta$ -casein fusion proteins, and the proposed limits and controls to restrict exposure of animals to the GMOs, including pest management practices. Further, GM seed consumption is unlikely to cause sustained benefit for animals. Therefore, this risk could not be considered greater than negligible and does not warrant further detailed assessment.

# Section 3 Uncertainty

- 151. Uncertainty is an intrinsic property of risk analysis and is present in all aspects of risk analysis. This is discussed in detail in the Regulator's Risk Analysis Framework document.
- 152. Uncertainty is addressed by approaches such as balance of evidence, conservative assumptions, and applying risk management measures that reduce the potential for risk scenarios involving uncertainty to lead to harm. If there is residual uncertainty that is important to estimating the level of risk, the Regulator will take this uncertainty into account in making decisions.
- 153. As field trials of GMOs are designed to gather data, there are generally data gaps when assessing the risks of a field trial application. However, field trial applications are required to be limited and controlled. Even if there is uncertainty about the characteristics of a GMO, limits and controls restrict exposure to the GMO, and thus decrease the likelihood of harm.
- 154. For DIR 215, uncertainty is noted particularly in relation to:
  - potential for increased toxicity to livestock/other desirable organisms or increased allergenicity to people of the GM canola
  - potential for the genetic modifications to increase plant competitiveness and survival
  - potential for the genetic modifications to increase competitiveness of pest species if consumed.
- 155. Additional data, including information to address these uncertainties, may be required to assess possible future applications with reduced limits and controls, such as a larger scale trial or the commercial release of these GMOs.
- 156. Chapter 3, Section 4, discusses information that may be required for future release.

## Section 4 Risk evaluation

157. Risk is evaluated against the objective of protecting the health and safety of people and the environment to determine the level of concern and, subsequently, the need for controls to mitigate or reduce risk. Risk evaluation may also aid consideration of whether the proposed dealings should be authorised, need further assessment, or require collection of additional information.

158. Factors used to determine which risks need treatment may include:

- risk criteria
- level of risk
- uncertainty associated with risk characterisation
- interactions between substantive risks.

159. Four risk scenarios were postulated whereby the proposed dealings might give rise to harm to people or the environment. In the context of the limits and controls proposed by the applicant, and considering both the short and long term, none of these scenarios were identified as substantive risks. The principal reasons for these conclusions are summarised in Table 6 and include:

- limits on the size and duration of the proposed release
- suitability of controls proposed by the applicant to restrict the spread and persistence of the GM canola plants and their genetic material
- the products of the introduced genes are not expected to be toxic
- GM canola plant material is not expected to confer increased fitness to pest species
- none of the GM plant material would enter commercial human food or animal feed.

160. Therefore, risks to the health and safety of people, or the environment, from the proposed release of the GM canola plants into the environment are considered to be negligible. The *Risk Analysis Framework* (OGTR 2013), which guides the risk assessment and risk management process, defines negligible risks as risks of no discernible concern with no present need to invoke actions for mitigation. Therefore, no additional controls are required to treat these negligible risks. Hence, the Regulator considers that the dealings involved in this proposed release do not pose a significant risk to either people or the environment.<sup>3</sup>

<sup>&</sup>lt;sup>3</sup> As none of the proposed dealings are considered to pose a significant risk to people or the environment, Section 52(2)(d)(ii) of the Act mandates a minimum period of 30 days for consultation on the RARMP.

# Chapter 3 Risk management plan

# Section 1 Background

- 161. Risk management is used to protect the health and safety of people and to protect the environment by controlling or mitigating risk. The risk management plan addresses risks evaluated as requiring treatment and considers limits and controls proposed by the applicant, as well as general risk management measures. The risk management plan informs the Regulator's decision-making process and is given effect through licence conditions.
- 162. Under section 56 of the Act, the Regulator must not issue a licence unless satisfied that any risks posed by the dealings proposed to be authorised by the licence can be managed in a way that protects the health and safety of people and the environment.
- 163. All licences are subject to 3 conditions prescribed in the Act. Section 63 of the Act requires that each licence holder inform relevant people of their obligations under the licence. The other statutory conditions allow the Regulator to maintain oversight of licensed dealings: section 64 requires the licence holder to provide access to premises to OGTR inspectors and section 65 requires the licence holder to report any information about risks or unintended effects of the dealing to the Regulator on becoming aware of them. Matters related to the ongoing suitability of the licence holder must also be reported to the Regulator.
- 164. The licence is also subject to any conditions imposed by the Regulator. Examples of the matters to which conditions may relate are listed in section 62 of the Act. Licence conditions can be imposed to limit and control the scope of the dealings and to manage risk to people or the environment. In addition, the Regulator has extensive powers to monitor compliance with licence conditions under section 152 of the Act.

### Section 2 Risk treatment measures for substantive risks

165. The risk assessment of risk scenarios listed in Chapter 2 concluded that there are negligible risks to people or the environment from the proposed field trial of GM canola. These risk scenarios were considered in the context of the scale of the proposed release (Chapter 1, Section 2.1), the proposed controls (Chapter 1, Section 2.2), and the receiving environment (Chapter 1, Section 5), and considering both the short and the long term. The risk evaluation concluded that no specific risk treatment measures are required to treat these negligible risks. Limits and controls proposed by the applicant and other general risk management measures are discussed below.

# Section 3 General risk management

166. The limits and controls proposed in the application were important in establishing the context for the risk assessment and in reaching the conclusion that the risks posed to people and the environment are negligible. Therefore, to maintain the risk context, licence conditions have been imposed to limit the release to the proposed size, location and duration, and to restrict the spread and persistence of the GMOs and their genetic material in the environment. The conditions are discussed and summarised in this Chapter and listed in detail in the draft licence.

#### 3.1 Limits and controls on the release

167. Sections 2.1 and 2.2 of Chapter 1 list the limits and controls proposed by the applicant. Many of these are discussed in the 4 risk scenarios considered in Chapter 2. The appropriateness of these limits and controls is considered further in the following sections.

### 3.1.1 Consideration of limits proposed by the applicant

168. The applicant proposes that the field trial would take place between May 2025 and December 2029. This would limit the duration of the trial to 4 years and 8 months. The applicant has also proposed that GM canola would be grown at a maximum of 2 sites per year in 2025, 5 in 2026, 10 in 2027, 15 in 2028 and 20

in 2029. Across all sites, GM canola is proposed to be grown on planting areas up to a combined total area of 1 ha in 2025, 5 ha in 2026, 25 ha in 2027, 105 ha in 2028, and 300 ha in 2029 – a total of 436 ha over the duration of the licence. The applicant proposes up to 7 ha per site in 2028 and 15 ha per site in 2029. However, monitoring such a large area could be challenging and, at this stage, the applicant does not have experience monitoring large sites for GM field trials. Therefore, it is considered appropriate to limit the maximum combined planting area for each site to 5 ha in 2028 and 2029 (a maximum of 206 ha over the duration of the licence, if issued). This would ensure effective monitoring for volunteer GM canola on each site. The size and short duration of the trial would restrict the exposure of people and desirable animals to the GMOs (Risk scenario 1).

169. The applicant proposes that only authorised and trained people would be permitted to deal with the GMOs. Standard licence conditions included in the draft licence state that only people authorised by the licence holder are covered by the licence and permitted to deal with the GMOs. In addition, the licence holder must inform all people dealing with the GMOs of relevant licence conditions. These measures would ensure that the field trial is conducted in accordance with the specified limits and controls (important for all risk scenarios).

# 3.1.2 Consideration of proposed controls regarding exposure to the GMOs

- 170. The applicant proposes to grow both GM canola and non-GM canola in the trial sites. The draft licence limits the plants that can be intentionally grown in the planting area to the GMOs, non-GM canola, and any plants approved in writing by the Regulator. As non-GM canola may be mingled with or fertilised by GM canola, a standard licence condition has been proposed requiring non-GM canola plants grown in a trial site to be treated as if they are GMOs. This measure manages the dispersal or persistence of GM seed (Risk scenario 2).
- 171. The applicant proposes that GM plants or products from the GM plants would not be used in commercial human food or animal feed, and this requirement has been included in the draft licence. This condition would maintain the risk context by restricting the exposure of people and desirable animals to the GMOs via consumption (Risk scenario 1) and would also minimise dispersal of the GMOs by livestock or during transport or processing for human food or animal feed use (Risk scenario 2).
- 172. The GM canola is not expected to be toxic (Chapter 1 Section 4.3), so there are no specific controls proposed to manage increased toxicity to people or animals. General controls included in the draft licence will limit exposure of people to the GMOs.
- 173. Any human sensory testing must be approved by a Human Research Ethics Committee in accordance with the National Statement on Ethical Conduct in Human Research. This condition would maintain the risk context by restricting the exposure of people to the GM canola by consumption (Risk scenario 1).
- 174. People who are allergic to β-casein or the other potentially allergenic protein constituent of the fusion protein could have an allergic reaction to GM canola as a result of the genetic modification. The applicant has proposed to restrict trial site access to authorised personnel. People working with the GM canola could be exposed to potentially allergenic  $\beta$ -casein fusion proteins expressed in seeds via contact with the GM canola seeds. However, allergic reactions to casein proteins via dermal contact is rare and both introduced proteins are too large to penetrate the skin to cause allergenicity (Section 2.4.1). Due to the limited scale of the proposed trial, only a limited number of people would be exposed to the GM canola, but it is possible that one or more of those people could have a known allergy to milk proteins or to the other fusion protein constituent. Consequences of allergic reactions to these proteins can be severe, including anaphylaxis. Although the  $\beta$ -casein fusion proteins would be expressed using a seed-specific promoter, the GM canola has not been characterised for fusion protein expression levels in seeds or other tissues. Therefore, it is possible  $\beta$ -casein fusion proteins could be expressed in tissues other than seeds. Therefore, an additional draft licence condition is included to not engage personnel with a known allergy to the proteins expressed as a result of modifications in the GM canola to conduct dealings that may expose them to the GM canola. It is considered that the proposed condition is appropriate to protect people with known allergies to these proteins from contact with GM canola seed (Risk scenario 1).

175. The applicant has proposed fencing around trial sites to restrict access by large animals. This would limit exposure to GM canola to large animals through direct contact with plant material (e.g. through grazing). If consumed, potential harm to desirable animals from the introduced genetic elements is expected to be minimal. In addition, the draft licence requires that the GMOs must not be used in a way that results in its use as animal feed, which would restrict access to the GMOs by animals. Therefore, the draft licence does not impose the use of fencing at trial sites.

### 3.1.3 Consideration of proposed controls regarding pollen flow from the GMOs

- 176. The applicant has proposed 3 different planting options to control pollen flow from the trial sites while the GMOs are flowering (Chapter 1, Figure 2).
- 177. The first option is to cover the planting area with an insect-proof tent, and to surround the planting area with a 10 m monitoring zone and a 390 m isolation zone (Chapter 1, Figure 2a). The tents would be in place at least 7 days before flowering until the GMOs complete flowering, and would be inspected for damage fortnightly and after any extreme weather event. The tents are expected to prevent insect-mediated pollen flow. The tents may also reduce wind-mediated pollen flow as it is expected that surrounding the GMOs with a tent would lessen air flow across the GMOs. Therefore, the use of an insect-proof justifies a reduced monitoring zone of 10 m and an isolation zone of 390 m. This option has been included in licences for previous GM canola field trials (e.g. <u>DIR 188</u>) and is considered an effective means of restricting pollen flow from canola.
- 178. The second option is to surround the planting area with a 15 m pollen trap of non-GM canola plants, a 35 m monitoring zone and a 350 m isolation zone (Chapter 1, Figure 2b). The pollen trap would be comprised of a mix of early, mid and late flowering non-GM canola to ensure synchronised flowering between the pollen trap and the GM canola. Pollen trap plants may provide sufficient forage for incoming pollinating insects so that they do not visit the GM plants, and any insects that reach the GM plants are expected to deposit most GM pollen on pollen trap plants while exiting the trial site. Pollen trap plants may also absorb some pollen dispersed by wind. The applicant has proposed an isolation zone of 390 m. As discussed in previous RARMPs for GM canola field trials (e.g. <u>DIR 164</u>, <u>DIR 188</u> and <u>DIR 205</u>), the use of a pollen trap justifies a reduced isolation zone of 350 m and thus an overall distance of 400 m between the GMOs and any crops of related species.
- 179. The third option is to surround the planting area with a 50 m monitoring zone and a 950 m isolation zone (Chapter 1, Figure 2c). A combined isolation distance of 1 km as proposed in this trial setup is considered appropriate where pollen tents or pollen trap crops are not used, or when a pollen trap fails to function (e.g. failure to grow to a required density, or to form a continuous barrier, or to flower at the same time as the GM plants) or the insect-proof tent fails. The Canadian Regulations and Procedures for Pedigreed Seed Crop Production (CSGA, 2022) require that foundation production of male sterile *B. napus* seed must be separated from other *B. napus* plants by an 800 m isolation distance, of which the first 50 m must be practically free from related plants, and the remaining distance must be reasonably free from related plants. Therefore, the proposed 50 m monitoring zone and combined 1 km isolation distance, which are more stringent than these Canadian requirements, are considered effective measures to restrict pollen flow from the GM canola. This option has been included in licences for previous GM canola field trials (e.g. DIR 164 and DIR 188) and is considered an effective means of restricting pollen flow from canola.
- 180. For all 3 proposed options, a draft licence condition requires that the monitoring zone would be inspected at least once every 35 days from 14 days prior to flowering of the GMOs until the GMOs are harvested, to ensure that it is free from any sexually compatible plants. This condition also requires that the isolation zone would be inspected at least once every 35 days from 14 days prior to flowering of the GMOs until the GMOs complete flowering, to ensure that it is free from intentionally planted sexually compatible plants. GM canola plants would not be planted at a trial site if any plants that are sexually compatible with canola were being grown in the monitoring or isolation zones.
- 181. The proposed measures to control pollen flow would minimise outcrossing between the GMOs grown on the trial sites and sexually compatible plants growing outside the trial sites (Risk scenario 3).

182. After harvest of the trial sites, the applicant proposes to inspect for volunteer canola plants at least once every 35 days for at least 24 months and until the site is free of volunteers for at least 12 months. Identified volunteer plants would be destroyed prior to flowering which would prevent further pollen flow. These post-harvest inspections are required in the licences for other GM canola field trials and are considered an effective means of restricting pollen flow from GM canola volunteers to plants outside the trial sites (e.g. <u>DIR 164</u> and <u>DIR 188</u>). These post-harvest inspections are considered appropriate to manage pollen flow and are included in the draft licence.

### 3.1.4 Consideration of proposed controls regarding dispersal of the GMOs

- 183. The applicant proposes that any equipment used with the GMOs would be cleaned as soon as practicable and before use for any other purpose, to avoid movement of viable plant material together with equipment. Personnel and clothing will also undergo physical examination before leaving the trial site to prevent unintentional movement of GM material. The applicant would contain the GM seeds during transport and storage in accordance with the Regulator's <u>Guidelines for the Transport, Storage and Disposal of GMOs</u>. These measures for the handling of GMOs would minimise exposure of people and other desirable organisms to the GMOs (Risk scenario 1), and dispersal of GMOs into the environment (Risk scenario 2) during transport, and have been included in the proposed licence conditions.
- 184. The applicant proposes to not locate the trial site in a flood-prone area in order to minimise the chance of viable plant material being washed away from the sites. This has been included as a condition in the draft licence. The draft licence also requires the trial sites to be located at least 50 m away from waterways as a standard licence condition for canola licences and that any extreme weather events must be reported to the Regulator. These measures would minimise dispersal of GM seeds by flooding (Risk scenario 2).
- 185. The GM canola seeds could be dispersed short distances from the trial sites during sowing, windrowing or harvest activities; by pod shattering, by seed-hoarding behaviours of animals such as ants or rodents; or by strong winds or runoff after heavy rain. As described in Section 3.1.3, the planting areas would be surrounded by monitoring zones that are inspected while the GMOs are growing, so any volunteers growing from dispersed GM seeds during this period would be detected and destroyed. Specific conditions to minimise dispersal of GM plant material from windrowed plants by wind or rain have also been included in the draft licence. The applicant also proposes to inspect the monitoring zones after harvest to destroy any volunteers growing from dispersed GM seeds. As the short-distance seed dispersal mechanisms listed above are unlikely to transport seeds further than 10 m from the trial sites, the draft licence only requires post-harvest inspections of the innermost 10 m of the monitoring zone.
- 186. The applicant proposes to control the number of rodents present at trial sites. This would limit the potential dispersal of GMOs outside the trial sites (Risk scenario 2). As discussed above, rodents are unlikely to transport seeds further than 10 m from the trial sites. Post-harvest inspection of the innermost 10 m of the monitoring zone is considered sufficient to detect volunteers growing from rodent-mediated dispersal. Therefore, additional rodent control measures are not included in the draft licence.
- 187. The draft licence includes additional conditions to manage short-distance dispersal of GM seeds. These include requiring the trial site to be cleaned within 14 days after harvest by a method that removes GM seeds from the soil surface, and requiring post-harvest inspections of any area used to clean equipment or any other area where GMOs are known to have dispersed. This combination of controls would minimise short-distance dispersal of GM seeds leading to establishment of volunteer populations outside the trial sites (Risk scenario 2).

### 3.1.5 Consideration of proposed controls regarding persistence of the GMOs

188. After harvest, the applicant proposes to destroy GMOs not required for further evaluation or future trials. This would involve both cleaning the trial site within 14 days after harvest in a manner that destroys any surviving GMOs and destroying any harvested GM seed that is not required for experimentation or future planting. The measures are considered appropriate and have been included in the draft licence.

- 189. The applicant has proposed that GMOs would be destroyed by destructive analysis (e.g. ground up, hammer milled and/or roller milled), herbicide application, root cutting and shredding/mulching, uprooting, burning/incineration, light tillage to a depth of no more than 5 cm, autoclaving, or seed burial to a depth of at least 1 m. These methods are considered effective for rendering canola plants and/or seed non-viable, and have been included in the licence. To ensure the effectiveness of destruction by seed burial, a draft licence condition specifies how this must be carried out, including a requirement that seeds must be wet at time of burial to encourage decomposition.
- 190. As discussed in Section 3.1.2, the applicant proposes to also grow non-GM canola on the trial sites, which would be treated as if they are GMOs. Non-GM canola in the trial site may be cross-pollinated by the GM canola, resulting in hybrid seeds. Therefore, it is appropriate to require non-GM canola to be destroyed in the same manner as GM canola, to manage persistence of the GMOs. This measure is included in the draft licence.
- 191. As detailed in Section 3.1.3, the applicant proposes to inspect the trial sites after harvest and destroy any identified volunteers. In minimum-tillage Australian farms, the canola seedbank is reported to decline rapidly, and no viable seed was recovered from the seedbank by 2.5 years after canola harvest (Baker and Preston, 2008). Similarly, OGTR monitoring data for 9 GM canola trial sites planted in 2015 found that in most sites no canola volunteers emerged more than one year after harvest and no volunteers emerged at any site more than 2.5 years after harvest. Therefore, the proposed monitoring for least 24 months, and until the site is free of volunteer canola plants for at least 12 months, is considered to be appropriate. This monitoring duration was also required for previous GM canola field trials and is considered effective for managing persistence of canola seed (e.g. DIR 164, DIR 188 and DIR 205).
- 192. To deal with the case of failed crops that are not harvested, draft licence conditions require that GMOs must be harvested or destroyed within 8 months after planting, and that if all GMOs in a planting area have been destroyed, then the area is considered to have been harvested and cleaned.
- 193. The applicant also proposes that canola volunteers identified during inspections would be destroyed prior to flowering, which would prevent GM canola seed dispersal (Risk scenario 2) and pollen flow to non-GM plants outside the trial site (Risk scenario 3). This measure is considered appropriate and has been included in the draft licence conditions.
- 194. The applicant proposes shallow cultivation of the trial sites to encourage seed germination. The draft licence conditions require that tillage depth would be no greater than 5 cm, to avoid deep burial of seed that could induce dormancy. The first tillage would occur within 60 days after harvest and the final tillage would occur during the volunteer-free period prior to sign-off. To ensure that the final tillage produces conditions that are conducive to germination of volunteers, the draft licence requires this tillage to be followed by specified levels of rainfall or irrigation that provide sufficient moisture to the seedbank.
- 195. The combination of control measures described in this section would minimise the persistence of GM seeds leading to establishment of GM volunteer populations in the environment (Risk scenario 2).

#### 3.1.6 Summary of draft licence conditions to be implemented to limit and control the release

- 196. A number of licence conditions are proposed to limit and control the release, based on the above considerations. These include requirements to:
  - limit the duration of the release to the period from May 2025 to December 2029
  - limit the size of the release to a maximum of 2 sites per year in 2025, 5 in 2026, 10 sites in 2027, 15 sites in 2028 and 20 sites in 2029, with planting areas up to a combined total area of 1 ha in 2025, 5 ha in 2026, 25 ha in 2027, 75 ha in 2028, and 100 ha in 2029
  - limit the location of the release to nominated local government areas in NSW, Vic, WA and SA
  - not allow GM plant material to be used in commercial human food or animal feed
  - treat any non-GM canola grown in planting areas like the GMOs

- not permit persons with an allergy to the proteins produced as a result of the modification, or related proteins to conduct dealings that may expose them to GM plant material
- control pollen flow from the trial sites using one of the following options:
  - a. cover the planting area with an insect proof tent, and surround the planting area with a monitoring zone of 10 m and an isolation zone of a further 390 m, or
  - b. surround the planting area with a pollen trap of 15 m, a monitoring zone of 35 m and an isolation zone of a further 350 m, or
  - c. surround the planting area with a monitoring zone of 50 m and an isolation zone of a further 950 m
- locate trial sites at least 50 m from any natural waterways
- transport and store the GMOs in accordance with the Regulator's guidelines
- destroy all GMOs not required for further evaluation or future trials
- clean equipment used with the GMOs before use for any other purpose
- clean the planting areas, monitoring zones, areas where equipment has been cleaned, and other areas where GMOs are known to have dispersed after harvest
- apply any measures to promote the germination of any canola seeds that may be present in the soil after harvest, including watering and shallow tillage
- monitor each trial site at least once every 35 days for at least 24 months after harvest and until no volunteers are identified for at least 12 months, and destroy any canola plants that emerge.

### 3.2 Other risk management considerations

197. All DIR licences issued by the Regulator contain a number of conditions that relate to general risk management. These include conditions relating to:

- applicant suitability
- contingency plans
- identification of the persons or classes of persons covered by the licence
- reporting requirements
- access for the purpose of monitoring for compliance.

#### 3.2.1 Applicant suitability

198. In making a decision whether or not to issue a licence, the Regulator must have regard to the suitability of the applicant to hold a licence. Under Section 58 of the Act, matters that the Regulator must take into account include:

- any relevant convictions of the applicant
- any revocation or suspension of a relevant licence or permit held by the applicant under a law of the Commonwealth, a State or a foreign country
- the capacity of the applicant to meet the conditions of the licence.

199. If a licence were issued, the conditions would include a requirement for Miruku to inform the Regulator of any information that would affect their suitability.

200. In addition, Miruku must have access to an Institutional Biosafety Committee (IBC) and be an accredited organisation under the Act.

### 3.2.2 Contingency plan

- 201. If a licence were issued, Miruku would be required to submit a contingency plan to the Regulator before planting the GMOs. This plan would detail measures to be undertaken in the event of any unintended presence of the GM canola outside permitted areas.
- 202. Before planting the GMOs, Miruku would also be required to provide the Regulator with a method to reliably detect the GMOs or the presence of the genetic modifications in a recipient organism.

#### 3.2.3 Identification of the persons or classes of persons covered by the licence

203. If issued, the persons covered by the licence would be the licence holder and employees, agents or contractors of the licence holder and other persons who are, or have been, engaged or otherwise authorised by the licence holder to undertake any activity in connection with the dealings authorised by the licence. Prior to growing the GMOs, Miruku would be required to provide a list of people and organisations that will be covered by the licence, or the function or position where names are not known at the time.

### 3.2.4 Reporting requirements

204. If issued, the licence would require the licence holder to immediately report any of the following to the Regulator:

- any additional information regarding risks to the health and safety of people or the environment associated with the dealings
- any contraventions of the licence by persons covered by the licence
- any unintended effects of the field trial.

205. A number of written notices would also be required under the licence regarding dealings with the GMO, to assist the Regulator in designing and implementing a monitoring program for all licensed dealings. The notices include:

- expected and actual dates of planting
- details of areas planted to the GMOs
- expected dates of flowering
- expected and actual dates of harvest and cleaning after harvest
- details of inspection activities.

### 3.2.5 Monitoring for compliance

206. The Act stipulates, as a condition of every licence, that a person who is authorised by the licence to deal with a GMO, and who is required to comply with a condition of the licence, must allow inspectors and other persons authorised by the Regulator to enter premises where a dealing is being undertaken for the purpose of monitoring or auditing the dealing. Post-release monitoring continues until the Regulator is satisfied that all the GMOs resulting from the authorised dealings have been removed from the release sites.

207. If monitoring activities identify changes in the risks associated with the authorised dealings, the Regulator may also vary licence conditions, or if necessary, suspend or cancel the licence.

208. In cases of non-compliance with licence conditions, the Regulator may instigate an investigation to determine the nature and extent of non-compliance. The Act provides for criminal sanctions of large fines and/or imprisonment for failing to abide by the legislation, conditions of the licence or directions from the Regulator, especially where significant damage to the health and safety of people or the environment could result.

# Section 4 Issues to be addressed for future releases

209. Additional information has been identified that may be required to assess an application for a commercial release of the GM canola, or to justify a reduction in limits and controls.

#### 210. This includes:

- additional molecular and biochemical characterisation of the GM canola lines, particularly with respect to expression of the introduced genes and proteins in the seeds
- biochemical characterisation of the GM canola lines, particularly with respect to potential for allergenicity related to the introduced genetic elements
- additional phenotypic characterisation of the GM canola lines leading to potential for increased weediness.

#### Section 5 Conclusions of the consultation RARMP

- 211. The risk assessment concludes that the proposed limited and controlled release of GM canola poses negligible risks to the health and safety of people or the environment as a result of gene technology. These negligible risks do not require specific risk treatment measures.
- 212. If a licence were issued, conditions would be imposed to limit the release to the proposed size, location and duration, and to restrict the spread and persistence of the GMOs and their genetic material in the environment, as these were important considerations in establishing the context for assessing the risks.

# **Chapter 4** Proposed licence conditions

# Section 1 Interpretations and definitions

#### 1. In this licence:

- (a) unless defined otherwise, words and phrases used in this licence have the same meaning as they do in the Act and the Regulations;
- (b) words importing a gender include every other gender;
- (c) words in the singular number include the plural and words in the plural number include the singular;
- (d) expressions used to denote persons generally (such as "person", "party", "someone", "anyone", "no one", "one", "another" and "whoever"), include a body politic or corporate as well as an individual;
- (e) references to any statute or other legislation (whether primary or subordinate) are a reference to
  a statute or other legislation of the Commonwealth of Australia as amended or replaced from
  time to time and equivalent provisions, if any, in corresponding State law, unless the contrary
  intention appears;
- (f) where a word or phrase is given a particular meaning, other grammatical forms of that word or phrase have corresponding meanings;
- (g) specific conditions prevail over general conditions to the extent of any inconsistency.

#### 2. In this licence:

'Act' means the *Gene Technology Act 2000* (Commonwealth) or the corresponding State law under which this licence is issued.

'Canola' means plants of the species Brassica napus L.

'Clean' means, as the case requires:

- (a) in relation to Equipment or a facility, remove and/or Destroy the GMOs; or
- (b) in relation to an area of land specified in this licence as requiring Cleaning:
  - i. Destroy Canola plants, if present, to the reasonable satisfaction of the Regulator, and
  - ii. remove Canola seeds from the soil surface to the reasonable satisfaction of the Regulator.

Note: The intent of removing seeds from the soil surface is to minimise seed dispersal. One method of removing seeds from the soil surface is Tillage, which moves seeds to under the soil. Tillage must be in accordance with Condition 38.

'Contingency Plan' means a written plan detailing measures to be taken in the event of the unintended presence of the GMOs outside an area that must be inspected. A Contingency Plan must include procedures to:

- (a) ensure the Regulator is notified immediately if the licence holder becomes aware of the event; and
- (b) recover and/or Destroy the GMOs to the reasonable satisfaction of the Regulator; and
- (c) inspect for and Destroy any Volunteers that may exist as a result of the event to the reasonable satisfaction of the Regulator.

'Destroy', (or 'Destruction') means, as the case requires, kill by one or more of the following methods:

- (a) uprooting;
- (b) root cutting and shredding/mulching;

- (c) Tillage, but only in accordance with Condition 38;
- (d) treatment with herbicide;
- (e) burning/incineration;
- (f) autoclaving;
- (g) milling/hammer milling;
- (h) crushing or grinding of seed;
- (i) burial, but only in accordance with Condition 39;
- (j) a method approved in writing by the Regulator.

Note: 'As the case requires' has the effect that, depending on the circumstances, one or more of these techniques may not be appropriate. For example, treatment with herbicide would not successfully kill GM seeds.

**'Equipment'** includes, but is not limited to, seeders, harvesters, tents, threshers, transport equipment (e.g. bags, containers, trucks), clothing, footwear and tools.

**'Extreme Weather'** includes, but is not limited to, fires, flooding, cyclones or torrential rain, that could disperse GMOs or affect the licence holder's ability to comply with licence conditions.

**'Flowering'** is taken to begin when any plant of the class of plants referred to in a particular condition first has an open flower, and is taken to end when all plants in the class of plants no longer have flowers.

'GM' means genetically modified.

**'GMOs'** means the genetically modified organisms that are the subject of the dealings authorised by this licence. GMOs include live plants and viable seed.

'Insect-proof' means sufficient to prevent the entry of insects that commonly pollinate Canola flowers.

'Isolation Zone' means an area of land extending outwards from the outer edge of the Monitoring Zone, as shown in Figure 1.

**'Logbook'** means a written or electronic record containing information required to be collected and maintained by this licence and which is able to be presented to the Regulator on request.

'Monitoring Zone' means an area of land extending outwards from the outer edge of the Planting Area, or the outer edge of a Pollen Trap if a Pollen Trap is employed, as shown in Figure 1.

'OGTR' means the Office of the Gene Technology Regulator.

**'Personal Information'** means information or an opinion about an identified individual, or an individual who is reasonably identifiable:

- (a) whether the information or opinion is true or not; and
- (b) whether the information or opinion is recorded in a material form or not.

'Planting Area' means an area of land where the GMOs and non-GM Canola are intentionally planted and grown pursuant to this licence, but does not include the Pollen Trap.

'Plant Material' means any part of the GM or non-GM Canola plants grown at a Planting Area or Pollen Trap, whether viable or not, or any product of these plants.

'Pollen Trap' means an area of land extending outwards at least 15 metres from the outer edge of a Planting Area, where only Pollen Trap Plants are grown, as shown in Figure 1.

'Pollen Trap Plants' means non-GM Canola grown in a Pollen Trap.

'Regulations' means the Gene Technology Regulations 2001 (Commonwealth) or the corresponding State law under which this licence is issued.

'Regulator' means the Gene Technology Regulator.

**'Related Species'** means plants of the species *Brassica napus*, *B. rapa*, *B. juncea*, *B. oleracea*, *Hirschfeldia incana*, *Raphanus raphanistrum* or *Sinapis arvensis*, but does not include plants intentionally grown in the Planting Area or Pollen Trap in accordance with licence conditions.

'Sign off' means a notice in writing from the Regulator, in respect of an area, that post-Cleaning obligations no longer apply to that area.

'Tillage' means the use of any technique to disturb the soil.

Note: Tillage must be in accordance with Condition 38.

'Volunteers' means GM or non-GM Canola plants, which have not been intentionally grown.

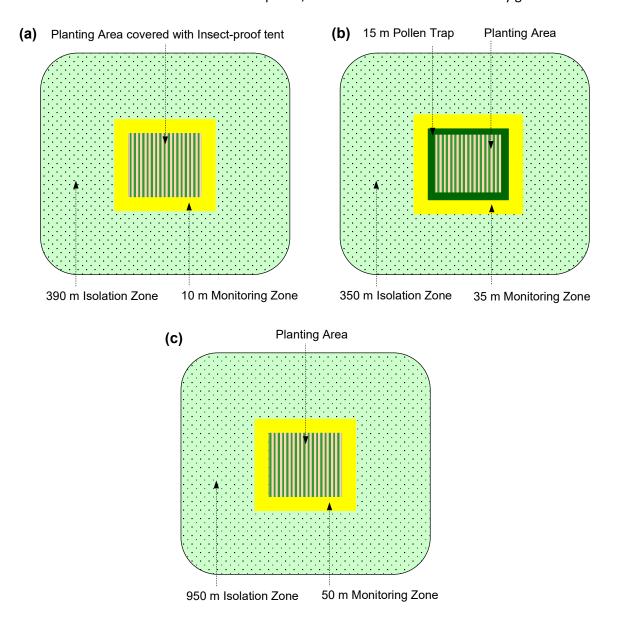


Figure 1. Diagrams (not to scale) showing the relationships between Planting Area, Pollen Trap, Monitoring Zone and Isolation Zone.

Site layout (a) with Insect-proof tent, (b) without Insect-proof tent and with Pollen Trap, and (c) without Insect-proof tent or Pollen Trap. Monitoring and Isolation Zones must be kept free of Related Species.

# Section 2 General conditions and obligations

- 3. This licence does not authorise dealings with the GMOs that are otherwise prohibited as a result of the operation of State legislation recognising an area as designated for the purpose of preserving the identity of GM crops, non-GM crops, or both GM crops and non-GM crops, for marketing purposes.
- 4. This licence remains in force until it is suspended, cancelled or surrendered. No dealings with the GMOs are authorised during any period of suspension.

Note: Although this licence has no expiry date, the period when GMOs may be grown is restricted in accordance with Condition 18.

- 5. The licence holder is Miruku Australia Pty Ltd.
- 6. The persons covered by this licence are the licence holder and employees, agents or contractors of the licence holder and other persons who are, or have been, engaged or otherwise authorised by the licence holder to undertake any activity in connection with the dealings authorised by this licence.
- 7. The GMOs with which dealings are authorised by this licence are those listed at Attachment A.
- 8. The dealings authorised by the licence are to:
  - (a) conduct experiments with the GMOs;
  - (b) make the GMOs;
  - (c) breed the GMOs;
  - (d) propagate the GMOs;
  - (e) use the GMOs in the course of manufacture of a thing that is not the GMOs;
  - (f) grow the GMOs;
  - (g) transport the GMOs;
  - (h) dispose of the GMOs;

and the possession, supply or use of the GMOs in the course of any of these dealings.

9. This licence does not apply to dealings with the GMOs conducted as a Notifiable Low Risk Dealing (NLRD) or pursuant to another authorisation under the Act.

Note: Dealings conducted as an NLRD must be assessed by an Institutional Biosafety Committee (IBC) before commencement and must comply with the requirements of the Regulations.

### General obligations of the licence holder

- 10. The licence holder must, at all times, remain an accredited organisation in accordance with the Act and must comply with its instrument of accreditation.
- 11. The licence holder must be able to access and control all Planting Areas, Pollen Traps, Monitoring Zones, Isolation Zones and approved facilities to the extent necessary to comply with this licence.
- 12. The licence holder must inform any person covered by this licence, to whom a particular condition of the licence applies, of the following:
  - (a) the particular condition, including any variations of it;
  - (b) the cancellation or suspension of the licence;
  - (c) the surrender of the licence.
- 13. The licence holder must not permit a person covered by this licence to conduct any dealing with the GMOs unless:
  - (a) the person has been informed of any applicable licence conditions, including any variation of them; and

- (b) the licence holder has obtained from the person a signed and dated statement that the person:
  - has been informed by the licence holder of the licence conditions including any variation of them; and
  - ii. has understood and agreed to be bound by the licence conditions, or variation.
- 14. The licence holder must inform the persons covered by this licence that any Personal Information relevant to the administration and/or enforcement of the licence may be disclosed to the Regulator.

#### General obligations of persons covered by the licence

15. If a person is authorised by this licence to deal with the GMOs and a particular condition of the licence applies to the dealing by the person, the person must allow the Regulator, or a person authorised by the Regulator, to enter premises where the dealing is being undertaken, for the purposes of auditing or monitoring the dealing.

Note: Under the Act, the definition of premises includes a building, area of land or vehicle.

### Section 3 Limits and control measures

#### 3.1 Limits on the release

The following licence conditions impose limits on where and when the GMOs may be grown.

- 16. The only plants that may be intentionally grown at a Planting Area are:
  - (a) the GMOs covered by this licence; and
  - (b) non-GM Canola; and
  - (c) plants approved in writing by the Regulator.
- 17. Non-GM Canola plants grown in a Planting Area must be handled as if they were the GMOs.
- 18. Planting and growing of the GMOs may only occur within the following limits:

#### Area and duration

Period	Maximum number of Planting Areas per year	Maximum size of any Planting Area
2025	2	0.5 ha
2026	5	1 ha
2027	10	2.5 ha
2028	15	5 ha
2029	20	5 ha

#### Local Government Areas in which Planting Areas may be located

New South Wales	Victoria	Western Australia	South Australia
Berrigan	Ararat	Albany	Adelaide Plains
Bland	Ballarat	Beverley	Barossa
Blayney	Benalla	Boddington	Light
Cabonne	Buloke	Boyup Brook	Wakefield
Coolamon	Campaspe	Bridgetown-Greenbushes	
Coonamble	Central Goldfields	Brookton	
Cootamundra-Gundagai	Colac Otway	Broomehill-Tambellup	
Cowra	Corangamite	Carnamah	
Dubbo	Gannawarra	Coorow	
Edward River	Glenelg	Corrigin	

New South Wales	Victoria	Western Australia	South Australia
Federation	Golden Plains	Cranbrook	
Forbes	Greater Bendigo	Cuballing	
Gilgandra	Greater Geelong	Cunderdin	
Greater Hume	Greater Shepparton	Dalwallinu	
Griffith	Hepburn	Denmark	
Gunnedah	Hindmarsh	Donnybrook-Balingup	
Gwydir	Horsham	Dowerin	
Hay	Indigo	Dumbleyung	
Hilltops	Loddon	Esperance	
Inverell	Macedon Ranges	Gnowangerup	
Junee	Mildura	Goomalling	
Leeton	Mitchell	Greater Geraldton	
Liverpool Plains	Moira	Jerramungup	
Lockhart	Moorabool	Katanning	
Mid-Western	Mount Alexander	Kent	
Moree Plains	Moyne	Kojonup	
Murray River	Northern Grampians	Manjimup	
Murrumbidgee	Pyrenees	Merredin	
Muswellbrook	Southern Grampians	Mingenew	
Narrabri	Strathbogie	Moora	
Narrandera	Swan Hill	Morawa	
Narromine	Wangaratta	Nannup	
Orange	West Wimmera	Narrogin	
Parkes	Wodonga	Northam	
Snowy Valleys	Wyndham	Perenjori	
Tamworth	Yarriambiack	Pingelly	
Temora		Plantagenet	
Upper Hunter		Quairading	
Wagga Wagga		Ravensthorpe	
Walgett		Tammim	
Warren		Three Springs	
Warrumbungle		Toodyay	
Weddin		Victoria Plains	
		Wagin	
		Wandering	
		West Arthur	
		Wickepin	
		Williams	
		Wongan-Ballidu	
		Woodanilling	
		Wyalkatchem	
		York	

#### 3.2 Control measures

The following licence conditions restrict the spread or persistence of the GMOs and their genetic material in the environment.

# Conditions to restrict exposure to the GMOs

- 19. Plant Material must not be used, sold or otherwise disposed of for any purpose which would involve or result in its use as food for humans other than that used in any sensory testing (see Condition 20) or feed for animals.
- 20. Products of the GMOs may be used in human sensory testing which may include taste testing, subject to those experiments being under oversight by a Human Research Ethics Committee, which is required

- to review and approve the research proposals in accordance with the National Statement on Ethical Conduct in Human Research.
- 21. The licence holder must not permit a person covered by this licence to conduct any dealing which may expose the person to Plant Material unless the person has provided a statement that the person does not have a known allergy to proteins expressed as a result of the modifications in the GM Canola.

Note: The licence holder should document compliance with this condition, as per Condition 13.

#### Conditions to restrict pollen flow

- 22. For each Planting Area, one of the following measures to limit gene flow must be adopted, either:
  - (a) cover every GMO planted in the planting area with Insect-proof tents from at least seven days prior to Flowering and until all GMOs have completed Flowering, and surround the Planting Area with a Monitoring Zone of at least 10 metres, and surround the Monitoring Zone with an Isolation Zone of at least 390 metres (as shown in Figure 1a); or
  - (b) surround the Planting Area with a Pollen Trap of at least 15 metres, and surround the Pollen Trap with a Monitoring Zone of at least 35 metres, and surround the Monitoring Zone with an Isolation Zone of at least 350 metres (as shown in Figure 1b); or
  - (c) surround the Planting Area with a Monitoring Zone of at least 50 metres, and surround the Monitoring Zone with an Isolation Zone of at least 950 metres (as shown in Figure 1c).
- 23. If a Pollen Trap is used in accordance with Condition 22, Pollen Trap Plants must:
  - (a) have a reasonably dense and vigorous growth; and
  - (b) be Flowering at the same time as the GMOs; and
  - (c) form a continuous barrier at least 15 metres wide around the Planting Area while the GMOs are Flowering, although one path of up to 3 metres in width is allowed in order to access the Planting Area; and
  - (d) be handled as if they were the GMOs.
- 24. The Monitoring Zone must be maintained in a manner appropriate to allow the identification and Destruction of Related Species while the GMOs are growing in the Planting Area.
  - Note: Measures to achieve this could include maintaining the area free of vegetation and/or keeping vegetation mown. Condition 48 requires details of current land use and recent land management practices to be recorded upon inspection of the Monitoring Zone.
- 25. The GMOs must not be planted in a Planting Area if any Related Species are being grown at the same time in the Monitoring or Isolation Zones.
  - Note: Refer to Condition 11 regarding access and control of areas.
- 26. While the GMOs are growing in a Planting Area, associated areas and Insect-proof tents must be inspected by people trained to recognise plants of Related Species, and actions must be taken as follows:

Area	Period of inspection	Inspection frequency	Inspect for	Action
Planting Area, Pollen Trap (if applicable) and Monitoring Zone	From 14 days prior to the expected commencement of Flowering of any GMOs* until all GMOs in the Planting Area have been harvested or Destroyed	At least once every 35 days	Related Species	Destroy before Flowering or prevent from Flowering
Insect-proof tents	While tents are in place	At least once every 14 days and after any Extreme Weather event	Damage that may render tents not Insect- proof	Repair any damage or replace if repair not possible
Isolation Zone	From 14 days prior to the expected commencement of Flowering of any GMOs* until all GMOs in the Planting Area have finished Flowering	At least once every 35 days	Intentionally planted Related Species	Destroy before Flowering or prevent from Flowering or Destroy the GMOs in the Planting Area

<sup>\*</sup>Condition 47(a) requires the licence holder to provide information to the Regulator on the expected Flowering period, however the inspection period should be based on the observed development of the GMOs, so that inspections commence prior to Flowering of any GMOs.

Note: Details of any inspection activity must be recorded in a Logbook (Condition 48) and reported to the Regulator (Condition 47).

#### Conditions to restrict seed dispersal

- 27. Equipment used in connection with the GMOs must be Cleaned as soon as practicable after use with the GMOs and before use for any other purpose.
- 28. Planting Areas and Pollen Traps must be at least 50 metres away from any permanent natural watercourses or man-made drainage features that flow into natural watercourses.

Note: This includes irrigation channels or storm water drains that flow into a natural watercourse.

- 29. Planting Areas and Pollen Traps must not be located in flood prone areas.
- 30. If the GMOs are windrowed, the licence holder must take, or have taken, measures to minimise the likelihood of dispersal of the GMOs by wind or rain. Appropriate measures may include:
  - (a) ensuring high density planting and growth of the GMOs prior to windrowing; or
  - (b) cutting/windrowing to allow maximum stubble height; or
  - (c) use of windrow roller; or
  - (d) appropriate site selection.

Note: Appropriate site selection includes avoidance of windy areas. Windrowing dates and details of measures used to minimise dispersal of GMOs must be reported to the Regulator (Condition 47(d)).

#### Conditions relating to harvesting

- 31. All GMOs planted within a Planting Area must be harvested or Destroyed within 8 months after the first planting of any GMO within that Planting Area.
- 32. If all GMOs in a Planting Area have been Destroyed, then for the purposes of this licence:
  - (a) the GMOs are taken to have been harvested; and
  - (b) the Planting Area is taken to have been Cleaned.

Note: Cleaning activities must be reported to the Regulator (Condition 47). Areas of land that have been Cleaned are subject to inspections (Condition 36).

- 33. The GMOs must be harvested and threshed separately from any other crop.
- 34. Harvested GM seed not required for experimentation or future planting must be Destroyed as soon as practicable.

### Conditions to restrict persistence of GMOs on trial sites

35. Areas of land used in connection with the GMOs must be Cleaned as follows:

Area	s of land to be Cleaned	When
i.	Planting Area,	Within 14 days after harvest of the GMOs
ii.	Pollen Trap, if used, and	
iii.	10 metres around each Planting Area, or around the Pollen Trap, if used (innermost 10 metres of Monitoring Zone)	
-	other area used to Clean any Equipment used in nection with the GMOs	As soon as practicable
	other area where the GMOs have dispersed, e.g. ng planting, growing, harvesting or Destruction	As soon as practicable

Note: Cleaning activities must be reported to the Regulator (Condition 47). Areas of land that have been Cleaned are subject to inspections (Condition 36).

36. After Cleaning, areas of land must be inspected by people trained to recognise Canola. Inspections must cover the entirety of areas to be inspected. Actions must be taken as follows:

Area	Period of inspection	Inspection frequency	Inspect for	Action
Planting Area, Pollen Trap, innermost 10 metres of Monitoring Zone and other areas of land that were Cleaned in accordance with Condition 35.	From the day of Cleaning, until:  i. the area is planted as a new Planting Area in accordance with Condition 16 and inspections required under Condition 26; or  ii. the Regulator has issued a Sign off for the area	At least once every 35 days	Volunteers	Destroy before Flowering

Note: Details of any inspection activity must be recorded in a Logbook (Condition 48) and reported to the Regulator (Condition 47).

- 37. While post-Cleaning inspection requirements apply to an area:
  - (a) the area must be Tilled within 60 days of harvest of the GMOs at a Planting Area, unless otherwise approved in writing by the Regulator; and

Note: If Tillage is used as a method of Cleaning, the Tillage done as Cleaning also meets the requirements for a Tillage within 60 days of harvest.

- (b) within the 12 months prior to submission of a Sign off application, the area must be Tilled and then receive a watering event as described in **Attachment B**; and
- (c) the area must be maintained in a manner appropriate to allow identification of Volunteers; and
- (d) the area must not be used for grazing livestock; and

- (e) no plants may be intentionally grown in the area unless:
  - i. the area is planted as a new Planting Area in accordance with Condition 16; or
  - ii. the plants are listed as post-harvest crops permitted for GM Brassica field trial sites in the OGTR Policy on Post Harvest Crops as current at the time of planting; or
  - iii. the plants are agreed to in writing by the Regulator.

Note: The OGTR's Policy on Post Harvest Crops can be found on the OGTR website.

#### Tillage

38. Any Tillage of the Planting Area and the Pollen Trap must be to a depth no greater than 5 centimetres.

#### **Destruction by burial**

- 39. If Destruction of GMOs occurs by burial:
  - (a) the GMOs must be buried in a pit and covered by a layer of soil at least one metre in depth, the top of which is no higher than the surrounding soil surface; and
  - (b) seeds must be wet when buried to encourage decomposition; and
  - (c) the licence holder must take measures to ensure that the burial site is not disturbed for a period of at least two years from the date of burial.

Note: If GMOs are dispersed on the soil surface during the process of burial, the burial site becomes an area of land that requires Cleaning under Condition 35, and is subject to post-Cleaning requirements.

Note: The date and location of burial, and measures used to ensure that the burial site is not disturbed, must be reported to the Regulator (Condition 47(g)).

#### Processing or experimentation with the GMOs

- 40. Treatment, threshing or processing of GM seed or experimentation or analysis with the GMOs may only be undertaken within:
  - (a) a Planting Area before Cleaning; or
  - (b) a Pollen Trap before Cleaning; or
  - (c) the innermost 10 m of a Monitoring Zone before Cleaning; or
  - (d) a facility approved in writing by the Regulator.

Note: This condition does not apply to dealings conducted as an NLRD (see Condition 9).

41. Within a facility approved in writing by the Regulator in accordance with Condition 40, any area that is used for treatment, threshing, processing, experimentation or analysis of the GMOs must be Cleaned as soon as practicable and before use for any other purpose.

#### Transport or storage of the GMOs

- 42. Transport or storage of the GMOs must:
  - (a) only occur to the extent necessary to conduct the dealings permitted by this licence or other valid authorisation under the Act; and
  - (b) be in accordance with the Regulator's *Guidelines for the Transport, Storage and Disposal of GMOs* for PC2 GM plants as current at the time of transportation or storage; and
  - (c) comply with all other conditions of this licence.

Note: Activities with the GMOs within a Planting Area prior to Cleaning are not regarded as transport or storage.

Note: Condition 13 requires signed statements for persons transporting the GMOs.

Note: This condition does not apply to dealings conducted as an NLRD (see Condition 9).

43. Methods and procedures used to transport GMOs must be recorded, and must be provided to the Regulator, if requested.

Note: The Contingency Plan must be implemented if the GMOs are detected outside areas under inspection (Condition 44).

#### Contingency plan

44. If any unintentional presence of the GMOs is detected outside the areas requiring Cleaning, the Contingency Plan must be implemented.

# Section 4 Sign off

- 45. The licence holder may make written application to the Regulator that planting restrictions and inspection requirements no longer apply to the Planting Area and other areas requiring Cleaning if:
  - (a) post-Cleaning inspection activities have been conducted for at least 24 months on the area; and
  - (b) conditions have been conducive for germination and detection of Volunteers; and
  - (c) no Volunteers have been detected in the area during the 12 months prior to the Sign off request.

Note: The licence requires two Tillages and a watering event prior to a Sign off application (Condition 37).

Note: The Regulator will take into account the management and inspection history for the Planting Area and other areas requiring Cleaning, including post-harvest crops planted (if any), Tillage, irrigation, rainfall, application of herbicide and occurrence of Volunteers, in deciding whether or not further inspections are required to manage persistence of the GMOs.

# Section 5 Reporting and documentation

The following licence conditions are imposed to demonstrate compliance with other conditions and facilitate monitoring of compliance by staff of the OGTR.

46. General notifications must be sent to the Regulator as follows:

Note: please send all correspondence related to the licence to <u>OGTR.M&C@health.gov.au</u>.

	Notice	Content of notice	Timeframe
a.	Changes to contact details	Changes to any of the contact details of the project supervisor that were notified in the licence application or subsequently	As soon as practicable
b.	Ongoing suitability to hold a licence	<ul> <li>i. any relevant conviction of the licence holder; or</li> <li>ii. any revocation or suspension of a licence or permit held by the licence holder under a law of the Australian Government, a State or a foreign country, being a law relating to the health and safety of people or the environment; or</li> <li>iii. any event or circumstances that would affect the capacity of the licence holder to meet the conditions of the licence; and</li> </ul>	As soon as practicable after any of these events occur
		<ul> <li>iv. any information related to the licence holder's ongoing suitability to hold a licence, that is requested by the Regulator</li> </ul>	Within the timeframe stipulated by the Regulator

C.	People covered by the licence	<ul> <li>i. names of all organisations and persons, or functions or positions of the persons, who will be covered by the licence, with a description of their responsibilities; and Note: Examples of functions or positions are 'project supervisor', 'site manager', 'farm labourer' etc.</li> <li>ii. detail of how the persons covered by the licence will be informed of licence conditions</li> </ul>	At least 14 days prior to conducting any dealings with the GMOs (to be updated within 14 days if the notified details change)
d.	Testing methodology	A written methodology to reliably detect the genetic modifications described in this licence. The detection method/s must be capable of identifying each GM Canola line planted under this licence	At least 14 days prior to conducting any dealings with the GMOs (to be updated within 14 days if the notified details change)
e.	Contingency plan	A Contingency Plan to respond to inadvertent presence of the GMOs outside an area that must be inspected	At least 14 days prior to conducting any dealings with the GMOs (to be updated within 14 days if the notified details change)
f.	Training records	Copies of the signed and dated statements referred to in Condition 13 if requested by the Regulator	Within the timeframe stipulated by the Regulator
g.	Additional information required by the Act	<ul> <li>i. additional information as to any risks to the health and safety of people, or to the environment, associated with the dealings authorised by the licence; or</li> <li>ii. any contraventions of the licence by a person covered by the licence; or</li> <li>iii. any unintended effects of the dealings authorised by the licence</li> <li>Note: The Act requires, for the purposes of the Condition 46(g), that:</li> <li>the licence holder will be taken to have become aware of additional information of a kind mentioned in Condition 46(g) if he or she was reckless as to whether such information existed; and</li> <li>the licence holder will be taken to have become aware of contraventions, or unintended effects, of a kind mentioned in Condition 46(g), if he or she was reckless as to whether such contraventions had occurred, or such unintended effects existed</li> <li>Note: Contraventions of the licence may occur through the action or inaction of a person.</li> </ul>	Without delay after becoming aware of any new information  Note: An example of notification without delay is contact made within a day of a contravention of the licence via the OGTR free call phone number 1800 181 030.  Notification without delay will allow the OGTR to conduct a risk assessment on the incident and attend the location, if required
h.	Further details regarding additional information	Any further details requested by the Regulator in relation to information provided under Condition 46(g)	Within the timeframe stipulated by the Regulator

# 47. Notifications relating to each trial site must be sent to the Regulator as follows:

Note: please send all correspondence related to the licence to <u>OGTR.M&C@health.gov.au</u>.

Notice	Content of notice	Timeframe
a. Intention to plant	<ul> <li>Details of the Planting Area including size, the local government area, GPS coordinates, a street address, a diagrammatical representation of the trial site (e.g. Google Maps) and any other descriptions</li> </ul>	At least 7 days prior to each planting (to be updated as soon as practicable if the notified details change)

	Notice	Content of notice	Timeframe
		<ul> <li>ii. Whether an Insect-proof tent or Pollen Trap will be used</li> <li>iii. Detail of how the licence holder will access and control the Planting Area and the associated Pollen Trap, Monitoring Zone and Isolation Zone, in accordance with Condition 11</li> </ul>	
		Note: this should include a description of any contracts, agreements, or other enforceable arrangements.	
		iv. Identity of the GMOs to be planted at the Planting Area (e.g. inserted vector name)	
		v. Date on which the GMOs will be planted	
		vi. Period when the GMOs are expected to Flower	
		vii. Period when windrowing (if intended) is expected to commence	
		viii. Period when harvesting is expected to commence	
		ix. How all areas requiring post-Cleaning inspections are intended to be used until Sign off, including proposed post-harvest crops (if any)	
		x. Details of how inspection activities will be managed, including strategies for the detection and Destruction of Volunteers and Related Species	
		xi. History of how the trial site has been used for the previous two years	
b.	Planting	<ul><li>i. Actual date(s) of planting the GMOs</li><li>ii. Any changes to the details provided under part (a) of this condition</li></ul>	Within 7 days of any planting
c.	Extreme Weather	Any Extreme Weather event that is expected to affect or has already affected an area where the GMOs are or may be present.	As soon as practicable
		Note: The Contingency Plan must be implemented if the GMOs are detected outside areas requiring Cleaning (Condition 44).	
d.	Windrowing	Actual date(s) of windrowing and details of measures used to minimise dispersal of the GMOs during windrowing (Condition 30).	Within 7 days of commencement of windrowing
e.	Harvest	Actual date(s) of harvesting the GMOs	Within 7 days of commencement of any harvesting
f.	Cleaning	Date(s) on which required Cleaning was performed on any areas of land	Within 7 days of completion of Cleaning
		ii. Method(s) of Cleaning	
g.	Destruction by burial	Date of burial, location of burial including GPS co-ordinates, and details of measures used to ensure that the burial site will not be disturbed for the period required by Condition 39	Within 7 days of burial of any GMOs
h.	Inspection activities	Information recorded in a Logbook as per the inspection requirements (Conditions 26, 36 and 48).	Within 35 days of inspection

Note: Additional records must be provided to the Regulator, if requested, in accordance with Condition 43.

- 48. Details of any inspection activity must be recorded in a Logbook and must include:
  - (a) date of the inspections; and
  - (b) name of the person(s) conducting the inspections; and
  - (c) details of the experience, training or qualification that enables the person(s) to recognise Canola and/or Related Species, if not already recorded in the Logbook; and
  - (d) details of areas inspected including current land use (including any post-harvest crops) and recent management practices applied; and

Note: management practices include Tillage events, spraying or maintenance measures used to facilitate inspections.

- (e) details of the developmental stage of the GMOs while they are being grown; and
- (f) details of any post-Cleaning rainfall events including measurements at or near the area, or any irrigation events; and
- (g) details of any Volunteers and/or Related Species observed during inspections or during land-management activities, including number, developmental stage and approximate position of the Volunteers and/or Related Species within each area inspected<sup>†</sup>; and
- (h) date(s) and method(s) of Destruction of or preventing Flowering of any Volunteers and/or Related Species, including destruction of Volunteers and/or Related Species during land-management activities; and
- (i) details of any damage and any repairs to the Insect-proof tents, while Insect-proof tents are required.

- descriptive text
- marking on a diagram
- indicating grid references on a corresponding map/sketch.

Note: Details of inspection activities must be provided to the Regulator (Condition 47). The Regulator has developed a standardised proforma for recording inspection activities. This can be made available on request.

<sup>&</sup>lt;sup>†</sup> Examples of acceptable ways to record the positional information for Volunteers and/or Related Species in the Logbook include:

#### **ATTACHMENT A**

**DIR No: 215** 

Full Title: Limited and controlled release of Canola genetically modified for diary protein

production

**Organisation Details** 

Postal address: PO Box 3555

South Brisbane

QLD 4101 Australia

**IBC Details** 

IBC Name: Miruku Australia

## **GMO Description**

### **GMOs** covered by this licence

Canola plants genetically modified by introduction of only the genes and genetic elements listed below.

#### **Parent Organism**

Common Name: Canola

Scientific Name: Brassica napus L.

#### **Modified traits**

Category: Composition – food (processing)

Selectable marker - herbicide

Description: Canola plants have been genetically modified for production of a chimeric dairy

protein and include a herbicide tolerance gene. The introduced genes are listed in

Table 1 and the associated genetic elements are listed in Table 2.

Table 1. Introduced genes in the GM Canola

Genetic element	Source organism	Encoded protein	Intended function
bar	Streptomyces hygroscopicus	Phosphinothricin acetyltransferase	Herbicide resistance
BCN2	Bos taurus	Miruku β-casein (CSN2) variant 2	β-casein production
BCN3	B. taurus	Miruku β-casein (CSN2) variant 3	β-casein production
CCI gene*	CCI*	CCI*	CCI*
CCI gene*	CCI*	CCI*	CCI*

<sup>\*</sup>This information has been declared Confidential Commercial Information (CCI) under s185 of the Act

Table 2. Introduced regulatory sequences in the GM Canola

Genetic element	Source	Intended function
AtCel1 cell wall signal	Cell wall signal from Arabidopsis thaliana CEL1 gene	Cell wall targeting sequence
CaMV35S promoter	Cauliflower mosaic virus	Constitutive promoter
CCI 3'UTR*	Glycine max	Terminator and polyadenylation signal
CCI promoter*	G. max	Seed-specific promoter
CT-CVS C- terminus	C-terminal beta-conglycinin vacuolar sequence from Glycine max	Terminator
ER Retention	C-terminal H/KDEL tag sequence from A. thaliana	Endoplasmic reticulum retention
GmCVS vacuolar signal	Vacuole signal from <i>G. max</i> beta-conglycinin gene	Vacuole targeting sequence
HvAVSP vacuolar signal	Vacuole signal from Hordeum vulgare aleurain gene	Vacuole targeting sequence
Prr	Nicotiana tabacum	Cell wall targeting sequence
nos 3'UTR	Agrobacterium tumefaciens	Terminator and polyadenylation signal

<sup>\*</sup>This information has been declared Confidential Commercial Information (CCI) under s185 of the Act

#### ATTACHMENT B

A watering event is irrigation or natural rainfall that provides sufficient soil moisture to promote germination of Canola seeds on a trial site.

Examples of acceptable watering events are:

- At least 26 millimetres of rainfall over one day; or
- At least 28 millimetres of rainfall over two days; or
- At least 30 millimetres of rainfall over three days; or
- At least 32 millimetres of rainfall over four days; or
- Irrigation that provides equivalent levels of soil moisture to one of the examples of rainfall above.

Rainfall measurements must be taken on the site or within 3 km of the site. An irrigation or natural rainfall that matches one of the examples listed above, and occurs during the time period specified for a watering event in Condition 37 of the licence, is considered a valid watering event. The licence holder should keep records of the date/s and amount of water applied during the watering event, and provide this information when requesting Sign off of the relevant site.

If an irrigation or natural rainfall does not match one of the examples listed above, the licence holder may submit a request to the Regulator for it to be considered a watering event. The request should provide:

- evidence of amount of water applied, such as rainfall measurements on the site or within 3 km of the site, and
- evidence that resultant soil moisture is suitable for germination, such as photos of germinating plants on the site.

It is recommended that any requests that an irrigation or natural rainfall be considered a watering event be submitted at the time of the event, to minimise potential delays to Sign off of the site.

# References

ABARES (2024a). Australian Crop Report, No. 210. (Canberra: Australian Bureau of Agricultural and Resource Economics and Sciences).

ABARES (2024b). Australian Crop Report, No. 212. (Canberra: Australian Bureau of Agricultural and Resource Economics and Sciences).

Alvarez, M.J., Estrada, J.L., Gozalo, F., Fernandez-Rojo, F., and Barber, D. (2001). Oilseed rape flour: another allergen causing occupational asthma among farmers. Allergy *56*, 185-188.

ANZFA (2001). Final risk analysis report - Application A372: Oil derived from glufosinate-ammonium tolerant canola lines Topas 19/2 and T45 and oil derived from glufosinate-ammonium tolerant and pollination controlled lines MS1, MS8, RF2 and RF3. (Canberra, Australia: Australia New Zealand Food Authority).

AOF (2019). Canola volunteer control 2019. (New South Wales, Australia: Australian Oilseeds Federation).

Arts, J.H.E., Mommers, C., and de Heer, C. (2006). Dose-response relationships and threshold levels in skin and respiratory allergy. Critical Reviews in Toxicology *36*, 219-251.

Baker, J., and Preston, C. (2008). Canola (*Brassica napus* L.) seedbank declines rapidly in farmer-managed fields in South Australia. Australian Journal of Agricultural Research *59*, 780-784.

Belter, A. (2016). Long-term monitoring of field trial sites with genetically modified oilseed rape (*Brassica napus* L.) in saxony-Anhalt, Germany. Fifteen years persistence to date but no spatial dispersion. Genes 7, 1-13.

Bonadonna, P., Senna, G., and Passalacqua, G. (2003). Dermatological powder as hidden cause of occupational allergy due to casein: a case report. Occup Environ Med *60*, 609-610.

Bos, J.D., and Meinardi, M.M. (2000). The 500 Dalton rule for the skin penetration of chemical compounds and drugs. Exp Dermatol *9*, 165-169.

Boyd, E.M., Krijnen, C.J., and Peters, J.M. (1967). Lethal amounts of casein, casein salts and hydrolyzed casein given orally to albino rats. J Nutr *93*, 429-437.

Brown, S. (2021). 2022 Victorian crop sowing guide. (Grains Research & Development Corporation, Agriculture Victoria).

Busi, R., and Powles, S.B. (2016). Transgenic glyphosate-resistant canola (*Brassica napus*) can persist outside agricultural fields in Australia. Agriculture, Ecosystems & Environment *220*, 28-34.

Cassiano, M.M., and Areas, J.A. (2003). Dependence of the interfacial behavior of beta-casein on phosphoserine residues. J Dairy Sci *86*, 3876-3880.

Creamer, L.K., Richardson, T., and Parry, D.A. (1981). Secondary structure of bovine alpha s1- and beta-casein in solution. Arch Biochem Biophys *211*, 689-696.

CSGA (2022). Canadian Regulations and Procedures for Pedigreed Seed Crop Production. (Canadian Seed Growers' Association).

Daniloski, D., McCarthy, N.A., Huppertz, T., and Vasiljevic, T. (2022). What is the impact of amino acid mutations in the primary structure of caseins on the composition and functionality of milk and dairy products? Curr Res Food Sci *5*, 1701-1712.

Darmency, H., and Fleury, A. (2000). Mating system in *Hirschfeldia incana* and hybridisation to oilseed rape. Weed Research *40*, 231-238.

Darmency, H., Lefol, E., and Fleury, A. (1998). Spontaneous hybridisations between oilseed rape and wild radish. Molecular Ecology 7, 1467-1473.

Dauphas, S., Mouhousriou, N., Metro, B., Mackie, A., Wilde, P., Anton, M., and Riaublanc, A. (2005). The supramolecular organisation of ?-casein: effect on interfacial properties. Food Hydrocolloids *19*, 387-393.

Delaney, B., Goodman, R.E., and Ladics, G.S. (2018). Food and feed safety of genetically engineered food crops. Toxicological Sciences *162*, 361-371.

Docena, G.H., Fernandez, R., Chirdo, F.G., and Fossati, C.A. (1996). Identification of casein as the major allergenic and antigenic protein of cow's milk. Allergy *51*, 412-416.

Farrell, H.M., Jr., Jimenez-Flores, R., Bleck, G.T., Brown, E.M., Butler, J.E., Creamer, L.K., Hicks, C.L., et al. (2004). Nomenclature of the proteins of cows' milk--sixth revision. J Dairy Sci 87, 1641-1674.

Felsot, A.S. (2000). Insecticidal genes part 2: Human health hoopla. Agrichemical & Environmental News 168, 1-7.

Fiocchi, A., Schunemann, H.J., Brozek, J., Restani, P., Beyer, K., Troncone, R., Martelli, A., et al. (2010). Diagnosis and Rationale for Action Against Cow's Milk Allergy (DRACMA): a summary report. J Allergy Clin Immunol 126, 1119-1128 e1112.

Ford, C.S., Allainguillaume, J., Grilli-Chantler, P., Cuccato, G., Allender, C.J., and Wilkinson, M.J. (2006). Spontaneous gene flow from rapeseed (*Brassica napus*) to wild *Brassica oleracea*. Proceedings of the Royal Society B: Biological Sciences *273*, 3111-3115.

FSANZ (2005a). Final assessment report- Application A533: Food derived from glufosinate ammonium-tolerant cotton line LL25. (Canberra, Australia: Food Standards Australia New Zealand).

FSANZ (2005b). Final assessment report - Application A543: Food derived from Insect-protected, glufosinate ammonium-tolerant corn line 59122-7. (Canberra, Australia: Food Standards Australia New Zealand).

FSANZ (2008). Final assessment report - Application A589: Food derived from glufosinate ammonium tolerant rice line LLRICE62. (Canberra, Australia: Food Standards Australia New Zealand).

FSANZ (2010a). Application A1028: Food derived from insect-protected & herbicide-tolerant cotton line T304-40 - Approval report. (Canberra, Australia: Food Standards Australia New Zealand ).

FSANZ (2010b). Application A1040: Food derived from insect-protected and herbicide-tolerant cotton line GHB119 - Approval report. (Canberra, Australia: Food Standards Australia New Zealand).

FSANZ (2013). Approval report - Application A1080. Food derived from herbicide-tolerant cotton line MON 88701. (Canberra, Australia: Food Standards Australia New Zealand).

FSANZ (2017). A1140 – Food derived from Herbicide-tolerant Canola Line MS11: Supporting document 1 - Safety Assessment (at Approval). (Canberra, Australia: Food Standards Australia New Zealand).

Giribaldi, M., Lamberti, C., Cirrincione, S., Giuffrida, M.G., and Cavallarin, L. (2022). A2 Milk and BCM-7 Peptide as Emerging Parameters of Milk Quality. Front Nutr *9*, 842375.

GRDC (2009). Canola best practice management guide for south-eastern Australia. (Canberra, Australia: Grains Research & Development Corporation).

GRDC (2011). Break crop benefits: fact sheet. (Canberra, Australia: Grains Research & Development Corporation).

GRDC (2015). GRDC GrowNotes Canola - Southern Region. (Canberra, Australia: Grains Research & Development Corporation).

GRDC (2017). GRDC Canola GrowNotes: Northern. (Grains Research and Development Corporation).

Gulden, R.H., Shirtliffe, S.J., and Thomas, A.G. (2003). Secondary seed dormancy prolongs persistence of volunteer canola in western Canada. Weed Science *51*, 904-913.

Hassanin, A.A., Osman, A., Atallah, O.O., El-Saadony, M.T., Abdelnour, S.A., Taha, H.S.A., Awad, M.F., et al. (2022). Phylogenetic comparative analysis: Chemical and biological features of caseins (alpha-S-1, alpha-S-2, beta- and kappa-) in domestic dairy animals. Front Vet Sci 9, 952319.

Hüsken, A., and Dietz-Pfeilstetter, A. (2007). Pollen-mediated intraspecific gene flow from herbicide resistant oilseed rape (*Brassica napus* L.). Transgenic Research *16*, 557-569.

Jensen, S.A., Fiocchi, A., Baars, T., Jordakieva, G., Nowak-Wegrzyn, A., Pali-Scholl, I., Passanisi, S., et al. (2022). Diagnosis and Rationale for Action against Cow's Milk Allergy (DRACMA) Guidelines update - III - Cow's milk allergens and mechanisms triggering immune activation. World Allergy Organ J 15, 100668.

Kay, R., Chan, A., Daly, M., and McPherson, J. (1987). Duplication of CaMV 35S promoter sequences creates a strong enhancer for plant genes. Science *236*, 1299-1302.

Keese, P. (2008). Risks from GMOs due to horizontal gene transfer. Environmental Biosafety Research 7, 123-149.

Keese, P.K., Robold, A.V., Myers, R.C., Weisman, S., and Smith, J. (2014). Applying a weed risk assessment approach to GM crops. Transgenic Research *23*, 957-969.

Masarwi, M., Gabet, Y., Dolkart, O., Brosh, T., Shamir, R., Phillip, M., and Gat-Yablonski, G. (2016). Skeletal effect of casein and whey protein intake during catch-up growth in young male Sprague-Dawley rats. Br J Nutr *116*, 59-69.

Matthews, P., McCaffery, D., and Jenkins, L. (2021). Winter crop variety sowing guide 2021. (NSW Department of Primary Industries).

McCarthy, N.A., Kelly, A.L., O'Mahony, J.A., and Fenelon, M.A. (2013). The physical characteristics and emulsification properties of partially dephosphorylated bovine beta-casein. Food Chem *138*, 1304-1311.

Moore, G., and Nazeri, N. (2022). Environmental weed risk assessment: Canola - oilseed rape (*Brassica napus*). (Western Australia: Department of Primary Industries and Regional Development).

Nakonechna, A., Matthews, D., and Sargur, R. (2019). Occupational asthma, rhinitis, contact dermatitis, and severe milk allergy caused by primary occupational exposure to casein. Ann Allergy Asthma Immunol *123*, 224-225.

OECD (2002). Series on Harmonization of Regulatory Oversight in Biotechnology, No 25. Module II: Phosphinothricin. Report No. ENV/JM/MONO(2002)14. (Organisation for Economic Cooperation and Development).

OGTR (2013). Risk Analysis Framework 2013, 4th edn (Canberra: Office of the Gene Technology Regulator).

OGTR (2024). The Biology of *Brassica napus* L. (canola) and *Brassica juncea* (L.) Czern. & Coss. (Indian mustard) (Canberra, Australia: Office of the Gene Technology Regulator).

Philips, J.G., Martin-Avila, E., and Robold, A.V. (2022). Horizontal gene transfer from genetically modified plants - Regulatory considerations. Front Bioeng Biotechnol *10*, 971402.

Pivard, S., Adamczyk, K., Lecomte, J., Lavigne, C., Bouvier, A., Deville, A., Gouyon, P.H., et al. (2008). Where do the feral oilseed rape populations come from? A large-scale study of their possible origin in a farmland area. Journal of Applied Ecology 45, 476-485.

Poikonen, S., Puumalainen, T.J., Kautiainen, H., Burri, P., Palosuo, T., Reunala, T., and Turjanmaa, K. (2006). Turnip rape and oilseed rape are new potential food allergens in children with atopic dermatitis. Allergy *61*, 124-127.

Randall, R.P. (2017). A global compendium of weeds, 3rd edn (Perth, Western Australia: R. P. Randall).

Rieger, M.A., Lamond, M., Preston, C., Powles, S.B., and Roush, R. (2002). Pollen-mediated movement of herbicide resistance between commercial canola fields. Science *296*, 2386-2388.

Salisbury, P.A. (2002). Genetically modified canola in Australia: agronomic and environmental considerations (Australian Oilseed Federation, Melbourne, Australia).

Schnell, J., Steele, M., Bean, J., Neuspiel, M., Girard, C., Dormann, N., Pearson, C., et al. (2015). A comparative analysis of insertional effects in genetically engineered plants: considerations for pre-market assessments. Transgenic Research 24, 1-17.

Sebastiani, C., Arcangeli, C., Ciullo, M., Torricelli, M., Cinti, G., Fisichella, S., and Biagetti, M. (2020). Frequencies Evaluation of beta-Casein Gene Polymorphisms in Dairy Cows Reared in Central Italy. Animals (Basel) *10*.

Shimada, K., Matsui, I., Inoue, K., Matsumoto, A., Yasuda, S., Katsuma, Y., Sakaguchi, Y., *et al.* (2020). Dietary casein, egg albumin, and branched-chain amino acids attenuate phosphate-induced renal tubulointerstitial injury in rats. Sci Rep *10*, 19038.

Shoormasti, R.S., Pourpak, Z., Yazdanyar, Z., Lebaschi, Z., Teymourpour, P., Barzegar, S., Tazesh, B., et al. (2011). The most common cow's milk allergenic proteins regarding to allergic symptoms. Clinical and Translational Allergy 1.

Soriano, V.X., Lee, H.Y., Dharmage, S.C., Perrett, K.P., Peters, R.L., and Koplin, J.J. (2023). Prevalence and risk factors of cow's milk sensitization and allergy in southeast Australia. J Allergy Clin Immunol Pract *11*, 3541-3543 e3541.

Steiner, H.Y., Halpin, C., Jez, J.M., Kough, J., Parrott, W., Underhill, L., Weber, N., et al. (2013). Evaluating the potential for adverse interactions within genetically engineered breeding stacks. Plant Physiology 161, 1587-1594.

Thompson, C.J., Movva, N.R., Tizard, R., Crameri, R., Davies, J., Lauwereys, M., and Botterman, J. (1987). Characterization of the herbicide-resistance gene *bar* from *Streptomyces hygroscopicus*. EMBO Journal *6*, 2519-2523.

Toden, S., Bird, A.R., Topping, D.L., and Conlon, M.A. (2005). Resistant starch attenuates colonic DNA damage induced by higher dietary protein in rats. Nutr Cancer *51*, 45-51.

Twigg, L.E., Taylor, C.M., Lowe, T.J., and Calver, M.C. (2008). Can seed-eating birds spread viable canola seed? Pacific Conservation Biology *14*, 119-127.

Vanhatalo, A., Varvikko, T., and Huhtanen, P. (2003). Effects of casein and glucose on responses of cows fed diets based on restrictively fermented grass silage. J Dairy Sci 86, 3260-3270.

Virtue, J.G. (2008). SA weed risk management guide. (Adelaide: Government of South Australia: Department of Water, Land and Biodiversity Conservation).

Warwick, S.I., Francis, A., and Gugel, R.K. (2009). Guide to Wild Germplasm of Brassica and Allied Crops (tribe Brassicaeae, Brassicaceae) 3rd Edition (Agriculture and Agri-Food Canada).

White, M., Cheal, D., Carr, G.W., Adair, R., Blood, K., Muir, A., and Meagher, D. (2022). Advisory list of environmental weeds in Victoria 2022. (Heidelberg, Victoria: Department of Environment, Land, Water and Planning).

Woodgate, J.L., Steadman, K.J., and Buchanan, K.L. (2011). A study of seed viability following consumption by birds. (Unpublished final report submitted to the OGTR).

Yoshimura, Y., Beckie, H.J., and Matsuo, K. (2006). Transgenic oilseed rape along transportation routes and port of Vancouver in western Canada. Environmental and Biosafety Research *5*, 67-75.

Zhang, Y., Singh, M.B., Swoboda, I., and Bhalla, P.L. (2005). Agrobacterium-mediated transformation and generation of male sterile lines of Australian canola. Australian Journal of Agricultural Research *56*.